

# **Recent Worldwide Research on Animal Pox Viruses**

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### **Executive Summary**

Although smallpox disease has been eliminated worldwide, concerns remain regarding variola virus (the cause of smallpox) and related poxviruses. It is widely feared that samples of variola virus may have been retained in several countries after a WHO directive to allow storage of such samples only in Russia and the United States. Even if the US and Russian stocks of variola virus were now destroyed, there would still be concern over possible hidden collections. A further worry is that some of the stocks of variola virus may have found their way into the hands of terrorists or rogue nations. It is even possible that the virus could be obtained from sites of human burials and grown up in a laboratory. These concerns are compounded by the suspension of vaccination against smallpox over the past three decades – with the eradication of smallpox as a naturally occurring disease – resulting in a population that lacks adequate immunity to the deadly virus.

With advances in molecular genetics technologies as well as in understanding of the disease-causing mechanisms of poxviruses, it is now possible to genetically engineer relatively benign poxviruses to make them more virulent. As a result, several available poxviruses already able to infect humans (monkeypox, buffalopox, cowpox, camelpox, and the vaccinia-like viruses circulating in the wild in Brazil) could potentially become serious human pathogens through evolutionary changes or laboratory manipulations, and perhaps become as worrisome as historical smallpox disease. Current vaccines will probably protect against the naturally occurring viruses as they evolve, but they might not protect against certain genetically engineered viruses.

The possibility that a poxvirus could be made more virulent for humans increased in likelihood when scientists in Australia found that a soluble factor of the immune system (called IL4) markedly worsened a poxvirus disease in mice. In that case, incorporation of the mouse IL4 gene into the mousepox virus genome led to fatal disease even in mice that normally had a genetic resistance to mousepox. Of great concern, the mousepox virus carrying the IL4 gene induced fatal disease in mice despite prior standard immunization against mousepox virus (immunization that protected mice from an unmanipulated mousepox virus). Therefore, it is possible that genetic modification of a poxvirus that causes mild human disease (vaccinia, monkeypox, cowpox, camelpox) might produce an engineered poxvirus that is as lethal as variola virus. Moreover, it is feared that such an engineered virus might cause fatal disease even in immunized people.

Apart from the Australian experiment, other countries have bioengineered Orthopoxviruses (vaccinia, monkeypox, cowpox, camelpox) by introducing different gene coding for different cytokines. In France, for example, scientists introduced IL2 genes into a vaccinia virus in order to study treatment of cancers. In Spain, researchers bioengineered the same virus with genes that produce two cytokines: IL8 and IL12. Japanese scientists did the same experiment using genes encoding for IL12, IL17 and IL23. The Chinese, on the other hand, separately engineered a human interleukin gene

and an interferon gene into an Othopoxvirus. Such experimentation could potentially lead to the creation of a super poxvirus with enhanced pathogenecity and increased lethality. In the wrong hands such an "enhanced" virus could readily be used as a BW agent.

The intensive Orthovirus genetics program that existed under the Soviet regime gives rise to concerns that advanced technology could transfer from Russia to countries harboring anti-US sentiments, third-world countries, terrorist organizations and/or non-state actors. Of utmost concern are Russian scientists with experience in Orthovirus bioengineering who have moved to other countries and continue to pursue this research topic. For example, an Australian paper on substituting vaccinia virus for variola virus for Orthoviruses research had six Russian scientists' names on it. Although the point of the experiment was to detect vaccinia virions in aerosolized material, this research shows knowledge of substituting vaccinia for variola and aerosolizing droplets (i.e. proliferation), then sampling and detecting for long-term monitoring of personnel exposure in a bioaerosol-contaminated environment (i.e. couunterproliferation). Under the right circumstances similar scientists could be persuaded to work for rogue entities or governments, thus raising the level of concern that poxviruses could be used in a bioweapon.

As a result of generalized concern about smallpox or a related virus as a bioterrorism agent, many countries have established detection assays and public health disaster plans to counter such an event. Some of these countries have begun to stockpile vaccines and to develop anti-viral drugs that might be effective should a poxvirus event occur.

Many Americans were vaccinated against smallpox decades ago, and it is likely that of these some individuals are no longer protected, some are partially protected and would therefore get mild disease if infected, and some are fully protected. The great majority of Americans born after 1972 were not immunized and would not be protected. The old vaccine, however, is too toxic in too many people to be given to everyone in a non-crisis situation. A newer vaccine with far fewer side effects might be acceptable if needed.

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### Analysis

Naturally occurring smallpox disease was eliminated worldwide in 1977. Routine vaccination of US civilians against smallpox was discontinued in 1971, but allowed for travelers to endemic regions until the late 1970s. In most other countries, vaccination of the general population ended by 1982. As a result of this halt in vaccination, most of the US population could now become ill with smallpox disease should it be reintroduced by accident or intentionally. In addition, humans are susceptible to several naturally occurring viruses related to smallpox, one of which could become a serious disease risk through natural evolution. Routine smallpox vaccination previously protected against these viruses. Finally, there is concern about the potential creation of a genetically engineered poxvirus that might be markedly pathogenic for humans.

When smallpox was being eradicated worldwide, the World Health Organization (WHO) moved to eliminate the causative variola virus from all laboratories and strain collections. The first step in 1984 was to confine variola virus samples to two centers, one in the United States and one in Russia. However, whether all other strain collections were destroyed or turned over to those two centers has been a matter of controversy and concern. It was feared that certain nations may have decided to retain samples of variola virus. At least as likely was the possibility that viral collections that included samples of variola viruses may have been retained by individual scientists in various countries through (i) defiance, (ii) laziness in not searching for variola samples, or (iii) inertia, despite knowing that such samples were present. The next step for the WHO was to propose eliminating variola virus samples even from the two designated centers. The United States balked, concerned that Russia might not destroy all of its stocks and that other collections of variola virus might still exist clandestinely. The United States argued that additional research was needed to better understand the virus in order to develop suitable anti-smallpox drugs that might be used should the disease recur for whatever reason. Indeed, laboratory security for serious pathogens throughout much of the world, including the United States, was rather lax until the recent past. As a result, thefts of worrisome bioagents could have been carried out many times, an issue with regard to potential reintroduction of smallpox disease into the human population from pilfered viruses (1).

A defector from Russia, who had been involved in biological warfare (BW) activities, stated and published that Russia had had a secret BW research program that included variola virus work and that the program had culminated in the weaponization of the virus for BW delivery (2). This testimony, along with the hint by Alibek that hidden stocks of smallpox virus might exist, increased the resolve of US officials to retain its samples of variola virus for defensive research. That position was supported by a study of the US National Academy of Sciences, which recommended areas of research that should be done with live variola virus, considering the bio-threat potential (3).

A Russian paper published in 2006 (4) stated that "smallpox virus is kept in laboratories of *some* or *certain* countries." [Italics added for emphasis.] This wording hints at possible Russian knowledge of variola virus stocks in countries other than the United

States and Russia, but may merely refer to the two countries known to have such stocks: Russia and the United States. Another Russian paper states that according to American experts: "at least 10 countries have such collections and this, through accidental leaks [spillages], could possibly endanger the entire world health protection system" (5). Indeed, many other countries are concerned about the existence of unofficial caches of variola virus (6-14).

Russian scientists at VECTOR created plasmids each carrying the complete genomes of one of thirteen strains of variola virus (15-17). Such plasmids could potentially be employed to reconstitute a viable variola virus or used to create a modified virus with unusual properties. Moreover, those plasmids could be used to separately transport non-infectious parts of variola virus for later reassembly. It is recognized that such a process would require sophisticated molecular virology procedures.

Concerns about *Orthopoxviruses* (the group of viruses that comprises variola virus and the poxviruses most closely related to variola virus) include the following:

- 1. *Orthopoxviruses* are stable and can be stored for long periods of time. A small sample of an *Orthopoxvirus* can be grown up in a suitable laboratory so that large batches of viruses are obtained.
- 2. Hidden caches of variola viruses that should have been destroyed decades ago, but which may not have been destroyed, might exist and be tapped to grow up large amounts of virus for potential use in a terrorist event (8).
- 3. Stocks of variola virus held in Russia might have been raided at a time in the past when security was lax, or taken by Russian laboratory workers leaving for jobs in other countries (7). A portion of such viral stocks might be grown up and used by terrorist groups.
- 4. Current Russian variola stocks might be diverted to terrorist groups or rogue states.
- 5. A virulent monkeypox virus (already a pathogen for humans) might be altered by natural means or by genetic engineering in a laboratory to make it more virulent, perhaps even as virulent as variola (smallpox) virus (18). For example, the gene encoding the IL1*beta*-related protein present in monkeypox virus but absent in variola virus could be deleted in monkeypox virus, which might make the resulting virus more deadly (19).
- 6. Another virus related to variola virus, such as cowpox virus, camelpox virus, or any of several vaccinia viruses, might be modified genetically so as to make it a serious pathogen. These genetic modifications could be mediated through *in vitro* recombination experiments, by random mutagenesis, or by directed addition of virulence genes.
- 7. A number of *Orthopoxviruses* are able to infect humans (e.g., cowpox, monkeypox, vaccinia variants, camelpox). Aside from variola and certain strains of monkeypox virus, they tend not to be passed efficiently from human to human by droplet transmission and they cause relatively mild disease. However, various genetic manipulations, including (but not limited to) the addition of a cytokine gene, such as that encoding interleukin 4 (IL4), might turn an *Orthopoxvirus* that

causes only mild to moderate disease in humans into a lethal infection. It is already known that recombinant vaccinia virus carrying the mouse IL4 gene is lethal for mice (20).

When smallpox was a widespread disease, a certain number of adverse effects of the vaccine were tolerated. However, a need for new, safer vaccines has arisen because the adverse events from the old vaccines were deemed unacceptable in the absence of the disease.

Worldwide concerns have caused many countries to disseminate response plans in case of an *Orthopoxvirus* outbreak, develop new vaccines (21-29), and construct mathematical models of disease spread (14, 30-40).

# Immunological Aspects of Poxvirus Infections

The mammalian immune system uses many soluble mediators to protect against a variety of infections. One of these soluble mediators, IL4, is produced by a subset of T lymphocytes (called Th2 cells) during an immune response. IL4 promotes antibody production but interferes with the development of T cell mediated immunity, including the formation of cytotoxic T cells and production of interferon-gamma. However, cytotoxic T cells and interferon-gamma are critical to eliminating viruses from the body during a viral infection. Consistent with these concepts, IL4 expression caused a delay in vaccinia virus clearance from target organs (41). In studies with far-reaching implications, Australian scientists engineered the gene encoding mouse IL4 into mousepox virus (also called ectromelia virus) and discovered that the mousepox virus bearing the IL4 gene had very different properties than ordinary mousepox virus (41, 42). The IL4-bearing mousepox virus acted as a super-pathogen. It caused disease in mouse strains normally genetically resistant to mousepox, leading to a high mortality. This finding with the IL4-modified mousepox virus has spread a fear that a genetically engineered Orthopoxvirus able to infect humans, such as vaccinia virus or cowpox virus, might be made more virulent for humans by similar genetic engineering. IL4 carrying vaccinia virus was found to be lethal for mice (20). It should be noted that human IL4 and mouse IL4 have structural differences (43). As a result, the effects on mice of a vaccinia virus carrying the mouse IL 4 gene may be species specific (43, 44). However, it is likely that a vaccinia virus carrying the gene for human IL4 would be a super pathogen in humans.

In the Australian studies, the mouse IL4 gene was incorporated into the mousepox virus, and the genetically modified virus administered to laboratory mice. That single genetic alteration (addition of the gene encoding IL4) led to fatal disease in all strains of mice tested. Of perhaps even greater concern, the IL4-bearing mousepox virus overcame standard immunizations that normally protect mice from mousepox disease. That is, mice vaccinated against mousepox resisted infection with the usual mousepox virus but contracted the fatal disease when they were given a mousepox virus genetically engineered to carry the mouse IL4 gene (42). The failure of immunization to protect

mice against such an engineered poxvirus suggests that the IL4-bearing virus interfered with the expression of memory immune responses. Indeed, the physiological basis of the IL4 effect is largely understood. It appears that two kinds of cells of the immune system, NK cells and cytotoxic CD8+ T cells, mediate resistance to mousepox virus both by direct cellular action and through production of interferon-gamma (42, 45). Apparently, IL4 inhibited these two cell types critical for clearing virus-infected host cells. The cytotoxic CD8+ T cells were indirectly inhibited by IL4. IL4 suppressed the development of Th1 type T cells (41). Such suppression interfered with generation of CD8+ cytotoxic T cells. Overall, the marked over-expression of IL4 caused by the genetically engineered viruses prevented viral clearance and markedly worsened the disease. It was also noted that the antiviral drug cidofovir failed to protect mice from the mousepox virus that bore the mouse IL4 gene (46).

This set of results indicates that it is entirely possible that genetic modification of an *Orthopoxvirus* that causes mild human disease (vaccinia, monkeypox, cowpox, camelpox, etc.) might produce an *Orthopoxvirus* that causes a disease in humans with the severity of smallpox. Thus, one might not even need to obtain samples of variola virus to create a serious poxvirus outbreak – modification of a readily available poxvirus might serve such a purpose. Moreover, an engineered *Orthopoxvirus* might even be virulent in people immunized against smallpox and other poxviruses. Therefore, experiments in which immune mediators are engineered into *Orthopoxviruses* should be viewed with concern.

As part of a substantial research program involving poxviruses, French scientists engineered human IL2 into vaccinia virus to study feline fibrosarcomas (47). Although that work is directed at treatment of cancers (47), such experiments are worrisome. They would be of greater concern if an adversary had performed them, because interleukins have the potential to make *Orthopoxviruses*, (including vaccinia viruses, monkeypox virus, cowpox virus, camelpox virus and even variola virus itself) more virulent than they otherwise would be. In other experiments, Spanish scientists engineered vaccinia virus to carry the genes encoding IL12 and IL18. Their studies demonstrated that those two cytokines facilitate the clearance of the virus (48). However, it is of concern that cytokines are being engineered into vaccinia viruses and perhaps other *Orthopoxviruses*.

Several *Orthopoxviruses* were found to carry genes encoding proteins able to bind IL18 (49, 50). The binding of IL18 increased virulence (50). Other methods of inhibition of IL18 also were found to increase virulence (56), which could point to various approaches to interfering with IL18 so as to render a poxvirus infection far more serious. Other *Orthopoxvirus* virulence factors also have been studied (52-58). Manipulation of some of these could possibly increase virulence.

Japanese scientists genetically engineered vaccinia viruses to bear human cytokine genes (59). The cytokine genes included those encoding IL12, IL23, and IL17 (59). Although none of these constructs are of direct concern, engineering of interleukin genes into a poxvirus pathogenic for humans is worrisome. These studies further found that IL17 and IL23 are important in resisting the virulence of the virus (59). Therefore, interfering with

IL17 and/or IL23 host responses could increase the severity of an *Orthopoxvirus* infection of humans.

Although general fears about potential genetically engineered viral and bacterial pathogens may be excessive in many circumstances, the cautions about an engineered *Orthopoxvirus* carrying a cytokine gene cannot be overemphasized. That is because one of the readily available relatives of the variola (smallpox) virus could possibly be made into a virus lethal to humans by such a maneuver.

A variety of host genetic factors – including but not limited to a preprogrammed suite of cytokine responses – may play a role in the relative susceptibility of different people to specific *Orthopoxviruses*. For example, a certain major histocompatibility complex type (HLA-A\*201) has been associated with cytotoxic T cell responses to a variola virus epitope (24). This trait may be one factor, along with preprogrammed cytokine production profiles and other factors, that underlies differences among people or groups of people in susceptibility to the virus.

In addition to its role as a vaccine for smallpox, vaccinia virus has been used as a vector for genetic engineering to deliver genes and/or to function as a carrier in immunizations against a variety of diseases, including HIV and many other infections (60-88). Vaccinia is especially useful in this regard because it can be genetically engineered without loss of its ability to replicate, and the virus can take a large amount of additional, engineered DNA. This function represents a potential dual use for vaccinia viruses.

During Soviet times, Russian scientists intensively studied Orthopoxviruses. They did report the first complete sequencing of a virulent variola strain in 1992 (89); however, the depth and extent of the Russian work may not be reflected in the open literature. Although the United States sequenced a different various strain a year after the Russians, it is only in relatively recent years that other countries, including the United States, have carried out systematic and comparative genetic analyses of *Orthopoxviruses*. As a result of its long commitment to understanding the genetics of variola and related viruses, Russia has substantial experience to draw upon in terms of manipulating Orthopoxviruses. That represents an important basis of concern for the United States in terms of (i) agreeing to the destruction of all variola virus stocks, and (ii) the possibility of creation of genetically manipulated Orthopoxviruses by an adversary. The second item includes but is not limited to the possibility of using genetic engineering to add to a naturally occurring Orthopoxvirus pathogenic for humans (such as a relatively virulent strain of monkeypox virus or cowpox virus or a now naturally occurring vaccinia virus variant) an immune-modulating gene or genes (such as the gene encoding human IL4) so as to construct a virus more virulent for humans. It should be noted that Orthopoxviruses are very large and able to readily accept large amounts of additional DNA by genetic engineering without a need for the very sophisticated methods sometimes required in the manipulation of smaller viruses with RNA genomes.

Camelpox virus rarely induces illness in people, and then typically only a mild case occurs, although the virus causes a serious and often lethal disease in camels. The full

genetic sequencing and analysis of camelpox virus led to the discovery that the camelpox virus is most closely related to variola virus, the cause of smallpox (90). Indeed, variola virus and camelpox virus produce the same kind of small white pocks on the chorioallantoic membranes of fertilized hens' eggs and have a similar ceiling temperature for the growth of such pocks. The two viruses are unable to grow in rabbit skin. Genetic comparisons indicate that (i) the genome sequence of the two viruses are closer than either is to any other virus, (ii) the arrangements of the open reading frames (protein coding sequences) are most similar to each other, (iii) the protein sequences themselves are most closely related to each other, and (iv) the nature and structure of the terminal repeats are most similar between variola and camelpox viruses (90). As camelpox is endemic in regions with large numbers of camels, the discovery of this close-relatedness between variola virus and camelpox virus triggered an immediate concern that terrorists or rogue states might first acquire camelpox virus from natural sources in their territories and then, somehow, convert the camelpox virus into a virus lethal for humans. Although this concern is appropriate, and it might be possible to convert camelpox virus into a variola-like virus lethal to humans, it also might take a substantial research program to turn camelpox virus into a new human pathogen. How many differences between camelpox virus and variola virus are responsible for the current modest virulence of camelpox virus for humans remains unclear. Stated another way, the number of genetic alterations that would have to be made in camelpox virus to convert it into a worrisome human pathogen is uncertain. It might be necessary to make modifications in many of the individual host-range genes of camelpox virus to convert that virus into a serious human pathogen. However, it also may be possible to add one or a few cytokine or cytokine receptor genes to achieve that goal.

In a study conducted by British and Irish scientists, it was observed that camelpox virus encodes a Schlafen-like protein that has mixed effects on virulence and is a gene that is disrupted in variola virus (91). It is conceivable that this gene inhibits virulence and that its disruption in camelpox virus might possibly make the latter more virulent for humans.

Over the years, Russian scientists sequenced the DNA genomes of many *Orthopoxviruses* and compared the encoded protein sequences (89, 92). For example, a 1995 paper compared in detail the sequences of a variola virus strain that had caused an outbreak of smallpox in India in 1967 with a variola strain that had caused an outbreak in Bangladesh in 1975 (93). Alignment of the DNA genomes showed a 99.3% homology. Of 200 protein sequences, 122 were identical, 42 had a substitution in a single amino acid, 11 had changes in two amino acids, and the other 25 had more extensive differences. It was noted that a region of conservation in variola virus is variable in vaccinia viruses (93). Such studies can assist in forensic analyses and in construction of virus variants.

The years of Russian work on poxviruses provided them with insights into the mechanisms by which *Orthopoxviruses* elude host immune responses. Such insights could have given Russian scientists clues to genetic alterations of *Orthopoxviruses* that might make them more virulent. This idea has been magnified by the Australian report of a marked increase in disease severity and mortality in mice after infection with a mousepox virus engineered to carry the gene for mouse IL4 (42).

As time passes, however, and modern molecular genetics techniques become both more sophisticated and disseminated throughout the world, whatever advantages the Russians may once have had should gradually have been reduced or even lost. This is attributed, in part, to a marked reduction in funding for biological research in Russia during the 1990s. That loss of research money led to Russia's failure to develop a modern biotechnology industry and the desertion of many biological scientists. It also impaired Russia's ability to build on its viral research. Indeed, recent studies of *Orthopoxviruses* from the United Kingdom, Australia, and France appear to be as sophisticated as the recent Russian studies or more so. Even the papers from Brazil are as advanced as those of the Russians.

Scientific collaborations between US and Russian scientists are also to be considered. Some such collaborations have been supported by US funds, and were designed in part to usefully employ Russian scientists formerly involved in Soviet-Russian BW programs. These relatively recent US-Russian collaborative studies have included genetic analyses of variola virus strains and related efforts (89, 94). In addition to that work, Russian scientists have independently published, in recent years, results of sequencing of a number of *Orthopoxviruses* (92, 95-97). These studies have provided insights into virulence mechanisms and *Orthopoxvirus* evolution (95, 97, 98). It is unclear how much of this work was performed recently and how much was carried out many years ago and was not published at the time, but has now been updated and published as new work. It is also possible that in addition to providing funds for useful employment of Russian scientists previously working in areas of concern, US scientific interactions with the Russian scientists could possibly have contributed to a greater sophistication in recent Russian *Orthopoxvirus* research.

A paper from Australia on aerosols of *Orthopoxviruses* – here using vaccinia virus as a surrogate for variola virus – had six authors, all with Russian names (99). The scientists used a Collison nebulizer to aerosolize droplets of vaccinia virus; virus recovery was then detected by sampling the aerosolized material using a prototype sampler. Although we made no effort to investigate former work by these scientists (such would be beyond the scope of this project), the publication is a reminder that a number of Russian scientists with expertise in *Orthopoxviruses* may have moved to other countries and continued to work in their field of expertise.

Human tumor necrosis factor (TNF), despite its name, is an inflammatory cytokine that protects against many of the adverse consequences of viral infections. TNF is targeted by *Orthopoxviruses*, including variola virus, as part of the viral strategy of interfering with host anti-viral immune responses. Russian scientists used standard molecular genetics techniques to produce recombinant proteins from variola virus, monkeypox virus, and cowpox virus, with particular emphasis on immune modulatory proteins (100, 101). Such work could provide additional insights into how *Orthopoxviruses* evade anti-viral immune responses, and also yield potential approaches to altering a less pathogenic virus into one more pathogenic. Russian scientists found substantial homology among aligned sequences of the anti-TNF proteins of variola virus and the anti-TNF proteins of both

cowpox viruses and monkeypox viruses (102). Not reported are results of similar studies with camelpox virus, of interest due to the homology between variola virus and camelpox virus (90). Scientists interested in such a comparison could perform analyses from published sequence data. Functional studies of the TNF-binding proteins showed some differences between the variola-derived protein and those from the other *Orthopoxviruses* studied (102). However, more work would need to be done to understand the genetic and functional differences in detail.

Poxviruses show an impressive homology of promoter sequences between genera. As a result, it is possible to heat-kill a pathogenic poxvirus but "rescue" it by adding a live second non-pathogenic virus which would provide the enzymes needed for transcription and perhaps for other functions. In such a circumstance, it may be possible to retain the DNA template of the heat-killed pathogenic virus, which could possibly be used to make more of the infectious pathogenic virus (103). This process of non-genetic activation potentially could be used to convert a non-infectious smallpox virus into an infectious one. It also might be used to obtain a virulent variola virus from its full genomic DNA, for example, as carried by Russian scientists in plasmids, though it would take a fairly sophisticated molecular virology laboratory to carry out such a procedure (15, 105). This phenomenon also has been called heterologous reactivation, a process whereby a cell infected with one poxvirus can produce infectious virus from the pure DNA of a second poxvirus (106). Such a method could allow resurrection of a no longer extant poxvirus from its known DNA sequence after synthesis of the DNA itself. In such a manner, DNA derived from the sequence of variola virus might possibly be used to grow variola virus that contained a complement of proteins required for the first rounds of replication followed by packaging of fully functional variola viruses. As a result, destruction of all variola virus stocks would not necessarily mean that the earth would thereafter forever be free of the virus.

Indeed, it has been suggested by Chinese scientists that global warming might release smallpox virus from sites in the tundra or in glaciers so as to induce a new smallpox pandemic (107). Although variola viruses are rather stable in the environment, the chance that a natural epidemic might occur in this fashion is conceivable but unlikely. It remains possible, however, that such a source of variola virus might be available even after all laboratory samples are destroyed. Viable variola virus might be obtained from a natural depot and then grown for various purposes. In this regard, Russian scientists have pointed out that variola viruses can live for long periods of time – a hundred years or more – in the tissues of corpses in permafrost areas or after burial in crypts (5, 108).

Homologous recombination is a frequent occurrence in poxvirus-infected cells (109-111). Thus, simultaneous infection of a cell with two different *Orthopoxviruses* can lead to emergence of new *Orthopoxviruses*, some of which might be especially virulent for humans. Indeed, studies by Norwegian scientists demonstrated *in vitro* recombination between a vaccinia virus strain and a cowpox virus strain (109). It is possible that recombination between, for example, a camelpox virus and a vaccinia virus might yield recombinant viruses of concern. In one study, Canadian scientists found that the vaccinia virus DNA polymerase has certain properties that allow recombination (112). If such a

recombination occurred, the resulting modification of the *Orthopoxvirus* – even without change in its virulence – might make initial detection difficult if based upon restriction endonuclease patterns (110).

In populations immune to smallpox, as in the past in the United States, a mild infection in a partially immune individual would pose little risk to contacts. The situation is very different for a non-immune population. If a partially immune person were to be infected with variola virus, the residual immunity could reduce the severity of disease in that individual, but he or she might not effectively clear the undetected virus. This scenario would allow him or her to shed the virus while not incapacitated, thereby potentially infecting many contacts (113).

This possibility is relevant to certain terrorist scenarios that have been advanced. For example, it has been suggested that fanatics might allow themselves to be purposely infected with the variola virus and then walk through US airports to infect people around them. Smallpox experts have suggested that such a scenario is unlikely as people with contagious smallpox would be rendered too ill by the disease to carry out such an assignment. However, if a person had minimal immunity (e.g., from a vaccination many years before), he or she might be able to secrete virus and yet be physically strong enough to try to spread disease in public places.

Russian scientists working at VECTOR, a former BW research center, have published several scenarios of potential future outbreaks of smallpox (36). They have emphasized mathematical modeling of the spread of smallpox under various situations of bioterrorism (36). These authors suggest a need for communications among responders and decision-makers; requirements for medical personnel, hospital beds, and medical materials; and estimates of the rate of spread of the disease. Other Russian scientists at the same institution published mathematical models of the spread of microbiological agents of concern through the atmosphere, including modeling of movement through indoor air (37). Such work has influenced the design of buildings in the region and the modification of ventilation systems (37).

Scientists at another former BW research center in Russia – Gamaleya Research Institute of Epidemiology and Microbiology – described a scenario in which variola virus was released in a city of a million people (38). The analysis included mathematical modeling of the spread of disease and potential consequences depending upon the reactions of public health authorities (38).

Between scenario development, computer modeling of the spread of smallpox, new oral vaccination methods, an initiative to immunize against smallpox, and development of human antibodies specific for variola virus, it appears that Russia is taking extensive measures to deal with a potential outbreak of smallpox. Russia has involved their Center for Military and Technical Problems of Biological Protection at the Ministry of Defense, as well as other agencies, in the mathematical modeling-computer simulation (39).

In contrast to the situation in Russia and the former Soviet Union, which had an intensive research effort into Orthopoxvirus genetics, China has shown little evidence in the open literature of a large, long-standing laboratory program into *Poxviruses*. However, in recent years Chinese scientists have manipulated poxviruses, and China's general programs in molecular virology appear to have narrowed the prior gap between itself and Russia. Indeed, Chinese scientists have analyzed immune modulating poxvirus proteins (114, 115), once a subject of substantial research by Soviet scientists. Chinese scientists have cloned an anti-apoptosis virulence factor of goatpox virus and analyzed it (116). Such an experiment raises the possibility of Chinese scientists inserting various virulence factors into an *Orthopoxvirus* readily able to infect humans. Indeed, the Chinese have separately engineered a human interleukin gene and an interferon gene into an Orthopoxvirus (75, 76). Although these last experiments are of no direct concern, the approach and the techniques could possibly lead to creation of an Orthopoxvirus virulent for humans. In this regard, some of China's capabilities in molecular biology generally and specific analyses of certain molecules are relevant to understanding virulence mechanisms of Orthopoxviruses (117).

China has published its awareness of the possible bioterrorism use of variola virus (9-11) and expressly indicated that cultures of the variola virus stored in the former states of the Soviet Union could possibly have been commandeered by terrorists considering the employment problems after the disintegration of the Soviet Union and the departure of many Soviet scientists (118, 119). To protect itself from such a possible event, China has been exploring the use of various viruses as vaccines, including cowpox strains (9).

Over the years, the effectiveness of vaccination against smallpox with vaccinia virus reduced the impetus to develop drugs effective against the virus for use either in prophylaxis or in therapy. However, with the eradication of smallpox as a naturally occurring disease and suspension of routine public health vaccination programs, a gap was increasingly appreciated. Basically, it was recognized that, despite its effectiveness in protecting against the dreaded smallpox, vaccination with the old vaccinia viruses used in the past for smallpox vaccines caused unacceptable adverse side effects in too many recipients to justify its use in large populations against a disease that no longer existed in nature (61, 120-123). However, in a desire to avoid the potential of being unprepared for a possible terrorist attack with variola virus (the causative agent of smallpox) or a related virus, many countries have tried to develop either less toxic vaccines or new drugs that might serve to prevent or treat smallpox or a related disease caused by another Orthopoxvirus. As a result of these new efforts, developmental work has covered (i) vaccines with fewer adverse events, and (ii) testing of known drugs and new compounds for potential use as anti-Orthopoxvirus agents. This type of work is ongoing in several countries and has yielded some promising results (24-26, 122, 124-131). A DNA molecule containing immune-stimulatory CpG sequences also may assist in the therapy of Orthopoxvirus infections (124). However, just as antibiotic-resistant bacteria have become a problem, the potential for development of anti-viral resistant *Orthopoxviruses* also is a concern. Indeed, it was found that vaccinia virus strains became resistant to the anti-viral drug cidofovir during serial passage in Vero cells in culture (130). Crossresistance was found for compounds related to cidofovir but not to unrelated nucleosides

(130). This result also raises the issue of potential purposeful development of a virulent *Orthopoxvirus* pathogenic for humans that might be resistant to the most promising antismallpox drugs currently under serious study.

As time passes, new issues not considered in the past become important. For example, bovine spongiform encephalopathy (Mad Cow Disease) may be passed to people via animal proteins that bear the inducing prions. To prevent this from occurring via animal proteins contaminating a smallpox vaccine, there is a push in some countries to use vaccine produced in tissue culture (132).

In the event of a new poxvirus outbreak in people, responses can include mass vaccination or vaccination and quarantine of index cases and their contacts. The details of expected public health responses to a poxvirus outbreak provide data for mathematical models. However, a predicted and required rapid response may not occur in time for contact (ring) vaccination plus quarantine to contain an outbreak.

In addition to its use as a vaccine for smallpox, during the past 25 years vaccinia virus has been employed for a variety of secondary purposes, such as vectors for antigens or genes (65-67, 70, 72-75, 80, 81, 83-88, 133-149, 150-189). The virus has been used to transfer genes in medical research – in cancer treatment, for example. Such work depends upon the ease in growing vaccinia virus and its stability during storage as well as the ability of vaccinia virus to carry substantial quantities of foreign DNA and yet continue to function normally (133). In addition, many countries have been using vaccinia viruses to carry other antigens in immunization efforts. This last use is based in part on the ability of vaccinia virus to stimulate host dendritic cells, which are critical to initiating a strong host immune response (138, 163, 190).

### **Conclusions**

Even if US and Russian stocks of *variola virus* are destroyed, the threat of smallpox will not necessarily have been eliminated. The virus may be sequestered elsewhere in hidden collections or could be recovered from burial sites of humans who died of smallpox (108).

Several *Orthopoxviruses* currently replicating in nature are able to infect humans: monkeypox, cowpox, vaccinia, and others. These could evolve to be worse human pathogens. Such a process, however, might take many decades. These other *Orthopoxviruses* also could be genetically engineered in the laboratory into more virulent human pathogens by techniques known to virologists around the world.

Currently available vaccines will probably protect against the naturally occurring viruses, such as monkeypox, cowpox, and vaccinia viruses as they evolve, but they might not protect against certain genetically engineered viruses.

There has been concern regarding the level of persistent immunity against smallpox in individuals vaccinated decades ago. In addition, a certain level of confusion has been introduced by variable results regarding the degree of residual immunity from different studies in various countries. Some studies have shown substantial antibody levels against vaccinia virus in a number of individuals, whereas other studies describe less robust immunity. In one series of studies of interferon-gamma-producing memory T cells, only 20% of people vaccinated 13-25 years previously had adequate responses (191). However, in another study of residual immunity against smallpox, the presence of a vaccination scar plus a history of vaccination within 20-30 years correlated with good T cell reactivity (192). Overall, among people vaccinated against smallpox in the distant past, it is likely that (i) some such individuals are no longer protected, (ii) some are partially protected and would therefore get mild disease if infected with the variola virus, and (iii) some are completely protected. The great majority of Americans born since 1972 would not have been immunized and would not be protected

Short of a recurrence of smallpox or the emergence of a new aggressive *Orthopoxvirus*, Americans who are not immune should not be given the old vaccinia smallpox vaccine. That historical vaccine is too toxic, inducing unacceptable adverse reactions in many people (61, 120-123). A newer vaccine with far fewer side effects might be acceptable as a relatively routine immunogen should one be required.

#### **Publications Reviewed**

More than 2,000 papers on poxviruses from the worldwide open literature published between 2002 and the latter part of 2007 were reviewed for this report, as well as a few earlier papers (most of those were from Russia). Papers selected for analysis primarily dealt with genetics, molecular genetic manipulations, virulence mechanisms and modifications of those, and approaches to therapy.

### **Background Information on Smallpox and Related Poxviruses**

### General

The *Poxviridae* family contains large double-stranded DNA viruses that encode between 150 and 300 proteins. The portion of this family that infects vertebrates, called *Chordopoxvirinae*, encompasses eight genera. The *Orthopoxvirus* genus – with which we are especially concerned herein – contains viruses with a linear double-stranded DNA genome of between ~170,000 and ~220,000 base pairs and includes the following species: (i) **Variola virus** (the causative agent of human smallpox), (ii) Monkeypox virus (strains of which can induce a generalized infection in humans), (iii) Cowpox virus (which can infect humans), (iv) Vaccinia virus (used to vaccinate humans against smallpox, and which can infect humans). [Buffalopox virus is a variant of vaccinia virus that can infect humans (193, 194). There are several Brazilian variants of vaccinia virus circulating in the wild, some of which have caused disease in people (195). Rabbitpox

virus is believed to be a vaccinia variant that did not directly evolve from any sequenced vaccinia virus strain (196). Although it causes severe disease in rabbits, that virus does not affect humans.] (v) Camelpox virus, which is closely related to variola virus and which has occasionally caused relatively mild disease in humans, (vi) volepox virus, (vii) skunkpox virus, (viii) racoonpox virus (197), (ix) Tatera virus (gerbilpox virus), (x) mousepox virus (called ectromelia virus), and (xi) Uasin Gishu (horsepox) disease virus (198). Racoonpox, volepox, and skunkpox viruses are North American viruses.

Variola virus, the causative agent of smallpox, is closely related by DNA homology studies to camelpox virus and Tatera (gerbilpox) virus and also is related to skunkpox and racoonpox viruses.

Overall, there are *Orthopoxviruses* that can cause generalized disease in humans, others that typically lead only to limited skin infections, and some that have not infected humans.

# Smallpox Epidemiological Issues

Smallpox (caused by variola virus) afflicted humans for thousands of years. When introduced into completely non-immune populations, such as the Native Americans of South America and North America, the death toll was especially severe. This is an issue today as routine vaccination against smallpox has been suspended worldwide since about 1982 with the worldwide elimination of the naturally occurring disease in 1977. As a result of this suspension of vaccination against smallpox, most of the US population is now susceptible to smallpox disease should it be reintroduced by accident. Moreover, there has been concern about the use of variola virus as a biological warfare (BW) or bioterrorism agent (2, 199).

After smallpox disease had been eradicated worldwide, the World Health Organization moved to eliminate the causative virus from all laboratories and strain collections. Variola viruses were first to be confined to two centers, one in the United States and one in Russia, and those two strain collections later destroyed. However, these viral collections have not yet been eliminated. Moreover, there is some concern that other variola stocks might exist in countries that may not have destroyed their variola virus collections as planned by the WHO. The United States has argued that additional research was needed to better understand the virus in order to develop suitable antismallpox drugs in case the disease recurred for whatever reason.

# Clinical Features of Smallpox

Person-to-person spread of smallpox is primarily by respiratory droplets and less often by viral transmission via scab material. The virus particles in the aerosol droplets of an infected individual originate in erupting lesions on the pharynx, larynx, tongue, and

uvula, and less often from the trachea and bronchi. Such viral secretion starts slightly before or at the same time as the skin rash and persists for approximately one week.

After a person is newly infected with variola virus, the incubation period prior to the development of clinical smallpox is approximately 10-14 days. The disease is characterized by the sudden onset of headache, backache, fever, and malaise. The typical rash occurs three to four days later: the first appearance is on the person's buccal and pharyngeal mucosae, followed quickly by eruptions on the face, forearms, and hands. Initially, the lesions are macules (flat), followed by firm papules (heaped up), vesicles (fluid filled), and finally pustules (with white blood cell accumulation). Eight to ten days after onset of the rash, the pustules dry up. Healing leaves pockmark scars, especially on the face. In smallpox, the rash typically occurs distally (i) on the face to a greater extent than on the trunk, (ii) on the legs and forearms more intensely than the arms and thighs. In contrast, chickenpox is distinguished by a central (trunk, arms, and thighs) rather than a peripheral pattern of rash. Rash on the hands and/or feet should point to smallpox rather than chickenpox.

One clinical type of smallpox, caused by variola major, had a case fatality rate of  $\sim 30\%$ , whereas another clinical type, variola minor, had a case fatality rate of only 1%. These clinical features correlate with genetic differences between the strains.

Vaccination with vaccinia virus vaccine strains is protective against smallpox for some five to ten years, with waning immunity thereafter. Even some immunity may render the disease much milder than would be the case without any prior immunizations. Nonetheless, as most people in the United States were last vaccinated before 1971 or not at all, the majority of the population is considered to lack immunity to smallpox.

# Laboratory Diagnosis of Smallpox and Other Orthopoxviruses

Prior to the elimination of smallpox as a naturally occurring disease, the clinical features were usually sufficient to make an accurate diagnosis of smallpox. For confirmation, virus-containing fluids from skin vesicles or scabs were tested on the chorioallantoic membranes of chick embryos, and the patterns and colors of the pocks that formed on those membranes were used to make a diagnosis of the virus species. Electron microscopic features also were examined for laboratory confirmation. Since then, PCR assays have been developed to detect *Orthopoxviruses* generically and also to distinguish among the eleven species of *Orthopoxviruses* (95, 97, 110, 200-220).

The *Orthopoxviruses* make an HA protein, whereas no other poxviruses make an HA protein. This has allowed development of a PCR reaction to distinguish *Orthopoxviruses* from other poxviruses.

PCR of the HA gene can distinguish the various species in the *Orthopoxvirus* genus. Such assays can even distinguish a variola major strain from a variola minor strain (221,

222). However, some studies suggest that certain cowpox virus strains have an HA gene that is detected by variola primers and probes. Therefore, where possible, viral genome sequencing should be performed to provide a definitive method of species and strain identification.

There is only about a 75% genomic DNA homology between Old World *Orthopoxviruses* and *Orthopoxviruses* isolated in North America. In contrast, among the Old World *Orthopoxviruses*, the sequence homology is ~ 95%. When *Orthopoxviruses* are divided into two groups, Group I includes Old World vaccinia virus (along with its variants: rabbitpox and buffalopox), monkeypox viruses, and the closely related ectromelia virus (mousepox). Group II includes variola virus (the causative agent of smallpox) and the closely related camelpox and tatera gerbilpox viruses. Also included are skunkpox and racoonpox viruses. Each of those two groups of viruses has a genetic relatedness to a different group of cowpox viruses, in keeping with the idea that an ancient cowpox virus may have given rise to all of the *Orthopoxviruses* (223).

# Monkeypox

With the eradication of smallpox as a disease in nature, monkeypox virus has become the naturally occurring *Orthopoxvirus* most able to cause systemic disease in immunologically intact people. Human disease caused by the monkeypox virus is found in the tropical rainforests of western and central Africa. Some monkeypox strains cause less severe disease in humans than others (224).

The incubation period for monkeypox is approximately 12 days, followed by marked malaise with severe muscle aches, headaches, and cough. The rash and clinical features of human monkeypox are indistinguishable from smallpox except that monkeypox is characterized by more marked lymph node enlargement (89). Person-to-person spread of monkeypox is less common than for smallpox, from rare up to 30% of human cases in different outbreaks (89, 225, 226). People most often become infected with monkeypox virus by handling infected animals (wherein the virus is thought to enter via cuts or abrasions) or from eating infected monkeys (bushmeat) or infected squirrels.

Although monkeys develop monkeypox disease, and many have antibodies to the virus, the animal reservoir for monkeypox virus is believed to be squirrels. The PCR test can diagnose monkeypox virus as an *Orthopoxvirus*, and specific primers can distinguish monkeypox virus from other *Orthopoxviruses*. Another laboratory method is the inoculation of virus-containing fluids from skin vesicles or scabs onto test chorioallantoic membranes of chick embryos. The appearance of the pocks that form on those membranes is used to make a species diagnosis. In addition, the broad susceptibility of various test animals to the virus assists in the identification of monkeypox virus.

Purposeful monkeypox virus infection of monkeys demonstrated that post-infection smallpox vaccination was not as good as anti-viral drug therapy (227). Whether this will hold for humans remains to be determined.

An outbreak of monkeypox occurred in 2003 in the United States as a result of the importation of a virus-carrying Gambian pouched giant rat. The African rodent infected pet prairie dogs (intermediate hosts) in the US facility, which led to disease in 72 people (half confirmed in the laboratory) some of whom became seriously ill (224, 228-230). Fortunately, this infection was from a variant strain of monkeypox virus that causes relatively mild disease and shows limited human-to-human transmission (224, 229). Infections occurred largely from contact, many presumably through skin lesions (230). Nonetheless, that outbreak raised the specter of a serious monkeypox outbreak outside of Africa and even the possibility that monkeypox might begin to fill the niche that smallpox had occupied for thousands of years.

Differences in the genetics of the different strains of monkeypox virus were found by Russian scientists to explain the differences in virulence of the strains (231). The Central African monkeypox virus has factors preventing host immune responses to infection with the virus, making it especially virulent. The West African strain has a deletion of one such gene, which encodes the complement-binding protein that interferes with host-antiviral responses. Without that gene product, the virus is more readily cleared by the host and it causes less severe disease (89, 231). Fortunately, it was the milder form that was imported into the United States in 2003.

Genetic studies suggest that monkeypox arose independently of smallpox (19, 89). The central regions of their genomes are 94-96% homologous; however, the outer, species-specific, regions are less homologous (19, 89, 226).

Genetic comparisons from a US-Russian collaboration suggest that the monkeypox virus would not easily transform into a virus like variola virus under natural selection in the wild in the near term, though that could possibly occur over the stretch of future history (19).

# Cowpox

Cowpox viruses have large genomes and are the *Orthopoxvirus* with the most complete set of genes, suggesting to scientists that they are the likely precursors of the other extant *Orthopoxviruses*. For example, several genes in cowpox viruses are found in only fragmentary forms in the other *Orthopoxviruses* (223, 232). Further analysis indicates that there are two genetically separable groups of cowpox viruses that may have diverged evolutionarily long ago (232). It is thought that each of those two cowpox virus groups is the precursor of the two major groups of *Orthopoxviruses*.

As a disease of cows, cowpox is manifested as ulcers on the teats, which leads to pustular lesions on the hands of people milking the cows. More than two centuries ago, such lesions on the hands of milkmaids were found to protect against the subsequent development of smallpox. However, the animal reservoirs of cowpox viruses are not cows but several kinds of rodents (bank voles, field voles, and wood mice in the UK and

elsewhere in Europe, susliks and gerbils in Turkmenistan, root voles in Russia, and lemmings in Norway) (233-235). Such animals can infect additional mammals, including domestic cats and zoo animals (236, 237). Humans infected with cowpox viruses can have virus circulating in their blood streams even if the disease is relatively limited (238). Human cowpox disease currently is contracted primarily from infected domestic cats (which had contracted the virus from rodents) (203, 239-244), although dogs (245) and horses (246) also may be implicated.

Cowpox in people has an incubation period of about seven days, followed by the sudden appearance of fever, malaise, headaches, and muscle aches, sometimes accompanied by nausea and vomiting. Skin lesions are most common on the hands and face. Local lymphadenopathy is marked. Healing of the skin takes place over several weeks, leaving a large depressed scar. The disease can be fatal.

It has been found that cowpox virus interferes with induction of host nuclear factorkappaB, a global immune regulatory transcription factor (247). This effect could contribute to the failure of host immune responses to adequately clear the virus. It is presumed that related mechanisms would occur in variola virus infection.

From an evolutionary perspective, a fit *Orthopoxvirus* would have the greatest host range (be able to infect many different animal species), be transmitted effectively from animal to animal in a not very dense population, would replicate rapidly and effectively in each host, and would balance its pathogenicity with its ability to reproduce and be passed on to other hosts. From this perspective, cowpox virus in perhaps the most evolutionarily fit *Orthopoxvirus* and variola virus the least fit (248).

#### Vaccinia

Vaccinia virus, the vaccine strain for preventing smallpox, was the first animal virus seen microscopically, grown in tissue culture, and purified. However, vaccinia viruses have a poorly understood origin. They may have originated from a now extinct *Orthopoxvirus* of horses that caused a disease called "grease" (249) or perhaps from a recombinant of cowpox virus and this grease-causing virus. The vaccine virus was passaged many times, which undoubtedly introduced several changes into the viral genome. Indeed, the resultant vaccine virus variants were selected for those strains that induced less pain, fever, and other complications after vaccination of people.

As a result of this process, vaccinia viruses were kept as laboratory strains for long periods of time. This led to a belief by many scientists that there was no natural reservoir for vaccinia (250). However, the genome of horsepox virus isolated from Mongolian horses points to a vaccinia-like structure (249). This finding suggests the possibility of a relationship between horsepox virus and the ancestral strain originally used for vaccination and from which vaccinia virus may have been derived.

Several variants of vaccinia virus have escaped into animal populations and now circulate in nature. Some of these vaccinia variants can cause disease in humans. For example, buffalopox in India is due to vaccinia virus variants, of which several have been distinguished (193, 194). A buffalopox virus can cause skin disease in humans who touch the infected teats of milking water buffalo or dairy cows, but systemic symptoms are usually quite mild or absent. Buffalopox viruses also cause oropharyngeal lesions in people who drink unpasteurized milk carrying the virus (193, 228). Rabbitpoxvirus, which infects rabbits, also is a vaccinia virus variant, but one that does not usually infect people and which may not have derived from a vaccine strain (196).

A number of strains of vaccinia virus circulate in various animals in Brazil (250-256), strains that show divergent evolution but which appears to have derived from vaccine strains. Many of these strains have infected people.

In addition to its role as a vaccine for smallpox, vaccinia virus has been used as a vector via genetic engineering to deliver genes and/or to function as a carrier in immunizations against a variety of diseases (including HIV and other infections). Vaccinia is especially useful in this regard because it can be genetically engineered to accept a large amount of additional DNA without loss of its ability to replicate.

# Camelpox virus

Camelpox virus is the virus most closely related to variola virus, the cause of smallpox (90). Variola virus and camelpox viruses produce the same small white pocks on the chorioallantoic membranes of fertilized hens' eggs and have a similar upper temperature limit for growth of such pocks. Neither of the two viruses can grow in rabbit skin. Genetic analyses show that (i) the sequences of the two viruses are closer than either is to any other virus, (ii) the arrangements of the protein coding regions are most similar to each other, (iii) the protein sequences themselves are most closely related, and (iv) the nature and structure of the terminal repeats are most similar between variola and camelpox viruses (90). The observation that camelpox is endemic in camels has raised the concern that a camelpox virus isolated from animals in one or more regions might somehow be converted into a lethal, variola-like human pathogen.

The susceptibility of various potential host species to camelpox virus has been incompletely studied. Rabbits, guinea pigs, and three-day old chickens are reportedly susceptible (257); however, studies with the M-96 strain of camelpox virus could not confirm those findings (257), suggesting possible variability from strain to strain.

Russian scientists also have compared the genetic organization of camelpox virus with those of other *Orthopoxviruses*. Their studies suggest that variola virus and camelpox viruses diverged approximately 6,000 years ago (96, 248), whereas modern *Orthopoxvirus* species derived from a precursor virus ~12,000-14,000 years ago (89).

### Other poxviruses that can sometimes cause disease in humans

**Parapoxviruses** cause disease in sheep and goats, called "orf," characterized by papules and vesicles on the mouth and nostrils of the animals (258). Such disease in lambs can cause lesions on udders of nursing ewes and vice versa. The diseases are found in Africa (especially north of the equator), western Asia, India, China, and Bangladesh (258). Although orf once had been thought to be caused by the same capripoxvirus in both sheep and goats, recent work suggests that the sheep and goat diseases may be caused by viruses that have genetic differences that allow molecular tests to discriminate them (258, 259). In fact, some of the capripoxviruses are specific for sheep (sheep pox viruses), some are specific for goats (goat pox viruses), and some may infect either sheep or goats (a less discriminating form of sheep pox virus) (258). Recombinants also occur between various strains of sheep pox virus. This type of disease can occur in cattle, caused by (i) bovine papular stomatitis virus or (ii) *Pseudocowpox* virus. Humans touching the teats during milking contract the virus and develop local lesions on the hands. The human lesions resolve within six weeks. There is also a seal parapoxvirus (Sealpoxvirus) that rarely has infected humans. Occasionally, poxvirus infections of humans can be misdiagnosed as an Orthopoxvirus when the infecting agent is a Parapoxvirus (260).

# **Yatapoxviruses**

Tanapoxvirus and Yaba monkey tumor poxvirus are *Yatapoxviruses* that occasionally infect humans. The animal reservoirs in the wild of these two viruses are unknown as is the mode of transmission to humans.

# Molluscum Contagiosum

A skin disease of humans in which the skin seems to be heaped up is caused by the Molluscum Contagiosum virus. Although especially found in children, in recent years this disorder has occurred in many AIDS patients. Indeed, Molluscum Contagiosum has become a marker for sexually transmitted diseases generally.

# Structure of Orthopoxviruses

These are big, brick-shaped viruses, able to be seen on light microscopy. The outer part of the viruses consists of a membrane composed of tube-like lipoprotein subunits. This membrane encloses a dumbbell-shaped core and two "lateral bodies" (261). The core consists of the viral double-stranded (ds) DNA and associated proteins. Cowpox virus is the largest of the *Orthopoxviruses* at ~220,000 base pairs. Variola viruses run at approximately 186,000 base pairs of dsDNA. The DNA has a C+G content of ~34%. These viruses have been sequenced (95-97, 221). Among the genes found in common, variola, vaccinia, and cowpox viruses have >90% genetic identity.

Each end of the virus DNA has an inverted terminal repeat (ITR) that is A+T rich and incompletely base paired, giving it a hairpin loop that connects the two DNA strands (sometimes called cross-linking). Genes encoding species-non-specific "housekeeping" proteins involved in replication and other critical viral functions are found in the central 90-100 kb of the *Orthopoxvirus* genome and are highly conserved (89, 262). In contrast, genes encoding species-specific proteins (such as those that interfere with the immune functions of a particular host species) are located peripherally (toward the ends of the DNA (89, 262).

# **Poxvirus Replication**

Infection of a host cell by an *Orthopoxvirus* leads to major cytopathic effects, changes in membrane permeability, and inhibition of cellular DNA, RNA, and protein synthesis as the virus essentially takes control of such functions and directs them to producing viral molecules.

Poxvirus replication occurs completely in the cytoplasm, a feature seen in only one other type of virus: African swine fever virus. An *Orthopoxvirus* gene product associates with host cell membranes, including those of the endoplasmic reticulum, in helping to determine the cytoplasmic sites of viral replication (263).

Some *Orthopoxviruses*, such as vaccinia viruses, can enter nearly all kinds of cell lines and infect them. As a result, such viruses appear not to take advantage of a specific receptor for viral attachment and entry but may use a virus-binding site that allows viral attachment but not entry. Entry occurs when the cell-associated enveloped virion (CEV) fuses with the host cell membrane with or without an endocytic process, and the viral core is released into the cytoplasm (264). The genomes of poxviruses encode dozens of enzymes needed for transcription and replication of the viral genome. Some of these factors are pre-formed and present in the virion, including DNA-dependent RNA polymerase, capping enzyme, methylating enzymes, and transcription factor.

Pre-formed viral transcriptase initiates transcription [copying of viral DNA to make messenger RNA (mRNA)]. As a result, functional capped, polyadenylated, and methylated viral mRNAs are produced within minutes after infection of a host cell. Polypeptides produced by translation (making a polypeptide from the viral mRNA) act to finish uncoating the viral core. Transcription of ~100 "early" genes occurs prior to the initiation of viral DNA synthesis. These "early" synthesized proteins include enzymes needed for replication of the viral DNA, such as DNA polymerase and thymidine kinase. Indeed, half of the vaccinia virus genome is transcribed even before replication of viral genomic DNA – an impressive "hitting the road running" adaptation.

This "early" process is followed by a shift in gene expression to "intermediate" gene transcription at approximately 100 minutes after infection of the host cell, and then by "late" gene transcription, which starts about 140 minutes after infection of the cell. These intermediate and late events happen via the action of specific viral proteins on promoter

sequences and may be controlled by positive and negative regulatory proteins. Most intermediate and late viral mRNA molecules have a 5' poly (A) tail, whereas most early mRNA do not. Viral DNA replication is thought to begin near the ends of the viral DNA genome.

Some viral proteins are secreted by infected cells. These proteins promote infection. Many are proteins that interfere with host immune responses, such as cytokine analogues and cytokine receptor analogues with functions relating to host interferons, interleukins, complement, chemokines, and tumor necrosis factor (89, 115, 228, 265-274). Included are (i) immunosubversive proteins that mimic host ligands, (ii) regulatory proteins ("virokines") such as homologues of epidermal growth factor, a complement regulatory protein, and virokines that confer resistance to host interferon, and (iii) "viroreceptors" that are homologues of cellular receptors but have lost their transmembrane anchors and signaling functions; these bind and sequester important immune regulatory molecules, preventing the molecules from reaching the appropriate host cell receptors. Included in this last category are molecules that bind tumor necrosis factor (TNF)-alpha, interleukin-1-beta, interleukin 18, CC chemokines, interferon alpha/beta, and interferon gamma (89).

Because of their importance for virulence, *Orthopoxviruses* produce two intracellular proteins and two extracellular proteins that interfere with the actions of host interferons (89). Besides interferons, tumor necrosis factors are the other critical immune actors in viral infections. Despite its name, human TNF is an inflammatory cytokine that protects against many adverse consequences of viral infections. TNF is targeted by *Orthopoxviruses*, including variola virus, as part of the viral strategy of interfering with host anti-viral immune responses (89). Russian scientists lined up the sequence of the anti-TNF protein of variola virus with anti-TNF proteins of cowpox viruses and monkeypox viruses and found substantial homology (102). Not reported are results of similar studies with camelpox virus, of interest because of the known homology between variola virus and camelpox virus. Functional studies of the TNF-binding proteins showed some differences between the variola-derived protein and those from the other *Orthopoxviruses* studied (102).

It has been found that cowpox virus and vaccinia virus produce factors that interfere with induction of host nuclear factor-*kappaB*, a global immune regulatory transcription factor (247, 268-270). This effect contributes to the failure of host immune responses to adequately clear the virus. It is presumed that related mechanisms would occur in variola virus infection. The more these virus-induced modulations are analyzed, the more viral proteins appear to be involved and the more complicated the interactions (268-270, 275).

The host complement system contributes to anti-viral responses. However, one of the vaccinia secreted proteins inhibits both the classical and alternative complement pathways of the host – this occurs through binding and inactivation of C4b and C3b complement molecules (231, 266, 267, 272, 276, 277). The variola virus complement control protein is more potent than that from vaccinia (272).

The *Orthopoxvirus* replication process within the infected cell uses specific areas of the cytoplasm for assembly of daughter viruses, sites that can be visualized by electron microscopy. A complete early transcription system is produced late in the infection of a cell and packaged within the core of infectious poxvirus particles. This allows the daughter virions to immediately begin their life cycles as soon as they enter other, non-infected host cells. Most of the daughter viral particles are released without envelopes via cell disruption. A smaller number are released with envelopes by exocytosis. Although both enveloped and non-enveloped viruses are infectious, the enveloped viruses are more infectious (278).

Homologous recombination is a frequent occurrence in poxvirus-infected cells. Thus simultaneous infection of a cell with two different poxviruses can lead to emergence of new poxviruses with recombined genes (232).

## Treatment and Vaccination – a 21st Century Issue

Vaccination with live vaccinia virus was effective in protecting against smallpox; however, the vaccine did cause adverse reactions, including generalized vaccinia disease in some people with eczema or immune deficiencies (61, 120-122, 279). Historically, approximately 30 people per million vaccinated had a life-threatening reaction to the vaccine and one to two people per million vaccinated died with one preparation (120, 122) and there were up to eight deaths per million people vaccinated with another vaccine preparation (120). Adverse reactions included eczema vaccinatum, progressive vaccinia, post-vaccinial encephalitis, fetal vaccinia, vaccinia keratitis, myopericarditis, and inadvertent transfer of virus from a newly vaccinated person to others (122, 123). Nowadays, up to 30% of people have contraindications to vaccination with live vaccinia virus, including cancer, HIV, heart problems, skin diseases, treatment with immunosuppressive medicines, pregnancy, or household contacts with such contraindications.

Despite these adverse events from vaccination against smallpox, over the years, the effectiveness of vaccination with vaccinia virus in preventing smallpox disease reduced the impetus to develop drugs effective against variola virus for use either in prophylaxis or in therapy of smallpox. However, with the eradication of smallpox as a naturally occurring disease and suspension of routine public health vaccination programs, a gap was increasingly appreciated. Despite its ability to protect against *Orthopoxviruses*, vaccination with vaccinia virus caused unacceptable side effects in too many recipients to justify its use in large populations against a disease that no longer existed in nature. In a desire to avoid the potential of being unprepared for a possible terrorist attack with variola virus (the causative agent of smallpox) or a related virus, many countries have tried to develop either less toxic vaccines or new drugs that might serve to prevent or treat smallpox or a related disease caused by a naturally occurring or genetically engineered *Orthopoxvirus*. As a result of these new efforts, developmental work has covered (i) vaccines with fewer adverse events, and (ii) testing of known drugs and new

compounds for potential use as anti-*Orthopoxvirus* agents. This work is ongoing in several countries and has yielded some promising results.

There is extensive immunological cross-reactivity and cross-protection among the *Poxviridae* strains within a given genus but little among strains in different genera. As a result, monkeypox virus, vaccinia virus, and cowpox virus protect against smallpox. This type of cross-protection has encouraged scientists to try several potential approaches to new vaccines. There is special interest in recombinant vaccines that might avoid the side effects seen in some people vaccinated with live vaccinia virus.

An effective immune response against a poxvirus includes both T cell-mediated immunity and antibody production. T lymphocytes in the form of cytotoxic T cells eliminate variola virus-infected cells even prior to viral replication, limiting an infection. Antibodies able to bind enveloped and non-enveloped virions protect against re-infection.

The standard US vaccine against smallpox was for many years a lyophilized live vaccinia virus preparation called Dryvax. A single vaccination protected against smallpox disease for about five years. A second vaccination protected for at least 10 years thereafter. Because of the long incubation period for smallpox, it is possible to effectively vaccinate someone even after he or she is exposed to the variola virus. Vaccination in the first four days after exposure can often prevent disease. Vaccine given on day 5 or 6 may ameliorate the disease to some extent (280). Patients with eczema may develop widespread disease from vaccination. Such complications have been treated successfully in the past with vaccinia immune globulin; however, such globulin is now in limited supply. The optimal time for harvesting such immune globulin is found to be 14 days after a revaccination (281).

For decades, Russian scientists have been evaluating an oral form of live vaccinia virus as a human vaccine. This oral vaccine has shown far fewer adverse reactions in humans than the standard skin scarification method with no diminution in immune protection (27). Russian workers also have combined the oral smallpox vaccine with an oral Hepatitis B vaccine (282).

A strain of vaccinia virus that had been used as a smallpox vaccine was found by Japanese scientists to become more virulent after growth in culture due to a mutation in a single gene (283). That gene could be eliminated without loss of protective immunity (283). The resulting strain was as protective as the US Dryvax strain and yet much less virulent. Additional work on the vaccine strain lacking the complicating [B5R] gene has produced what appears to be a much safer vaccine, one that might be sufficiently devoid of serious adverse effects to be considered for large population groups (21-23). This vaccine protects early in infection by inducing cell-mediated immunity more than antibodies (284). However, British scientists argue that retention of the B5 protein is critical for a smallpox vaccine because that immunogen is what is recognized by important antibodies (285). This is an issue in the trade off between good immunization and adverse events.

Monkeypox virus infection of monkeys demonstrated that post-infection vaccination was not as good as anti-viral drug therapy (227). Whether this tendency will hold for humans remains to be determined.

A DNA polymerase inhibitor used in HIV infections called cidofovir has anti-*Orthopoxvirus* activity; however, that drug can be nephrotoxic. Some related compounds may be given orally and cause less toxicity (127, 228). Ribavirin, another antiviral drug, may also have anti-*Orthopoxvirus* effects.

### **Selected Open Source Publications by Country**

#### Russia

During Soviet times, Russia carried out intensive studies of *Orthopoxviruses*. For many years, the research into variola genetics was part of a vigorous military program. However, the extent of that work may not be reflected in the open literature. As a result of its long commitment to understanding variola virus genetics, Russia has substantial experience to draw upon in terms of manipulating *Orthopoxviruses*. That represents an important basis of concern for the United States, Canada, Mexico, Europe, Japan, Australia, and other nations in terms of the potential for creation of genetically manipulated *Orthopoxviruses* pathogenic for humans. Such a concern is superimposed upon that relating to the possibility that some Russian variola or variola-like viral stocks may have fallen into the hands of terrorists.

Although the present report focuses on open source publications in the years 2002-2007, we are aware of several earlier Russian papers. Russian scientists reported the complete sequence of a virulent variola virus in 1992 (89). A 1992 publication reported the cloning of the growth factor gene of vaccinia virus (286). Studies from 1993 compared variola and vaccinia viruses (287, 288). Another 1993 paper provided information on a study of genetic engineering of vaccinia virus, which may have been part of a systematic analysis of *Orothopoxvirus* genes (289). A 1996 paper also reported on genetic engineering of Orthopoxviruses by adding genetic material (290). Some of these experiments analyzed virulence mechanisms of the virus (288). Papers from 1995 compared in detail the sequences of a variola virus strain that had caused an outbreak of smallpox in India in 1967 with a variola strain that had caused an outbreak in Bangladesh in 1975 (93, 291). Further studies compared the sequences with vaccinia virus sequences and analyzed their genomic organization (291, 292). Alignment of the DNA genomes showed a 99.3% homology. Of 200 protein sequences, 122 were identical, 42 had a substitution in a single amino acid, 11 had changes in two amino acids, and the other 25 had bigger differences. Such studies can assist in forensic analyses. It also was noted that a region of conservation in variola virus is variable in vaccinia viruses (93).

The extensive work on *Orthopoxviruses* gave Russian scientists insights into the mechanisms by which such viruses elude host immune responses (89, 98, 232, 293). Such information could have provided them with clues to genetic alterations of *Orthopoxviruses* that might make the viruses more virulent to humans. This concern has been magnified by the reported increase in virulence seen after vaccinia virus (20) and then mousepox virus were manipulated so as to carry the gene for mouse interleukin 4 (IL4) (42). Russians have published papers about their genetic manipulations of *Orthopoxviruses*, so far emphasizing changes that make the viruses less pathogenic (294-297); however, some of the same techniques could possibly lead to more virulent cowpox or vaccinia viruses. There is concern that an available *Orthopoxvirus* engineered to carry the human IL4 gene might be especially virulent.

Overall, it appears that as modern molecular genetics techniques have become more sophisticated and disseminated worldwide, whatever advantages the Russians may once have had may have been reduced or even lost. Indeed, studies from Australia and France appear to be as sophisticated as the recent Russian studies or more so. Papers from Brazil may be in the same class as those of the Russians. During the 1990s, Russia experienced a marked reduction in funding for biological research, which led to their failure to develop a modern biotechnology industry and the desertion of many biological scientists. It also has prevented Russia from maintaining and building on its prior gains in viral research. Shortage of funds has led to publications describing experiments that used old-fashioned equipment and a limited supply of reagents. This, along with internal political factors, often leads to publications in Russian journals not readily available to the international scientific community.

Also to be considered are the efforts at collaboration between US and Russian laboratories in recent years. This has included funding of Russian scientists by the US Civilian Research and Development Foundation (219) and the International Science and Technology Center (89), in part in an effort to employ Russian experts who might otherwise take their expertise and skills to other countries. These collaborations have included genetic studies (19, 94, 298). The full sequence of a monkeypox virus (197 kb) was compared with that of a variola strain (~11-12 kb smaller). The central regions, containing genes necessary for survival and replication in a host cell, were 96.3% identical. However, there were considerable differences in the ends of the genomes where species-specific factors are encoded (19). Comparisons of variola virus strains (94) grouped the viruses into three phylogenetic clusters: (i) consisting of three subgroups – China-Korea-Sumatra; Japan; Central Asia-Near East, (ii) an Asian cluster restricted to an India-Nepal group and a Bangladesh group, and (iii) West African and South American stains, perhaps connected by the early slave trade (94).

Russian scientists also have independently published results of sequencing of a number of *Orthopoxviruses* (95-97). Those studies have provided insights into pathogenetic mechanisms of infection with such viruses and the evolution of *Orthopoxviruses* (95, 97). It remains unclear how much of this work was performed recently and how much was carried out many years ago, was not published at the time, and has now been updated and published as new work.

A recent Russian paper (4) stated that "smallpox virus is kept in laboratories of [some or certain] countries." (Brackets and italics added). This wording may hint at Russian knowledge of stocks in countries other than the United States and Russia, but could refer only to the United States and Russia. Another Russian paper states that according to US experts: "At least 10 countries have such collections and this, through accidental leaks (spillages), could possibly endanger the entire world health protection system." (5).

Studies of the ability of different variola strains to infect different classes of host cells indicate that the envelope structure of the virus may contribute importantly to virulence (299).

Russian scientists have developed PCR and microarray assays for the *Orthopoxvirus* genus and for specific *Orthopoxviruses*, such as variola virus (the causative agent of smallpox), monkeypox virus, vaccinia virus, cowpox virus, camelpox virus, and others (95, 97, 110, 215-220, 298, 300-306). Such assays distinguish the varicella-zoster virus (the causative agent of chickenpox and a virus unrelated to *Orthopoxviruses*). This is important because clinical chickenpox may be confused with the clinical presentation of various *Orthopoxviruses*.

Russian scientists at VECTOR created sets of plasmids with each set carrying the complete genome of one of thirteen strains of variola virus (15-17). Such plasmids could potentially be used to reconstitute a viable variola virus or used to create a modified virus with unusual properties. Such plasmids could possibly be used to separately transport non-infectious parts of variola virus to be reassembled after the travel was completed.

In preparation for the plasmid work, it was found that 32 of 54 variola virus isolates were viable and 29 suitable for the relevant long PCR (307).

A phage combinatorial library was screened for scFv single-chain human antibodies reactive with variola, vaccinia, and cowpox viruses (308-317). The purpose was to identify and then grow large quantities of antibodies in case of a new outbreak of *Orthopoxvirus* disease in humans (308, 318). Russian scientists also have produced full-length recombinant antibodies (319).

Between scenario development, computer modeling of the spread of smallpox, new oral vaccination methods, and development of human antibodies specific for variola virus, it appears that Russia is taking extensive measures to deal with a new outbreak of smallpox.

Human tumor necrosis factor (TNF) is an inflammatory, anti-viral cytokine that protects mammalian hosts against many of the adverse consequences of viral infections. TNF is targeted by Orthopoxviruses, including variola virus, as part of the viral strategy of interfering with host anti-viral immune responses (265, 320). Russian scientists used standard molecular genetics techniques to produce recombinant proteins from variola virus, monkeypox virus, and cowpox virus, especially immune modulatory proteins (98, 100, 101, 320). They have analyzed in detail the TNF-binding proteins (98, 101, 265). Such work could provide insights into how *Orthopoxviruses* evade anti-viral immune responses and also yield potential approaches to alter a less pathogenic virus to make it more pathogenic. Russian scientists aligned the sequence of the CrmB protein, the anti-TNF protein of variola virus, with anti-TNF proteins of cowpox viruses and monkeypox viruses and found substantial homology (~90%), but with a number of differences as well, with the lowest homology (85%) between variola virus and monkeypox virus proteins (102, 265). Not reported are results of similar studies with camelpox virus, of interest because of the known strong homology between variola virus and camelpox virus. However, scientists interested in such a comparison could probably perform the analyses from published sequence data. Functional studies of the TNF-binding proteins showed some differences between the variola-derived protein and those from the other

Orthopoxviruses studied (102), with the suggestion of species-specific effects (265). More work would need to be done to understand the genetic and functional differences in detail. Nonetheless, the Russian scientists did find that recombinant variola virus CrmB worked effectively in vitro (321) and decreased the mortality of mice in an endotoxic shock experiment (322). These last studies demonstrate the functional significance of variola proteins that interfere with host immune responses. It also points to a dual use situation in which variola proteins may be used for clinical therapeutic purposes.

Studies related to the TNF protein of *Orthopoxviruses* involved interferon-gammabinding proteins of these viruses, proteins that also facilitate viral virulence by interfering with anti-viral host responses (323). New therapeutic uses of the proteins are being sought (323).

Mousepox, caused by ectromelia virus, was ameliorated by treatment with interferongamma or tumor necrosis factor alpha or both (324). These results may be relevant to treatment of *Orthopoxvirus* infections of humans.

A Russian study showed that ectromelia virus (mousepox) and cowpox virus (also thought to be carried by rodents) differed in terms of their pathological effects on mice. The cowpox virus strain used caused a localized type of infection, whereas ectromelia produced a more disseminated infection (325). However, it is possible that other cowpox virus strains might have different effects. Indeed, sequencing of cowpox virus strains has indicated that cowpox virus has the most complete set of *Orthopoxvirus* genes and that cowpox strains vary to a greater extent than strains from other *Orthopoxviruses* (223). This work points to cowpox viruses as the precursors of variola virus, vaccinia virus, and ectromelia virus because several genes in cowpox virus are found in only fragmentary forms in the other *Orthopoxviruses* (223).

Morphological aspects of various *Orthopoxviruses* were reported with an emphasis on their appearance while growing on chick embryos (326) as well as in various cell types (327).

Russian scientists have reported that variola virus can live for long periods of time in the tissues of corpses, a hundred years or more in permafrost areas or after burial in crypts (5, 108).

Russian scientists, like scientists in many other countries, have used vaccinia virus for several purposes: (i) immunizing against antigens of other pathogens, (ii) anti-cancer treatments, and (iii) gene therapy (65, 66, 67).

Intranasal infection of mice with a cowpox virus strain was studied (328). Intranasal studies have similar implications to aerosol studies. In this circumstance, virus replication in the respiratory tract was found and analyzed.

Aerosol studies were carried out to optimize detection of an airborne release of *Orthopoxviruses*, in this case using vaccinia virus (215, 216). It was found that use of

PCR in the test process markedly increased the sensitivity and decreased the time of analysis.

Russian scientists working at a former BW research center (VECTOR) have published scenarios of potential future outbreaks of smallpox (36). They have emphasized mathematical modeling of the spread of smallpox under various bioterrorism scenarios (36). Requirements for medical personnel, hospital beds, and medical materials; estimates of the rate of spread of the disease; and the need for communications among responders and decision-makers are included. Other scientists at the same Russian institution published mathematical models of the spread through the atmosphere of microbiological agents of concern. These models included dissemination of microbial agents through indoor air (37). Such work has influenced the design of buildings and has led to the modification of ventilation systems (37).

Russian scientists at another former BW research center – Gamaleya Research Institute of Epidemiology and Microbiology – described a scenario involving the release of variola virus (the causative agent of smallpox) in a city of one million people (38). Analyses of this release included mathematical modeling of the spread of disease, with different consequences depending upon the reactions of the public health sector (38).

Russia has involved their Center for Military and Technical Problems of Biological Protection at the Ministry of Defense in the mathematical modeling-computer simulation activity (39). Included in their analyses are releases in (i) closed systems, such as subways, (ii) open air places where people congregate, and (iii) other sites (39).

The genes of Kelch-like proteins from variola virus, monkeypox virus, cowpox virus, and vaccinia virus were compared. These gene sequences were found only in variable terminal regions of the *Orthopoxvirus* genomes (329).

Russian scientists have been evaluating an oral form of live vaccinia virus as a human vaccine for decades. This oral vaccine showed far fewer adverse reactions than did the standard skin scarification method in more than 6,000 human subjects (27, 248, 330, 331). Moreover, protective immune responses after oral vaccination were as good as after scarification, both in terms of antibody titers and cell-mediated immunity. This oral vaccine also showed protection of people from smallpox in Ethiopia in 1972-1973 (27). The Russians believe that oral immunization is much safer than the older scarification method (248, 332) and should be the preferred method in the event of a new outbreak of smallpox (4). It was noted that children playing with ampules of the vaccinia virus could and did become ill (333).

Russian scientists also have developed an oral vaccine against both smallpox and hepatitis B (334-340), apparently based on the oral vaccine with which they had experience (4, 27). Such work is consistent with their encouragement of the simultaneous administration of vaccines against multiple agents, a strategy designed to save money and reduce the number of required medical visits. These studies suggested

that prime and boost with just the oral vaccine may be sufficient to induce a good antismallpox response (334).

A 2005 paper described oral immunization of rabbits with vaccinia virus (341). It would seem that a 35-year experience with oral immunization of people would not be followed by rabbit studies, but that animal experiments would precede human use. It is possible that the reported rabbit studies filled some gap in the human trials. Alternatively, the rabbit studies may have been carried out in the distant past and only recently published.

Russian scientists have also developed a DNA vaccine against variola virus, so far tested in mice (26).

Several studies show that cidofivir and related compounds are effective against *Orthopoxviruses*. Ribavirin appears to have some activity against *Orthopoxviruses*. Combinations of anti-virals may be useful (342). Additional anti-smallpox drugs are under study (343); however, most do not have activity against *Orthopoxviruses* (344).

Many rat monoclonal antibodies to ectromelia virus cross-reacted with certain specific determinants on vaccinia virus, cowpox virus, and smallpox virus (345). Only those antibodies reactive with a 14 kDa polypeptide were cross-neutralizing of *Orthopoxviruses* (345). In addition, some ectromelia virus-specific antibodies were found. Related antibody studies also were reported in another paper (346).

Molecular genetics techniques were used to distinguish vaccine and epidemic strains of sheeppox virus and to distinguish sheeppox and goatpox viruses (259, 347). Other studies established techniques for producing and measuring antibodies to such viruses (347-349).

Genetic variability among strains of monkeypox virus was found by Russian scientists to explain the differences in virulence of the strains (231). The Central African monkeypox virus has virulence factors preventing host immune responses to infection with the virus. The West African strain has a deletion of one such gene (the complement binding protein), preventing production of the gene product that interferes with host-antiviral responses. Without that gene product, the virus is more readily handled by the host, and it causes less severe disease (231). Fortunately, it was the milder form that was imported into the United States in 2003. Had the more virulent form been imported into the United States, much more severe disease might have occurred.

#### China

In contrast to the situation in Russia and the former Soviet Union, which had an intensive research effort into *Orthopoxvirus* genetics, based upon the open source literature, China has not had a large, long-standing laboratory program into *Orthopoxviruses*. However, in recent years Chinese scientists have manipulated such viruses; moreover, its general programs in molecular virology have narrowed the prior gap between itself and Russia.

Chinese scientists have reviewed the world literature on *Orthopoxvirus* genomic organization (350) and have studied individual genes (351, 352). They also have analyzed immune-modulating poxvirus proteins (114, 115), a subject of great interest to Soviet scientists. Chinese scientists have cloned an anti-apoptosis virulence factor of goatpox virus and analyzed it (116). Such an experiment raises the possibility of Chinese scientists inserting various virulence factors into an *Orthopoxvirus* readily able to infect humans. Of special note is the genetic engineering of a human interleukin gene and an interferon gene into *Orthopoxviruses* (75, 76). Although such experiments are of no direct concern, the approach and the techniques could possibly lead to creation by genetic engineering of an *Orthopoxvirus* virulent for humans.

China has published its concern about the potential bioterrorism use of variola virus (9-11, 353). The Chinese expressly indicate that cultures of the variola virus stored in the former states of the Soviet Union could possibly have been commandeered by terrorists, in light of the employment problems after the disintegration of the Soviet Union and the departure of many Soviet scientists (118, 119). Moreover, Chinese scientists see variola virus as easy to grow and aerosolize (353). Chinese papers even discuss creation of an *Orthopoxvirus* that is engineered to carry genes from Ebola virus or from monkeypox virus (353). To protect itself from such a possible event, China has been exploring the use of various viruses as vaccines, including cowpox strains (9), and has analyzed the potency and host range of vaccinia strains (354-356). Chinese scientists also have studied anti-smallpox immunoglobulin (357).

China has used vaccinia virus as a carrier for various antigens in immunization efforts (70, 72-75, 80, 81, 83, 139-162,). It has used vaccinia virus for presenting Nipah virus antigens to the immune system (140, 358). Such studies used altered codons to make them most compatible with mammalian immune systems (74, 79, 140, 150). Chinese scientists also used vaccinia for attempts to immunize against HIV (70-83, 359). In addition to its utility as a vaccine vector, vaccinia virus has been employed as a vector of gene therapy, especially for malignancies (68, 69).

Assays for detection of *Orthopoxviruses* have been developed, including PCR reactions and microarrays (207-214, 360-365). Included are assays to distinguish variola virus from monkeypox virus (360, 361).

Scientists in China have compared the genomes of various *Orthopoxviruses* and have constructed a phylogenetic tree (366).

Chinese scientists have genetically engineered poxviruses. They have studied capripox virus (49, 208, 367-374) and have modified a capripoxvirus (capripoxviruses cause a pox disease in goats and sheep as well as lumpy skin disease in bovines, and they occasionally infect humans) for use in immunization strategies (208, 367-373, 375). The studies included deletion of a virulence gene. In principle, addition rather than deletion of virulence genes also could be performed. In other studies, vaccinia virus was modified to make it safer by removing a gene encoding an interferon-*gamma* binding protein (352).

Of note, capripoxvirus was found to carry a gene encoding a protein able to bind IL18 as did ectromelia virus and vacccinia virus (49). The binding of IL18 increases virulence.

Studies of vaccinia virus have included production of recombinant viruses (355). Although less virulent recombinants may be useful for vaccine strains, the more virulent ones may be problematic in the future. An enhancer element appears to play a role in gene expression (376).

Methods for the large-scale production of smallpox vaccine viruses have been established (377). Such methods could also be used to grow up large numbers of other poxviruses. The method involved growth in 15 liter bottles, rather than in large bioreactors. Survival of vaccine virus was greater at -20 degrees C than at room temperature (377).

Chinese scientists studied the host range of vaccinia virus with the apparent goal of using vaccinia vaccine in horses and other equines (77). However, any studies of host range should be scrutinized as there are concerns that the information might be used to expand the host range of animal poxviruses to humans.

China's interest in mousepox (caused by ectromelia virus) is consistent with the occurrence of that disease in institutional animal facilities, as occurs in the West (378). However, studies of mousepox virus may have application to variola studies.

Chinese scientists have reviewed the non-Chinese research into monkeypox viruses (379-382) and have expressed concern about the possible introduction of monkeypox into China.

One paper suggested that global warming might release variola virus from sites in the tundra or in glaciers so as to induce a new smallpox pandemic (107). Although the variola virus is rather stable in the environment, this suggestion seems unlikely. It remains possible, however, that viable variola virus might be obtained in such a manner and then grown up, which would be a serious concern.

Like scientists in several other countries, scientists in China have established mathematical models of the spread of disease (40). In one smallpox model, 100 infected people would produce 16,530 infected people in 30 days (40).

### Australia

Overall, Australian scientists have produced several interesting and important papers on poxviruses, some of which describe sophisticated scientific methods and thinking.

Perhaps the most important research has to do with IL4 effects on mousepox virus disease. In studies with far-reaching implications, Australian scientists engineered the gene encoding the mouse IL4 gene into mousepox virus (also called ectromelia virus) and discovered that the mousepox virus bearing the IL4 gene had very different properties

than ordinary mousepox virus (41, 42). **The IL4-bearing mousepox virus acted as a super-pathogen**. IL4 expression caused disease in mouse strains normally genetically resistant to mousepox, leading to a high mortality (41, 42). IL4 expression also led to a delay in vaccinia virus clearance from target organs and death of mice infected with vaccinia virus bearing the mouse IL4 gene (20, 41). The physiological basis of these findings is understood. It is known that Th1-type T cells are critical for the cell-mediated immunity and interferon gamma that clear virus-infected cells and that IL4 suppresses the development of such cells (41). Therefore, the marked over-expression of IL4 caused by the modified viruses prevents viral clearance. It was also noted that the antiviral drug cidofovir failed to protect mice from the mousepox virus that bore the mouse IL4 gene (46).

This finding with the IL4-modified mousepox virus has raised the real possibility that a genetically engineered Orthpoxvirus able to infect humans, such as vaccinia virus or cowpox virus, might be made more virulent for humans by similar genetic engineering with the human IL4 gene. In the Australian study, the mouse IL4 gene was incorporated into the mousepox virus and the genetically modified virus administered to laboratory mice. That single genetic alteration (addition of the gene encoding IL4) led to fatal disease in all strains of mice tested, even in mice normally resistant to mousepox. Of perhaps even greater concern, the IL4-bearing mousepox virus overcame standard immunizations that normally protect the mice from mousepox disease. That is, mice vaccinated against mousepox resisted the usual mousepox virus but came down with fatal disease when they were given the IL4-containing mousepox virus (42). The failure of immunization to protect suggests that the IL4-bearing virus interfered with the expression of memory immune responses. This set of results indicates that it is entirely possible that genetic modification of an Orthopoxvirus that causes mild human disease (vaccinia, monkeypox, cowpox, camelpox, etc.) might give rise to an Orthopoxvirus that is as lethal as variola virus. Thus, one might not even need variola virus to cause a serious Orthopoxvirus outbreak – modification of a readily available Orthopoxvirus might serve that purpose. Moreover, such an engineered *Orthopoxvirus* might even be virulent in people immunized against smallpox. Therefore, experiments in which immune mediators are engineered into *Orthopoxviruses* should be viewed with suspicion.

Mouse strains (C57BL6) that produce lots of IFN gamma, IL2, and TNF produce cytotoxic T cells and are relatively resistant to mousepox infection; in contrast, mouse strains that primarily generate an IL4 response (BALB/c and A/J) fail to produce enough cytotoxic T cells (383). The mice that fail to make cytotoxic T cells succumb to the infection, whereas those that do make cytotoxic T cells survive a mousepox infection (383). Some of the interferon effects may be mediated by interferon inducible chemokines (384).

Australian scientists found that antibody is critical for control of secondary poxvirus infections and that interferon is critical in primary infections (385). The ability of poxviruses to produce factors that inhibit interferon function worsens disease.

Australia is concerned about smallpox stocks being in the possession of "rogue states" or bioterrorists, raising the issue of the need for vaccination against smallpox (12).

Live vaccines for smallpox require refrigerated transport and storage (29), suggesting a requirement for alternate approaches. One such alternative might be peptide vaccines; however, they may not provide long-term immunity and might not induce immunity in people with certain MHC types (29). In addition, somewhat toxic adjuvants may be required to induce an adequate immune response with peptide antigens. A lipid-core peptide system that incorporates a lipid adjuvant, carrier, and peptide antigen may overcome these drawbacks (28, 29). Moreover, particles with both immunological adjuvant and carrier functions activate host dendritic cells, which is a critical feature for working very well as immunogens in vaccines (28, 138).

Like scientists in many countries, Australian scientists are developing vaccinia virus as a carrier for other antigens for treating varied infections or cancer (163-168, 386).

Tumor necrosis factor (TNF) production by host cells is critical to recovery from poxvirus infections (387). Studies of mice infected with ectromelia virus (mousepox virus) show that TNF regulates the traffic of immune cells into infected tissues. However, poxviruses produce factors (homologues of receptors for TNF) that bind TNF and, thereby, reduce the effects of host TNF responses (387).

Studies in mice showed that vaccinia virus had developed ways to prevent the action of interferon-inducible T cell alpha chemoattractant, which causes inflammation after other viral infections (388). This factor is induced by interferon-gamma. It is likely that vaccinia-produced proteins that bind to interferon gamma are responsible for this observation.

T cell responses were studied after people were vaccinated with vaccinia virus (389). CD4+, CD38+, CCR5+ T cells and interferon-gamma producing cells peaked at 14 days (389). Neutralizing antibodies increased between days 14 and 21.

Papers from Australia on aerosols of *Orthopoxviruses* – here using vaccinia virus as a surrogate for variola virus – had six authors, all with Russian names (99, 390). This work is a reminder that Russian scientists with expertise in *Orthopoxviruses* may have moved to other countries and continued to work in the field. The studies themselves involved development of personalized samplers for measuring exposure to *Orthopoxviruses* released into the atmosphere.

Like many other countries, Australia has developed an emergency plan to deal with a reintroduction of smallpox (30). Mathematical models of such an event indicate that rapid identification of cases, combined with quarantine and prevention of contact would be effective (30). They suggest that prior mass vaccination would be inefficient (30). However, this conclusion is predicated on a rapid identification of a small number of affected individuals, which may not be possible in the United States.

### New Zealand

New Zealand scientists, like those from other countries, have used vaccinia virus as a vector for other antigens (169).

### Taiwan

Residual immunity against smallpox was measured by assessing T cell reactivity to vaccinia virus (192). The presence of a vaccination scar plus a history of vaccination within 20-30 years correlated with good T cell reactivity.

Aspects of vaccinia virus morphogenesis were studied with an emphasis on temperature sensitivity of various critical proteins (391).

Vaccinia virus entry into host cells was found to involve plasma membrane lipid rafts (392).

Scientists in Tawian have used vaccinia virus to carry other antigens in cancer treatment and in immunizations against other infections (393).

# Japan

Japanese scientists and public health officials are concerned about a possible reemergence of smallpox and have undertaken plans for responses to such an event (394, 395).

Japan's stocks of vaccine for smallpox were destroyed in 1997 because the vaccine had lost its potency (13). A new vaccine was developed because of fears of bioterrorism (13, 14). Nonetheless, some humoral immunity appears to persist even in people last vaccinated nearly 30 years in the past (396, 397), especially in those individuals with at least three vaccinations (31, 398).

A strain of vaccinia virus that had been used as a smallpox vaccine was found to become more virulent after growth in culture due to a mutation in a single gene (283). That gene could be eliminated without loss of protective immunity (283). The resulting strain was as protective as the US Dryvax strain and yet much less virulent. Additional work on the vaccine strain lacking the complicating (B5R) gene has produced what appears to be a much safer vaccine, one that might be sufficiently devoid of serious adverse effects to be considered for large population groups (21-23, 399). This vaccine appears to protect early in infection by inducing cell-mediated immunity more than antibodies (284). British scientists also have been studying B5R effects in a vaccine and believe that the B5 is needed for good immune responses (400).

Mathematical modeling of a potential new outbreak of smallpox suggests that mass vaccination is preferable to vaccination of only case contacts when the Ro (the measure of the number of individuals infected by a single individual with the disease) is high and when public health intervention is delayed (14). Thirty million to fifty-six million doses of vaccine would be needed for such mass vaccination (14, 31).

Japanese scientists reviewed the literature on monkeypox and appeared to be concerned about possible importation of that disease (401-403). More recently, they have carried out experiments in monkeys with the virus obtained from human patients with monkeypox disease in Zaire (Congo Basin virus) and Liberia (West-African virus) (404). They found – as had others previously – that the Congo Basin type of monkeypox is more severe than the West African type of monkeypox (404).

Taterapox virus is closely related to camelpox virus and variola virus. Sequences in the mammalian poxviruses have derived from reptilian sequences (405).

The same vaccine strain of vaccinia virus could give different results in a plaque reduction neutralization assay if different cell types were used for plaquing (406).

Japanese scientists created vaccinia viruses that bore human cytokine genes by genetic engineering (59). The cytokine genes included those encoding IL12, IL23, and IL17 (59). Although none of these constructs are of direct concern, any engineering of interleukin genes into a poxvirus pathogenic for humans is worrisome. These studies further found that anti-IL17 antibodies could make vaccinia virus more virulent in mice (59). IL23 also contributes to resistance to vaccinia virus (59). Therefore, interfering with IL17 and/or IL23 could make an *Orthopoxvirus* infection of humans more severe.

Scientists in many countries have used vaccinia virus as a vector for other antigens or for gene delivery. Japanese scientists also have so employed vaccinia virus (170, 171, 407-413). In addition, Japanese scientists have been developing detection methods for various *Orthopoxviruses* (414).

### South Korea

A South Korean study found persistent humoral (antibody) immunity but not cell-mediated immunity against vaccinia virus in people vaccinated more than 25 years previously (415). For short-term effects, dilution of the vaccine did not impair development of T cell mediated immunity (416). Previously immunized people had higher antibody titers than naïve individuals prior to revaccination, as would be expected, and had fewer adverse events (417).

South Korean scientists found that vaccinia-related kinase 3 suppresses extracellular signal-regulated kinase (ERK) activity through direct binding to a vaccinia H1-related MKP, which inactivates the ERK (418). Indeed, vaccinia regulates the level of

transcription factors (419). An efficient protein expression system was used to express vaccinia-related kinase 1 (420, 421).

Vaccinia virus was used to carry other genes in cancer therapeutics (422). A Yatapoxvirus protein that interferes with host interferon was found to complex with the so-called Z form of DNA (423).

A mathematical model of a smallpox epidemic was developed to evaluate spread in a Korean city and the roles of possible interventions (32).

### **Thailand**

Scientists in Thailand developed mathematical models of the epidemiological outcome of a smallpox outbreak in *Japan* (33). They found that residual immunity in a portion of the population would markedly reduce the magnitude of the crisis (33).

### India

Several outbreaks of buffalopox were reported (424). The genetics of buffalopox virus isolates were studied. These viruses were found to be highly homologous to vaccinia virus (194) but must be distinguished from pseudocowpox, which also occurs (425).

Indian scientists have been analyzing the detailed mechanisms by which the *Orthopoxvirus* complement-binding protein interferes with host complement activity (266, 267).

Scientists in India like those in many other countries have been using vaccinia virus to carry genes for genetic therapy or for immunization against the encoded antigens (426).

### Kazakhstan

A strain of camelpox virus (M-96) was administered to a variety of farm animals and poultry; however, none became infected (257). It is possible that this strain was less virulent than other strains of camelpox virus because previous reports had indicated that three-day-old chicks, guinea pigs, and rabbits were susceptible to camelpox (257).

## **Turkey**

Concerned about biological warfare, Turkey has been developing educational and laboratory plans in case of an emergency (427).

### Israel

Responses to revaccination of Israeli adults were measured (428). Only 56-61% of the people had successful revaccination rates; however, the levels of pre-existing antibodies inversely correlated with the "take." Therefore, a greater percentage may actually be protected. Consistent with that result, the greater the time since last vaccination, the greater the rate of conversion. The optimal time for harvesting vaccinia immune globulin was found to be 14 days after a revaccination (281).

Virus encoded growth factor plays an important role in the *Orthopoxvirus* life cycle (429).

## Lithuania

Public health responses are being planned to counter bioterrorism events, including one with smallpox, (430).

## Czech Republic

Vaccinia virus was found to induce apoptosis in a macrophage cell line through the action of early gene expression (431). That result is superficially at variance with the finding of an apoptosis inhibitor in vaccinia virus (432).

In a collaborative study between Czech and US scientists, it was found that vaccinia virus strains became resistant to the anti-viral drug cidofovir during serial passage in Vero cells in culture (130). Cross-resistance was found for compounds related to cidofovir but not to unrelated nucleosides (130). This result raises the issue of potential purposeful development of a virulent *Orthopoxvirus* pathogenic for humans that might be resistant to the most promising anti-smallpox drugs currently under serious study.

A paper from 1983 indicated that only a few camel herdsmen developed skin lesions from camelpox and that worries about human camelpox are overblown (433).

### Slovakia

The soluble vaccinia virus-produced protein B18R contributes to virulence by binding Type I interferons produced by the host (434).

#### **Poland**

Polish scientists studied certain cellular transcription factors after vaccinia virus infection of human macrophages (435). The experiments provide information on mechanisms of

protein transport in human macrophages during infection with vaccinia virus. The studies are reasonably sophisticated. In somewhat related experiments, Polish scientists studied expression and localization of heat shock proteins after macrophages were infected with vaccinia virus (436). The data suggest that vaccinia viruses use several cellular anti-apoptotic mechanisms to prolong the viability of the macrophages that harbor the virus and, thereby, facilitate viral replication. Studies of host Bcl-2 expression after vaccinia virus infection of human monocytes has shed further light on the anti-apoptosis processes (437).

A vaccinia virus protein able to bind IL1 showed immunosuppressive activity (271). This is one of the immune mechanisms used by *Orthopoxviruses* to resist immune clearance by the host.

It was found that treatment of infected cells with IL1, IL6, and corticosteroids might delay virally induced inhibition of host protein synthesis (438). However, it is not clear whether such approaches would be clinically useful.

## Serbia

Serbia is concerned about possible bioterrorism with smallpox (439).

## Bulgaria

Bulgarian scientists found a synergistic therapeutic benefit of cidofovir and idoxuridine (IUdR) on vaccinia virus replication in cell cultures of chick embryo fibroblasts (440). However, it is unclear if this is applicable to treatment of people because IUdR is toxic to humans.

# Spain

Spanish scientists like those of many other nations have used vaccinia viruses as vectors to deliver antigens or genes (172-179, 441-447). They also have developed detection methods for *Orthopoxviruses* (448).

Recombinant vaccinia viruses were constructed by homologous recombination (449). In addition, Spanish scientists genetically engineered vaccinia virus to carry the genes encoding IL12 and IL18. Experiments with these genetically engineered viruses demonstrated that IL12 and IL18 cytokines contribute to the clearance of the virus (48). This clearance was also facilitated by the involvement of host NK cells and T cells (48). However, it is of concern that cytokines are being engineered into vaccinia viruses and perhaps other *Orthopoxviruses* because some of the cytokines may yield a much more virulent *Orthopoxviruses* as when the mouse IL4 gene was engineered into ectromelia

(mousepox) virus (42). Moreover, interfering with such a cytokine as IL18 might increase virulence.

Scientists in Spain are analyzing events after vaccinia virus infection of human cell lines in an attempt to better understand *Orthopoxvirus* infections, including variola (450). Spanish studies demonstrated that the B1R kinase of vaccinia virus is an early gene required for viral DNA synthesis. The BIR kinase hyper-phosphorylates host p53 molecules which leads to degradation of the p53, contributing to viral control of the host cell machinery (451). They also discovered that a Wiskott-Aldrich protein played a role in vaccinia virus infections (451).

Spanish scientists studied the interactions of vaccinia with host cell membrane proteins (453). Vaccinia virus was found to infect host monocytes most readily, followed by B cells and NK cells (454). Very few T cells were infected.

# Italy

Like scientists from many other countries, Italian scientists have used vaccinia virus to carry antigens for cancer therapy or for immunization against various infectious diseases (455-457).

T cell responses after smallpox vaccination were monitored by flow cytometry (458).

# **Switzerland**

CpG motifs packaged into viral-like particles induced protective cytotoxic T cell responses (459). Vaccinia virus has been used to carry gene segments in cancer therapy (460, 461).

# France

Residual immunity to variola virus may be inadequate to protect the civilian population. In studies of interferon-gamma-producing memory T cells, only 20% of people vaccinated 13-25 years previously had adequate responses (191).

French health officials have been considering potential responses to the threat of smallpox reintroduction. If vaccination with live vaccinia virus of the populace were to be resumed, in addition to the right vaccinia preparation, a good stock of vaccinia immune globulin would be needed (462). Work on such a supply of immune globulin is underway in France. Also, a national network of suitable laboratories for anti-vaccinia immunoglobulin is being established (463). Decontamination methods would be needed as well. Toward this end, a non-corrosive method for decontaminating *Orthopoxvirus*-harboring environmental areas and surfaces has been established (464).

New vaccine production capabilities are being developed (465). Non-replicating vaccinia viruses are being studied for potential utility in humans as smallpox vaccines (466). Such studies are first carried out in a mouse cowpox model (25). Although potentially less likely to cause serious side effects in people, the non-replicating vaccinia strains conferred *less* long-term immunity than classical replicating vaccinia vaccine strains.

French scientists also have been searching for anti-variola drugs. Delivery systems may be critical for the use of such drugs because of the poor *in vivo* conversion of nucleoside analogue drugs to their active triphosphate nucleotide forms (467). In an attempt to overcome this problem, the pro-drugs were encapsulated into a poly(iso-butylcyanoacrylate) aqueous core nanocapsule. However, this did not work as well for these mononucleotides as had been found for oligonucleotides (467). Additional work on formulation will be required. One possible approach is putting the nucleoside analogues, such as cidofovir, into lipids for better delivery (468). Preliminary studies suggest that this may be a useful approach (468). However, issues of drug resistance also have arisen (469).

Detection methods for variola virus and viruses causing diseases resembling smallpox have been developed in France (470, 471).

France, like many other countries, has been using vaccinia viruses to carry other antigens (134-138, 472-474). This ability is based in part on the stimulation of host dendritic cells, which are critical to initiating a strong host immune response (138).

Mathematical models were developed to assist in the management of a potential biological attack with variola virus in France (34). One such model dealt with a smallpox outbreak in a city of two million people (34). Three critical parameters were the basic reproduction number (Ro, the number of cases a single affected individual would cause), time to intervention by the public health authorities, and the proportion of contacts of cases that were traced and vaccinated (34).

French scientists engineered human IL2 into vaccinia virus to study feline fibrosarcomas (47). Although this work is directed at treatment of human cancers (47), such studies are a concern and would be a greater one if an adversary had performed such a study, because certain interleukins have the potential to make poxviruses, including vaccinia virus, monkeypox virus, and variola virus, much more virulent than they otherwise would be.

Using bioinformatics, a French scientist studied the evasion of Toll-like receptors by vaccinia virus by production of a protein that acts on the TLR-IL1 receptor (475). This receptor undergoes dimerization and then complexing to other molecules.

French scientists have studied the *Orthopoxvirus* DNA repair protein called uracil-DNA glycosylase (UNG), which is important in viral pathogensis (476). This enzyme preferentially cuts out uracils when opposite a cytosine (476). Future research will be

directed at developing inhibitors to UNG. In other experiments, French scientists have analyzed the immune response to cowpox virus in mice (477).

# Germany

Codon usage affects the rate of expression of genes, especially genetically engineered genes (478). Low translation initiation and elongation rates are especially associated with insufficient amounts of a corresponding tRNA and to a changed start codon context (478). The nucleotide composition of the mRNA (codon usage) was found to limit gene expression; however, a vaccinia virus polymerase hybrid system improved such expression (478). The results suggested that codon usage may be important in limiting heterologous gene expression in mammalian cells. As a result, modifying the codons of inserted genes prior to their insertion can make them reasonably compatible and more readily expressed.

Aptamers able to bind *Orthopoxviruses* have been developed (479).

German scientists reported that an infection in a colony of 80 monkeys in 2002 with 30 deaths was caused by a poxvirus related to cowpox virus (480). The monkeys had been exposed to rodents that showed evidence of prior poxvirus infection; rodents are known to be a reservoir for cowpox (480).

PCR reactions have been developed to distinguish among the various *Orthopoxviruses* (202, 481) and to diagnose cowpox in a human (203). The assay was 100 times more sensitive than traditional virological culture on Vero cells. German scientists also have developed a PCR assay followed by a DNA melting analysis for detection of *Orthopoxviruses* and of variola virus specifically (204, 205). A RAPD assay has been used to discriminate closely related cowpox viruses (482); however, PCR plus sequencing may be more specific.

A large percentage of cowpox infections of humans have been traced to cats (203, 240), which apparently contract the virus while hunting cowpox-carrying rodents.

A certain major histocompatibility complex type (HLA-A\*201) has been associated with cytotoxic T cell responses to a variola virus epitope (24). This finding may be one factor, along with preprogrammed cytokine production profiles, that underlies differences among people or groups of people in susceptibility to the virus.

German scientists have found that a vaccinia virus gene product associates with host cell membranes, including those of the endoplasmic reticulum, in helping to determine the cytoplasmic sites of viral replication (263, 483).

Studies with a modified vaccinia virus Ankara, typically used to carry genes for immunization or medical treatments, were conducted to maximize efficient production of recombinant viruses (484). In other experiments with the modified vaccinia virus, a

method was developed to rapidly determine neutralizing antibodies in smallpox immunization efforts (485). Indeed, a modified vaccinia virus Ankara has been shown to protect mice from a respiratory challenge with virulent vaccinia virus (24); however, the applicability of that approach to humans and the strength and duration of protection remain to be determined. One paper dealt with the risks of vaccination in people with skin diseases (486).

Like scientists from many other countries, German scientists have used modified vaccinia viruses to deliver genes or to carry genes for antigens from pathogens for immunizations (487-492). Genetic requirements for *in vitro* propagation of this virus have been studied (491, 493-495).

Studies in chickens showed that vaccinia infection induced interferon-beta as well as interferon alpha and gamma (496).

German scientists have tried to develop anti-viral agents that might be effective against a poxvirus infection (497). Unfortunately many of the compounds tested – (i) cyclic and acyclic nucleoside analogues, (ii) a prodrug of cidofovir, (iii) humic acid type phenolic polymers, and (iv) pyrophosphate analogue – have been rather toxic.

It was found that the frequency of secondary transmissions of variola might be highest between three and six days after onset of fever (498). Such information could be used to improve models of smallpox spread and for public health planning.

# United Kingdom

A camelpox protein and vaccinia proteins were found to modulate apoptosis of host cells; they interact with Bcl2 (432, 499, 500). Such viruses also have the ability to modulate host cell actin filaments, which enhances cell-to-cell spread of the virus (501). Moreover, a vaccinia virus virulence factor, N1, has a Bcl-2-like anti-apoptosis action (502).

Sequencing and analysis of camelpox virus led to the discovery that the camelpox virus is most closely related to variola virus, the causative agent of smallpox (90). Both produce the same kind of small white pocks on the chorioallantoic membranes of fertilized hens' eggs and have a similar ceiling temperature for the growth of the pocks. Both viruses are unable to grow in rabbit skin. Comparisons indicate that (i) the genome sequences of the two viruses are closer than either is to any other virus, (ii) the arrangements of the open reading frames (protein coding sequences) are most similar to each other, (iii) the protein sequences themselves are most closely related to each other, and (iv) the nature and structure of the terminal repeats are most similar between variola and camelpox viruses (90). Nonetheless, the number of genetic alterations that would have to be made in camelpox virus to convert it into a serious human pathogen is uncertain.

The camelopox virus encodes a schlafen-like protein that has a mixed affect on virulence (91). This gene is disrupted in variola virus, which could possibly contribute to increased

virulence. Disruption of the camelpox virus gene could possibly increase the virulence of that virus.

British scientists engineered vaccinia viruses to carry Type III (lambda) interferons with no increase in virulence (503).

British scientists studied the immune cells entering the site of a vaccination in a mouse model (504). Viral strains lacking the kelch protein A55 had larger than normal skin lesions (505). They also found that protein B14 is a vaccinia virulence factor (506). Other virulence factors also were studied (52-58). Manipulation of some of these could possibly increase virulence. Of note, inhibition of IL18 was found to increase virulence (56), which could point to various approaches to interfering with IL18 so as to increase virulence.

Vaccinia virus was found to encode an inhibitor of apoptosis, the result of which would be to allow the virus to replicate without host cells dying before viral progeny could be released to continue the infection (507).

Methods, including PCR assays, have been developed for detection of variola virus and *Orthopoxviruses* generally (206).

The variola complement control protein is more potent than the vaccinia complement control protein, which may contribute to the increased virulence of variola virus (272).

The *Orthopoxvirus* protein that binds interferon *gamma* is a dimer in vaccinia, cowpox, and camelpox viruses (273).

Human cowpox infections occur in Europe. Between two and four cases are reported from the UK each year. These infections are commonly contracted from domestic cats that acquire the virus while hunting virus-carrying small rodents (239).

It has been suggested that an allele in European populations may have been under selective pressure in the past because of smallpox (508).

The exit of mature *Orthopoxvirus* particles from host cells has been under consideration (509-511). In addition, viral effects in skin appear to be different from those in other organs (512, 513).

The *Orthopoxvirus* thymidine kinase has been studied in terms of association with other molecules and as a potential target for treatment (514).

Newer antiviral nucleosides are being developed as anti-poxvirus agents (515, 516). A DNA molecule containing immune-stimulatory CpG sequences also may assist in the therapy of *Orthopoxvirus* infections (124).

The adverse reactions of 200 vaccinated health care workers were recorded (517); however, this represents too small a number of people to provide meaningful population information. Modified vaccinia Ankara, a virus that does not replicate in humans, was studied as a possible smallpox vaccine (518). New tissue culture derived smallpox vaccines are being developed (519). British scientists also have been studying the potential use of DNA vaccines against smallpox in a mouse model (520). Such vaccines induce good interferon *gamma* responses but low antibody responses.

British scientists argue that retention of the B5 protein is critical for a smallpox vaccine because that immunogen is what is recognized by important antibodies (285).

Vaccinia immune globulin was tested in a mouse model of disease (521). It was found that the immune globulin might be useful for treating reactions to vaccination with vaccinia virus but would not have a high enough titer to treat smallpox infections.

British scientists have been using modified vaccinia viruses to deliver genes or to carry antigens for immunization against various infections, including malaria and HIV, and in cancer therapy (51, 522-555). They found that deletion of gene A41L increased vaccinia virus immunogenicity (556).

In addition, a vaccinia virus CC-chemokine inhibitor is being developed to treat atherosclerosis (557, 558).

Mathematical models of a potential smallpox outbreak have been developed (559). They emphasize the importance of contact tracing and quarantining; however, these measures are only effective when the outbreak is discovered very early in its evolution.

## Ireland

Mechanisms used by vaccinia virus to interfere with host immune responses were studied (268-270, 275). These included actions on (i) NuclearFactor-*kappaB*, a transcription factor involved in global host immune regulation, (ii) IL10, (iii) TLRs, (iv) IL1, (v) IL18, and (vi) TNF and its receptor, with complex molecular interactions.

### Iceland

Scientists studied the effect of the *Orthopoxvirus* complement-binding protein on atherosclerosis in mice (560).

#### Denmark

In a study done in Guinea-Bissau, Danish scientists found that prior vaccination with vaccinia virus was associated with improved mortality in adults for unknown reasons, but apparently was unrelated to poxvirus infections (561).

Vaccinia virus was found to stimulate dendritic cells, which may play a role in its use as a carrier for various immunogens (190).

## Sweden

Swedish scientists, like those of other nations, have used vaccinia viruses as a vector to deliver antigens or genes (180). They also have developed detection methods for *Orthopoxviruses*.

Swedish scientists have studied complement-binding proteins, including the vaccinia virus complement binding protein. They found that the vaccinia virus complement-binding protein is quite stable and is composed of certain specific structural domains, which may provide the stability (276).

Cowpox infections occur in Sweden (243).

## Norway

A modified vaccinia virus is being developed as a vector for immunization against infectious diseases and for cancer treatment (562).

Norwegian scientists tried to recombine several poxviruses, including cowpox virus (109). If done by an adversary, such experiments would be of some concern.

To better understand the details of vaccinia virus infection of human cells, scientists in Norway have analyzed effects of vaccinia infection of a human cell line (563).

Cowpox infections occur in Norway and Sweden (242, 567).

#### **Finland**

Human cowpox disease currently is contracted primarily from infected domestic cats, who in turn contract the virus from rodents, although dogs also may be implicated (245).

A vaccinia protein with anti-inflammatory properties has been used in graft treatment (564).

### **Netherlands**

Vaccinia virus was used as a vector for tumor-associated antigens in cancer therapy (62) or in immunization against another infection (63, 64, 565, 566).

A generalized infection with cowpox virus was found in an animal in an animal park (237).

Experimental monkeys were infected via the trachea with lethal monkeypox virus. Those monkeys given an acyclic nucleoside phosphonate drug starting 24 hours after infection and continuing for 13 days responded better than monkeys vaccinated with the human vaccinia vaccine given 24 hours after infection (125).

Topoisomerases cut and rejoin one strand of a DNA duplex which allows removal of supercoiled regions of the DNA that can form during replication and/or transcription. The interaction between the vaccinia virus topoisomerase IB enzyme and DNA was studied (567).

Mathematical models of a smallpox outbreak suggested that ring (contact) vaccination can be successful if instituted rapidly (35).

### Austria

A joint Austrian-US effort to produce quantities of smallpox vaccine involved cloning of the old Dryvax vaccinia virus strains and growing up batches in tissue culture (568). Some of the clones were unacceptably virulent in animals. One less virulent strain, found to be as immunogenic as the original Dryvax, was selected for use as the new vaccine and grown in MRC-5 human diploid cells (568). This vaccine induced both cell-mediated immune responses and neutralizing antibody. However, it turned out to be as reactogenic as Dryvax, which is thought by many to be unacceptable for routine immunizations, even if it might be used in a dire emergency. They have also developed a method for the generation of replication – defective vaccinia viruses that could be used as vectors (569). One such modified vaccinia was found to produce long-lasting immunity in mice with the expectation of fewer adverse reactions in humans (61); nonetheless, studies from other countries suggest that replication-defective vaccinia viruses do not produce persistant immunity in humans as do viruses that replicate.

Efforts at filtration of the virus during vaccine preparation were encouraging (570).

Austrian scientists have reviewed the use of vaccinia virus as a vector for a variety of genes (60, 571).

# Belgium

More than 50 years ago, thiosemicarbazones were found to have anti-viral activity, and one of those, methisazone, was investigated in the prophylaxis of smallpox (131). However, with the effective use of vaccinia as a smallpox vaccine, such studies of thiosemicarbazones were abandoned (131).

Cidofovir, used in the therapy of AIDS, also has good activity against *Orthopoxviruses* (572, 573). One newer drug type with activity against *Orthopoxviruses* is an acyclic nucleoside phosphonate [a pyrimidine with a C-2 amino group and a C-6-2 (phosphonomethoxy)ethoxy group or with another phosphonate group] (129). The base consists of a pyrimidine with an amino group at C-2. The inhibition was similar to that of cidofovir, to which some poxviruses are becoming resistant (129). Other nucleoside and nucleotide analogues also have been found to have antiviral activity and could possibly be developed for anti-smallpox purposes (127, 574, 575). Other drugs might target viral enzymes, such as DNA polymerase (131).

Belgian scientists have been interested in the use of poxviruses as vectors for other antigens (576).

A joint British, Belgian and US group found that a Yaba-like disease virus produced an IL10-like protein that could modulate host immune responses after a poxvirus infection (577). Although many cytokine analogues worsen poxvirus diseases, this protein may not. Additional work will be needed to determine its role in naturally occurring poxvirus infections.

Belgian workers analyzed thymidylate kinase from vaccinia virus with the aim of developing new anti-*Orthopoxvirus* drugs directed at that enzyme (126).

Nomadic herders have developed their own vocabulary for dealing with camel diseases, including camelpox (578). Native Taureg populations of Niger can identify *erk eshik* (camelpox) and distinguish it from other disorders of camels.

## Egypt

Egyptian scientists reported an outbreak of buffalopox on a private farm containing 216 buffalo (579). The attack rate was 8%. Disease severity varied from mild to fatal.

A vaccinia virus encoding T7 polymerase was used as an expression vector system (580).

## Saudi Arabia

Sheep and goats were immunized with a sheeppox live vaccine from Romania to counter an outbreak of capripox (581).

## South Africa

Modified vaccinia virus was used by South African scientists as a vector for rabies antigens (582). A vaccinia virus immunosuppressive protein (the complement binding protein) has been studied for its ability to prolong graft survival after transplantation (583). The protein also causes improvement after experimental brain trauma (584) or spinal cord trauma (585). Humanizing the gene encoding that protein makes the engineered protein 100 times more potent (586). South African scientists also have found a potent virulence factor of vaccinia virus that enhances viral growth in the brain (587).

A joint Gambian, Senegal, and British study used the vaccinia virus as a vector for tuberculosis antigens (588). Gambian workers are using vaccinia for carrying malaria antigens for purposes of immunization (589, 590). Studies from Kenya use vaccinia for both malaria and HIV antigens (181).

An Ethiopian study assessed the economics of camels and the diseases affecting them (591).

### Cuba

Cuban scientists have used vaccinia and fowlpox viruses in vaccination strategies against other diseases, including HIV (182, 183, 592).

## Mexico

Mexico has developed a comprehensive national preparedness plan to be used in the event of a bioterrorism event with smallpox (593).

Mexican scientists have used vaccinia virus as a vector for other antigens (184, 594, 595).

### **Brazil**

Faced with naturally occurring outbreaks of *Orthopoxviruses* that infect farm animals and people (195, 250-256, 596-601), Brazil has performed good work (i) on characterizing the viruses as vaccinia virus variants – many now naturally occurring throughout the country (598, 600, 601, 603, 604), (ii) in developing detection methods (250, 251, 255, 256, 600, 601, 605-607), and (iii) in studying aspects of the *Orthopoxvirus* replicative cycle within mammalian host cells (608-617). Brazilian scientists also have used vaccinia virus as a vector for carrying other antigens (185-187).

The genomes of vaccinia and related *Orthopoxviruses* contain genes that encode proteins that trick the host immune system (i) in a non-specific manner, or (ii) by their homology with host soluble factors or host receptors for such factors (598, 599). Among the latter

type are the IL1 beta inhibitor, a serine protease inhibitor that interferes with the enzyme IL1 convertase, and cysteine aspases (caspases) involved in apoptosis (598) as well as a protein that binds to interferon-1 and thereby serves as an inhibitor of interferon actions (599). These processes were studied in recombinant vaccinia viruses (598). In addition, the virus uses the early growth response gene (egr-1) to facilitate replication (616).

Outbreaks of vaccinia virus were found to affect both cattle and people in several regions of Brazil (195, 596, 597, 599, 600). Affected people often had a poxvirus rash (on the face and arms especially), lymphadenopathy, and fever. At least six strains of *Orthopoxviruses* have been isolated around Brazil (599, 604, 618); sometimes more than one strain was found in a given outbreak (195). Based upon analyses of thymidine kinase, vaccinia growth factor (VGF), hemagglutinin (HA), and acidophilic inclusion body genes, these appear to be vaccinia viruses or vaccinia-like viruses from various smallpox vaccination regimens, but which now have become established in natural habitats (598, 600, 601, 604). In addition, unusual variability has been observed (619).

Studies show that vaccinia virus induces extracellular signal-regulated kinases (ERKs) early after infection (609, 612, 614, 620, 621). This induction appears to play a critical role in allowing viral replication within the host cell (612). Mitogen-activated protein kinase (MAPK) signaling pathways MAPK JNK and p38 are activated in host cells late in infection (609, 614, 620). The vaccinia growth factor (VGF) is important in this latter activation (620, 622). Some similar results were found for cowpox virus; however, cowpox and vaccinia viruses recruit different pathways for JNK activation (610, 611, 621, 622).

The early response to growth gene Egr-1 of vaccinia virus appears to encode an important transcription factor impinged upon by signals involved in cell cycle progression, differentiation, and apoptosis (611, 615). Such factors as hormones, neurotransmitters, and growth and differentiation factors stimulate Egr-1 (611).

Scientists in Brazil studied the vaccinia virus F11L protein, which is expressed early in infection but for which no function had been described (608, 613). This gene product was found to interact with the G2 post-replicative transcription elongation factor. Additional studies show that F11L shifts the ratio of extracellular enveloped viral particles and cell-associated enveloped viral particles, which can increase virulence (608).

Lambda interferons were found to have little effect on vaccinia virus (623).

A study of FK506 suggests that the drug may have anti-*Orthopoxvirus* activity targeting the stage of viral DNA replication (624).

# Argentina

Like many countries, Argentina has studied antibody titers to vaccinia virus used in prior vaccination of people against smallpox. A good inverse correlation was found between antibodies and time since last vaccination (625). However, virus neutralizing antibodies were less frequent than antibodies measured by an immunoassay, suggesting that much of the population is not protected against a potential recurrence of smallpox.

Modified vaccinia virus Ankara, which does not replicate in recipient mammalian cells, is being developed for use as a carrier for other genes (188, 189).

In light of the potential for a recurrence of smallpox, scientists studied an old batch of smallpox vaccine called the Malbran strain, produced between 1945 and 1949. Live virus was grown and found to protect mice from cowpox virus challenge (626). The strain was related to the Lister strain. This work shows that a 60-year-old *Orthopoxvirus* is still able to be grown up and used.

### Canada

Tests were conducted to verify the potency of smallpox vaccine stocks (627). The chorioallantoic membrane test and Vero assays for vaccinia virus strains were reestablished for this work. Also, in light of the concerns about bovine spongiform encephalopathy (known as Mad Cow Disease) in North America, vaccine production will proceed without the use of animal proteins in the culture media (132).

A family of poxvirus virulence proteins as studied (628). These proteins were involved in localization of daughter virus production within the cytoplasm of an infected cell and with a role in apoptosis (628).

The *Orthopoxvirus* glutaredoxin gene product was studied by X-ray crystallography (629, 630). In other studies, *Chordopox* virion structure was analyzed (631).

Scientists in Canada, like those in many other countries, have used vaccinia virus as a vector for antigens in vaccine development for other (non-*Orthopoxvirus*) infectious diseases or for gene transfers in the therapy of cancer and other medical conditions (84-88, 632-637).

Infection of human host cells by vaccinia virus causes down-regulation of MHC I molecules on those host cells, which predisposes them to elimination by natural killer (NK) cells (638). The virus also interferes with NK cell functions that might eliminate vaccinia virus. A vaccinia virus F1L protein allows virus-carrying macrophages to survive and viral replication to proceed (639, 640).

A Canadian group developed bioinformatics methods to identify functional genes in *Orthopoxviruses* by identifying purine skewing to predict their A-T rich genes (641). This group also analyzed the G5R protein of vaccinia virus, explored its evolutionary relationships, and found similarities to a human protein (642).

Canadian scientists sequenced a rabbitpox virus genome and compared it with vaccinia virus sequences (196). The work confirms that rabbitpox virus is closely related to vaccinia virus but indicates that rabbitpox virus did not directly evolve from any sequenced vaccinia virus strain (196).

Studies of vaccinia virus associated with a vaccine complication revealed that (i) Dryvax vaccine is not clonal but contains several vaccinia variants, and (ii) the viral lesions in the patient with progressive vaccinia disease appeared to be clonal (643).

In recent years, it has been found that RNA interference (RNAi) can allow mammalian cells to resist viral infection. The vaccinia virus EL3 protein can inhibit RNAi – perhaps via inhibition of interferon-induced protein kinase R (PKR) – thereby reducing this anti-viral effect (644).

*Orthopoxviruses* can recombine within a host cell. Canadian scientists found that the vaccinia virus DNA polymerase had certain properties that allowed such recombination (112, 645).

A Canadian cat was found to be infected by racoonpox virus (646).

The antiviral drug cidofovir was found to inhibit the vaccinia virus DNA polymerase, a mechanism that may be critical to the anti-poxvirus effects of the drug (647). Canadian scientists also have been studying the mechanism of action of another anti-viral, ribavirin (648).

Canadian scientists studied the phenomenon of heterologous reactivation (106). This is a process whereby a cell infected with one poxvirus can produce infectious virus from the pure DNA of a second poxvirus. Such a method could allow resurrection of a no longer extant poxvirus from its known DNA sequence after synthesis of the DNA itself.

### **Discussion of Treatment**

Over the years, the effectiveness of vaccination against smallpox with vaccinia virus reduced the impetus to develop drugs effective against the virus for use either in prophylaxis or in therapy. However, with the eradication of smallpox as a naturally occurring disease and suspension of routine public health vaccination programs, it was recognized that vaccinia virus caused unacceptable adverse side effects in too many recipients to justify its use in large populations against a disease that no longer existed in nature. However, in a desire to be prepared for a possible terrorist attack with variola virus, many countries have tried to develop either less toxic vaccines or new drugs that

might prevent or treat smallpox or a related disease. Such developmental work has covered (i) development of vaccines with fewer adverse events, and (ii) testing of known drugs and new compounds for potential use as anti-*Orthopoxvirus* agents. This type of work is ongoing in several countries and has yielded some promising results.

Vaccination with a standard vaccinia virus preparation causes a mild viral disease and stimulates an immune response that protects against other *Orthopoxviruses*. Many people with atopic dermatitis (a large group, as ~10% of Americans have had eczema) do not produce two antimicrobial peptides: cathelicidinia and beta-detensina in amounts sufficient to prevent serious systemic vaccinia disease (121). This condition is called eczema vaccinatum, which is most often contracted from a family member or other contact who has been vaccinated. Australian scientists have made similar observations (649).

A study of FK506 suggests that the drug may have anti-*Orthopoxvirus* activity, targeting the stage of viral DNA replication (624).

A US team produced monoclonal antibodies to a glycoprotein (A33 glycoprotein) of vaccinia virus and to another antigen (B5 protein) of vaccinia virus (650, 651). The monoclonals also bound to the homologous variola virus antigenic determinants and inhibited the spread of variola *in vitro*. Such monoclonal antibody reagents might be suitable for use in treatment of complications of vaccinations with vaccinia virus or against the spread of smallpox, should there be a new outbreak of that disease, or against monkeypox.

Live vaccines for smallpox require refrigerated transport and storage (29), leading to investigations into alternative approaches. Peptide vaccines have been considered; they may not provide long-term immunity and might not induce immunity in people with certain MHC types (29). In addition, toxic adjuvants may be required to induce an adequate immune response with peptide antigens. A lipid-core-peptide system that incorporates a lipid adjuvant, carrier, and peptide antigen may overcome these drawbacks (28, 29). Moreover, particles with both immunological adjuvant and carrier functions activate host dendritic cells, which is a critical feature for effective immunogens in vaccines (28).

Russian scientists have been evaluating an oral form of live vaccinia virus as a human vaccine for decades. This oral vaccine showed far fewer adverse reactions than the standard skin scarification method and provided protection of people from smallpox in Ethiopia in 1972-1973 (27). Russian scientists also have developed a DNA vaccine against variola virus, so far tested in mice (26).

More than 50 years ago, thiosemicarbazones were found to have anti-smallpox activity (131). However, with the effective use of vaccinia as a smallpox vaccine, studies of thiosemicarbazones were abandoned (131). Cidofovir has good activity against orthopoxviruses. Other drugs might target (i) cellular enzymes, such as IMP dehydrogenase, SAH hydrolase, OMP decarboxylase, and CTP synthetase, and (ii) viral

enzymes, such as DNA polymerase (131). One newer drug type with activity against *Orthopoxviruses* is an acyclic nucleoside phosphonate [a pyrimidine with a C-2 amino group and a C-6-2 (phosphonomethoxy)ethoxy group or with another phosphonate group] (129). The base consists of a pyrimidine with an amino group at C-2. The inhibition was similar to that of cidofovir, to which some *Orthopoxviruses* are becoming resistant (129, 130). Other nucleoside and nucleotide analogues also have been found to have antiviral activity and could possibly be developed for anti-smallpox purposes (127, 128).

Ribavirin appears to have some activity against *Orthopoxviruses*. Combinations of antiviral drugs may be useful (342).

In a collaborative study between Czech and US scientists, it was found that vaccinia virus strains made resistant to cidofovir during serial passage in Vero cells in culture had cross-resistance to compounds related to cidofovir but not to unrelated nucleosides (130). This result raises the issue of potential purposeful development of a virulent *Orthopoxvirus* pathogenic for humans that might be resistant to the most promising anti-smallpox drugs currently under serious study.

In searching for anti-variola drugs, it was found that delivery systems may be critical because of the poor *in vivo* conversion of nucleoside analogue drugs to their active triphosphate nucleotide forms (467). In an attempt to overcome this problem, the prodrugs were encapsulated into a poly(iso-butylcyanoacrylate) aqueous core nanocapsule. However, this did not work as well for these mononucleotides as had been found for oligonucleotides (467). Additional work on formulation will be required.

French and Belgian workers analyzed thymidylate kinase from vaccinia virus with the aim of developing new anti-*Orthopoxvirus* drugs directed at that enzyme (126).

Lethal intra-tracheal infection of monkeys with monkeypox virus was found to respond better to an acyclic nucleoside phosphonate drug started 24 hours after infection and given for a 13-day period, rather than to vaccination with the human vaccine given 24 hours after infection (125).

Unfortunately, some drugs with activity against monkeypox virus and other *Orthopoxviruses* may have limited activity against variola virus (652).

A DNA molecule containing immune-stimulatory CpG sequences also may assist in the therapy of *Orthopoxvirus* infections (124).

Non-replicating vaccinia viruses were studied for utility as potential smallpox vaccines in a mouse cowpox model (25). These conferred less long-term immunity than classical replicating vaccinia vaccine strains. A second generation vaccine would consist of live virus grown in tissue culture (such as human fetal lung fibroblasts) under aseptic conditions (122, 466). Second generation vaccines eliminate concerns about contamination of calf-lymph derived vaccinia with such things as microbes, prions that

might induce Mad Cow Disease, or bovine antigens in the preparation that could cause undesirable immune responses.

A third-generation vaccine would have much enhanced safety and yet retain an ability to induce a long-lasting immune response (466). Although a modified vaccinia virus Ankara has been shown to protect mice from a respiratory challenge with virulent vaccinia virus (24), the applicability of that approach to humans and the strength and duration of protection remain to be demonstrated.

A strain of vaccinia virus that had been used as a smallpox vaccine was found by Japanese scientists to become more virulent after growth in culture due to a mutation in a single gene (283). That gene could be eliminated without loss of protective immunity (283). The resulting strain was as protective as the US Dryvax strain and yet much less virulent. Additional work on the vaccine strain lacking the complicating B5R gene has produced what appears to be a much safer vaccine, one that might be sufficiently devoid of serious adverse effects to be considered for large population groups (21-23). This vaccine appears to protect early in infection by inducing cell-mediated immunity more than antibodies (284). However, British scientists argue that B5 is critical for an immunogen because it is what is recognized by important antibodies (285). Thus, although progress is being made on both new vaccines and new drugs for poxvirus infections, work continues in the search for better pharmaceuticals.

# **Bibliography**

- 1. Daniel Cressey "Not so secure after all" Nature Publishing Group 448: (2007): 732-733.
- 2. K. Alibek "Biohazard" Random House (1999).
- **3.** Charles C.J. Carpenter, Ann M. Arvin, R. Palmer Beasley et al. "Assessment of Future Scientific Needs for Live Variola Virus" *National Academy Press* (1999): 1-118.
- **4.** G.G. Onishchenko, V.A. Maksimov, A.A. Vorob'ev et al. "The topicality of return to smallpox vaccination: problems and prospects" *Bulletin of the Russian Academy of Medical Sciences* 2006 (7): (2006): 32-38.
- **5.** P.I. Ogarkov, K.K. Rayevskiy "Problems Related to Orthopoxvirus Infections Decades after the Global Smallpox Eradication" *Russian Scientific and Practical Conference "Infectious Diseases: Problems of Public Health and Military Medicine"* (2006): 237.
- **6.** Morten Tryland "Should the human smallpox virus (variola) be destroyed?" *Journal of the Norwegian Medical Association* 124 (20): (2004): 2642-2643.
- 7. P.P. Mortimer "Why, which, how, who, when? A personal view of smallpox vaccination for the 2000s" *Communicable Disease and Public Health* 7 (2): (2004): 145-150.
- 8. Michael Gross "Smallpox heads back onto research agenda" Current Biology 15 (12): (2005): R434.
- **9.** Wang Ninglan, Wang Xiujun "Preparation and Development of New Smallpox Vaccines" *Chinese Journal of Frontier Health and Quarantine* 25 (z1): (2002): 92-93.
- **10.** Huang Jianshi "Epidemiological Characteristics of Bioterrorism-related Diseases" *Chinese Journal of Preventive Medicine* 38 (2): (2004): 78-80.
- **11.** Wang Luyu, Zhao Cui "Functions of Vector Animals in Bioterrorism and Precautionary Strategies" *Chinese Journal of Vector Biology and Control* 15 (3): (2004): 238-240.
- **12.** M.J. Selgelid "Bioterrorism and smallpox planning: information and voluntary vaccination" *Journal of Medical Ethics* 30 (6): (2004): 558-560.
- **13.** Kiyoshi Horiuchi "Topics in Preventive Inoculation: Development and Administration of New Smallpox Vaccine Programs" *Japanese Journal of Pediatric Medicine* 36 (3): (2004): 493-499.
- **14.** Yasushi Ohkusa, Kiyosu Taniguchi, Ichiro Okubo "Prediction of smallpox outbreak and evaluation of control-measure policy in Japan, using a mathematical model" *Journal of Infection and Chemotherapy* 11 (2): (2005): 71-80.
- **15.** I.N. Babkina, P.F. Safronov, I.V. Babkin et al. "Construction of DNA-fragments' libraries with complete genomes of different variola strains" *Problems of Virology* 50 (2): (2005): 18-23.
- **16.** Ilya G. Drozdov, A.N. Sergeev, S.N. Shchelkunov et al. "An update on variola virus stocks. Conservation of genetic material of different variola virus strains from the Russian collection at the WHO Collaborating Centre for Orthopoxvirus Diagnosis and Repository for Variola Virus Strains and DNA" *Meetings of the WHO Advisory Committee on Variola Virus* (2006).
- 17. I.N. Babkina, Sergei N. Shchelkunov, Lev S. Sandakhchiev "Conservation of genetic material of various variola virus strains from the Russian Collection" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- 18. G.L. Smith, G. McFadden "Smallpox: anything to declare?" Nature Reviews Immunology 2 (7): (2002): 521-527.
- **19.** Sergei N. Shchelkunov, Alexei V. Totmenin, Igor V. Babkin et al. "Human Monkeypox and Smallpox Viruses: Genomic Comparison" *FEBS Letters* 509: (2001): 66-70.
- 20. Martin Enserink, Richard Stone "Dead Virus Walking" Science 295 (2002): 2001-2005.

- **21.** Minoru Kidokoro, Masato Tashiro, Hisatoshi Shida "Genetically stable and fully effective smallpox vaccine strain constructed from highly attenuated vaccinia LC16m8" *Proceedings of the National Academy of Sciences of the United States of America* 102 (11): (2005): 4152-4157.
- **22.** Minoru Kidokoro, Masayuki Saijo, Yasushi Ami et al. "Protective Effect of Improved Smallpox Vaccine Strain m8-Delta Against Monkeypox Infection in Cynomolgus Monkeys" *Program and Abstracts from the Annual Meeting of the Japanese Society for Virology* 54: (2006): 394.
- **23.** Masayuki Saijo, Yasushi Ami, Yuriko Suzaki et al. "LC16m8, a Highly Attenuated Vaccinia Virus Vaccine Lacking Expression of the Membrane Protein B5R, Protects Monkeys from Monkeypox" *Journal of Virology* 80 (11): (2006): 5179-5188.
- **24.** I. Drexler, C. Staib, W. Kastenmueller et al. "Identification of vaccinia virus epitope-specific HLA-A\*0201-restricted T cells and comparative analysis of smallpox vaccines" *Proceedings of the National Academy of Sciences of the United States of America* 100 (1): (2003): 217-222.
- 25. Robert Drillien, Audrey Ferrier-Rembert "Smallpox vaccines: pathogenicity-safety, humoral and cellular immunogenicity, protection" *Thesa Database* (2006).
- **26.** R.A. Maksyutov, I.N. Babkina, A.E. Nesterov et al. "Construction of a candidate DNA vaccine against human orthopoxvirus infections" *Russian Journal of Biotechnology* 2006 (4): (2006): 23-30.
- **27.** A.A. Vorobyev, V.N. Podkuyko, V.A. Maksimov "Oral vaccination against smallpox (on the return of smallpox vaccination)" *Bulletin of the Russian Academy of Medical Sciences* 2003 (1): (2003): 5-10.
- **28.** Anita Gamvrellis, David Leong, Jennifer C. Hanley et al. "Vaccines that facilitate antigen entry into dendritic cells" *Immunology and Cell Biology* 82 (5): (2004): 506-516.
- **29.** Peter M. Moyle, Aniko Horvath, Colleen Olive et al. "Development of lipid-core-peptide (LCP) based vaccines for the prevention of group A streptococcal (GAS) infection" *International Journal of Peptide Research and Therapeutics* 10 (5-6): (2004): 605-613.
- **30.** G.K. Aldis, M.G. Roberts "An integral equation model for the control of a smallpox outbreak" *Mathematical Biosciences* 195 (1): (2005): 1-22.
- **31.** Yasue Takeuchi, Masaru Hara, Kosaku Aoyama "Anti-Smallpox Antibody Retention 28 Years Following the Cessation of Smallpox Vaccinations" *Modern Media* 52 (2): (2006): 50-53.
- **32.** Ch'o'n Byo'ng-ch'o'l "Epidemic Modeling and Table-top Exercise for Emerging Infectious Diseases in Korea" *Korean Journal of Epidemiology* 28 (1): (2006): 47-63.
- **33.** Nishiura Hiroshi, Tang I. Ming "Modeling for a smallpox-vaccination policy against possible bioterrorism in Japan: the impact of long-lasting vaccinal immunity" *Journal of Epidemiology* 14 (2): (2004): 41-50.
- **34.** J. Legrand, C. Viboud, P.Y. Boelle et al. "Modelling responses to a smallpox epidemic taking into account uncertainty" *Epidemiology and Infection* 132 (1): (2004): 19-25.
- **35.** Mirjam Kretzschmar, Susan van den Hof, Jacco Wallinga et al. "Ring vaccination and smallpox control" *Emerging Infectious Diseases* 10 (5): (2004): 832-841.
- **36.** Aleksandr Grigoryevich Bachinskiy, I. Fominykh "Analysis of Scenarios of Development of Outbreaks as Applied to Smallpox" *International Conference "Development of International Collaboration in Infectious Disease Research"* 2004 (1): (2004).
- **37.** B.M. Desyatkov, Aleksandr Ivanovich Borodulin, S.R. Sarmanayev "Use of Mathematical Modeling Methods of Atmospheric Admixture Spreading for Solving Applied Medicobiological Problems" *International Conference* "Development of International Collaboration in Infectious Disease Research" (2004).
- **38.** A.A. Vorob'ev, B.V. Boev, V.M. Bondarenko et al. "Problem of bioterrorism under modern conditions" *Journal of Microbiology, Epidemiology and Immunobiology* 2002 (3): (2002): 3-12.

- **39.** V.P. Myakishev, V.V. Filonov "Prediction of the Consequences of Bioterrorist Acts as the Basis for Organizing Counter-Measures to the Artificial Spread of Particularly Dangerous Infections" *Russian Congress of Infectious Disease Specialists* (2003).
- **40.** Feng Dan, Cao Wuchun "Establishment of Injury Effect Model for Aerosol Release of Biological Warfare Agents" *CNKI China Doctor/Master Dissertations Full-text Database* (2004).
- **41.** Dharam P. Sharma, Alistair J. Ramsay, Deborah J. Maguire et al. "Interleukin-4 Mediates Down Regulation of Antiviral Cytokine Expression and Cytotoxic T-Lymphocyte Responses and Exacerbates Vaccinia Virus Infection In Vivo" *Journal of Virology* 70 (10): (1996): 7103-7107.
- **42.** Ronald J. Jackson, Alistair J. Ramsay, Carina D. Christensen et al. "Expression of Mouse Interleukin-4 by a Recombinant Ectromelia Virus Suppresses Cytolytic Lymphocyte Responses and Overcomes Genetic Resistance to Mousepox" *Journal of Virology* 75 (3): (2001): 1205-1210.
- **43.** Dominikus Bonsch, Winfried Kammer, Antje Lischke et al. "Species-specific Agonist/Antagonist Activities of Human Interleukin-4 Variants Suggest Distinct Ligand Binding Properties of Human and Murine Common Receptor y Chain" *The Journal of Biological Chemistry* 270 (15): (1995): 8452-8457.
- **44.** Briggs W. Morrison, Philip Leder "A Receptor Binding Domain of Mouse Interleukin-4 Defined by a Solid-phase Binding Assay and in Vitro Mutagenesis" *Journal of Biological Chemistry* 267 (17): (1992): 11957-11963.
- **45.** Gabrielle T. Belz, Ken Shortman, Michael J. Bevan et al. "CD8?+ Dendritic Cells Selectively Present MHC Class I-Restricted Noncytolytic Viral and Intracellular Bacterial Antigens In Vivo" *Journal of Immunology* 175 (1): (2005): 196-200.
- **46.** Samantha J. Robbins, Ronald J. Jackson, Frank Fenner et al. "The efficacy of cidofovir treatment of mice infected with ectromelia (mousepox) virus encoding interleukin-4" *Antiviral Research* 66 (1): (2005): 1-7.
- **47.** T.M. Jourdier, C. Moste, M.C. Bonnet et al. "Local immunotherapy of spontaneous feline fibrosarcomas using recombinant poxviruses expressing interleukin 2 (IL2)" *Gene Therapy* 10 (26): (2003): 2126-2132.
- **48.** M. Magdalena Gherardi, Juan C. Ramirez, Mariano Esteban "IL-12 and IL-18 act in synergy to clear vaccinia virus infection: Involvement of innate and adaptive components of the immune system" *Journal of General Virology* 84 (8): (2003): 1961-1972.
- **49.** Liu Wenqiang, Lu Jianbiao "Capripoxvirus Genome Encoding IL-18-binding Protein Gene" *Shandong Science* 19 (6): (2006): 53-55.
- **50.** Vincent P. Smith, Neil A. Bryant, Antonio Alcami "Ectromelia, vaccinia and cowpox viruses encode secreted interleukin-18-binding proteins" *Journal of General Virology* 81 (5): (2000): 1223-1230.
- **51.** Natasha Larke, Aileen Murphy, Christoph Wirblich et al. "Induction of human immunodeficiency virus type 1-specific T cells by a bluetongue virus tubule-vectored vaccine prime-recombinant modified virus Ankara boost regimen" *Journal of Virology* 79 (23): (2005): 14822-14833.
- **52.** Aylwin Ng, David C. Tscharke "The vaccinia virus A41L protein is a soluble 30 kDa glycoprotein that affects virus virulence" *Journal of General Virology* 82 (9): (2001): 2095-2105.
- **53.** Elizabeth C. Mathew, Christopher M. Sanderson, Ruth Hollinshead et al. "A mutational analysis of the vaccinia virus B5R protein" *Journal of General Virology* 82 (5): (2001): 1199-1213.
- **54.** Nathan Bartlett, Julian A. Symons, David C. Tscharke et al. "The vaccinia virus N1L protein is an intracellular homodimer that promotes virulence" *Journal of General Virology* 83 (2002): (2002): 1965-1976.
- **55.** Timothy P. Newsome, Niki Scaplehorn, Michael Way "Src Mediates a Switch from Microtubule- to Actin-Based Motility of Vaccinia Virus" *Science* 306 (5693): (2004): 124-129.
- **56.** Julian A. Symonsa, Elizabeth Adams, David C. Tscharke et al. "The vaccinia virus C12L protein inhibits mouse IL-18 and promotes virus virulence in the murine intranasal model" *Journal of General Virology* 83 (2002): (2002): 2833-2844.

- **57.** Ferran Valderrama, Joao V. Cordeiro, Sibylle Schleich et al. "Vaccinia virus-induced cell motility requires F11L-mediated inhibition of RhoA signaling" *Science* 311 (5759): (2006): 377-381.
- **58.** J.A. Symons, D.C. Tscharke, N. Price et al. "A study of the vaccinia virus interferon-gamma receptor and its contribution to virus virulence" *Journal of General Virology* 83 (2002): (2002): 1953-1964.
- **59.** Shunsuke Kohyama, Satoshi Ohno, Akihiro Isoda et al. "IL-23 Enhances Host Defense against Vaccinia Virus Infection Via a Mechanism Partly Involving IL-17" *Journal of Immunology* 179 (6): (2007): 3917-3925.
- 60. F.G. Falkner, G.W. Holzer "Vaccinia viral/retroviral chimeric vectors" Current Gene Therapy 4 (4): (2004): 417-426
- **61.** J. Mayrhofer, M. Gerencer, F.G. Falkner et al. "The nonreplicating smallpox candidate vaccines defective vaccinia Lister (dVV-L) and modified vaccinia Ankara (MVA) elicit robust long-term protection" *Virology* 341 (1): (2005): 91-101
- **62.** Hetty J. Bontkes, Janneke J. Ruizendaal, Marco W.J. Schreurs et al. "Antigen gene transfer to human plasmacytoid dendritic cells using recombinant adenovirus and vaccinia virus vectors" *Cellular Oncology* 27 (3): (2005): 175-182.
- **63.** Gerrit Koopman, Daniella Mortier, Sam Hofman et al. "Vaccine protection from CD4+ T-cell loss caused by simian immunodeficiency virus (SIV) mac251 is afforded by sequential immunization with three unrelated vaccine vectors encoding multiple SIV antigens" *Journal of General Virology* 85 (10): (2004): 2915-2924.
- **64.** Leon de Waal, Linda S. Wyatt, Selma Yuksel et al. "Vaccination of infant macaques with a recombinant modified vaccinia virus Ankara expressing the respiratory syncytial virus F and G genes does not predispose for immunopathology" *Vaccine* 22 (8): (2004): 923-926.
- **65.** M.V. Khoretonenko, M.F. Vorovitch, L.G. Zakharova et al. "Vaccinia virus recombinant expressing gene of tickborn encephalitis virus non-structural NS1 protein elicits protective activity in mice" *Immunology Letters* 90 (2): (2003): 161-163.
- **66.** S.E. Aleshin, A.V. Timofeev, M.V. Khoretonenko et al. "Combined prime-boost vaccination against tick-borne encephalitis (TBE) using a recombinant vaccinia virus and a bacterial plasmid both expressing TBE virus non-structural NS1 protein" *BMC Microbiology* 5 (1): (2005): 45.
- **67.** A.Ye. Nesterov, I.A. Ryazankin, Ye.V. Bessmeltseva et al. "Design of Vaccinia virus recombinant strain producing a chimeric TBI-HBsAg protein" *Russian Journal of Biotechnology* 2004 (1): (2004): 3-12.
- **68.** Wu Ting, Ou Shanhai, Chen Min et al. "The infection efficiencies and EGFP expression levels of recombinant vaccinia virus in different mammalian cell lines" *Journal of Xiamen University (Natural Science)* 44 (6): (2005): 866-869.
- **69.** Li Jinzhou, Qi Yanchao, Luo Chaoquan "Preventive and Therapeutic Effect of CEA Recombinant Vaccinia Virus on CEA Positive Tumor" *Journal of Zhengzhou University (Medical Sciences)* 40 (4): (2005): 667-669.
- **70.** Teng Honggang, Jiang Chunlai, Yu Xianghui et al. "Construction and Immunogenicity of Recombinant Modified Vaccinia Ankara Expressing HIV-1 Gag-Po1 Antigen" *Chinese Journal of Biologicals* 20 (1): (2007): 25-28.
- **71.** Liu Ying, Duan Danli, Peng Hong et al. "Immunogenicity analysis of HIV vaccine based on replicating vaccinia virus Tiantan vector" *Chinese Journal of Experimental and Clinical Virology* 18 (3): (2004): 281-283.
- **72.** Xin Wei, Guo Fei, Lu Roujian et al. "Expression of nef gene of HIV-1 subtype C bostwana in non-replicative vaccinia virus and its immune effect in mice" *Chinese Journal of Virology* 20 (2): (2004): 97-103.
- 73. Zhou Xiaoyun, Liu Ying, Rao Liqun et al. "Construction of vaccinia virus vector for vaccine screening" *Chinese Journal of Virology* 20 (4): (2004): 315-321.
- **74.** Zhang Xiangmin, Xin Wei, Wang Shifeng et al. "Optimizing the codon usage of HIV-1 gag gene according to the codon bias of vaccinia virus improves the gag gene expression" *Chinese Journal of Virology* 21 (3): (2005): 210-216.
- **75.** Wang Yuhong, Li Tiyuan, Liu Suhuan et al. "Coexpression of HIV-1 ENV and IL-6 Gene in Recombinant Vaccinia Virus" *Chinese Journal of Zoonoses* 16 (4): (2000): 12-14.

- **76.** Wang Hongjun, Jin Ningyi, Nian Chunzhi et al. "Lymphocyte transformation and CTL induced with expression of HIV-1 env/INF?-2b" *Chinese Journal of Zoonoses* 20 (2): (2004): 85-87.
- 77. Ma Yan, Meng Qinglai, Duan Danli et al. "Study on the infectivity of vaccinia virus in equine cells" *Chinese Journal of Zoonoses* 23 (2): (2007): 149-152.
- **78.** Yao Xiangjie, Guo Jia, Zheng Congyi et al. "Establishment of an in vitro cell culture system transfected by full-length HCV cDNA genome" *Chinese Science Bulletin* 49 (13): (2004): 1358-1363.
- **79.** Li Haishan, Liu Yong, Li Dingfeng et al. "Enhancement of DNA vaccine-induced immune responses by a 72-bp element from SV40 enhancer" *Chinese Medical Journal* 120 (6): (2007): 496-502.
- **80.** Zhou Xiaoyun, Shao Yiming, Rao Liqun et al. "Construction of recombinant vaccinia virus expressing HIV-1 polyvalent synthetic gene for vaccine" *Journal of Microbiology* 25 (5): (2005): 1-5.
- **81.** Dai Kaifan, Liu Ying, Zhou Xiaoyun et al. "Construction of a Recombinant Vaccinia Virus Expressing the gagpol Gene of HIV-I Chinese Epidemic Recombinant Strain CN54, Clade B'/C" *Virologica Sinica* 18 (5): (2003): 441-445.
- **82.** Xu Jianqing, Ren Li, Huang Xianggang et al. "Sequential priming and boosting with heterologous HIV immunogens predominantly stimulated T cell immunity against conserved epitopes" *AIDS* 20 (18): (2006): 2293-2303.
- **83.** Huang Xianggang, Xu Jianqing, Qiu Chao et al. "Mucosal priming with PEI/DNA complex and systemic boosting with recombinant TianTan vaccinia stimulate vigorous mucosal and systemic immune responses" *Vaccine* 25 (14): (2007): 2620-2629.
- **84.** Zhou Xing, Michael Santosuosso, Sarah McCormick et al. "Recent advances in the development of adenovirus- and poxvirus-vectored tuberculosis vaccines" *Current Gene Therapy* 5 (5): (2005): 485-492.
- **85.** Gilbert Philippe-Alexandre, Grant McFadden "Poxvirus cancer therapy" *Recent Patents on Anti-Infective Drug Discovery* 1 (3): (2006): 309-321.
- **86.** R. Mouchantaf, H.L. Watt, T. Sulea et al. "Prosomatostatin is proteolytically processed at the amino terminal segment by subtilase SKI-1" *Regulatory Peptides* 120 (1-3): (2004): 133-140.
- **87.** Yu Hong, Huang Hui, Xiang Jim et al. "Dendritic cells pulsed with hepatitis C virus NS3 protein induce immune responses and protection from infection with recombinant vaccinia virus expressing NS3" *Journal of General Virology* 87 (6): (2006): 1761.
- **88.** Markus Czub, Hana Weingartl, Stefanie Czub et al. "Evaluation of modified vaccinia virus Ankara based recombinant SARS vaccine in ferrets" *Vaccine* 23 (17-18): (2005): 2273-2279.
- **89.** S.N. Shchelkunov "Results of the Study of the Variola Virus Genome" *Russian Journal "AIDS, Cancer and Public Health"* 10 (3): (2006): 33-41.
- **90.** Caroline Gubser, Geoffrey L. Smith "The sequence of camelpox virus shows it is most closely related to variola virus, the cause of smallpox" *Journal of General Virology* 83: (2002): 855-872.
- **91.** Caroline Gubser, Rory Goodbody, Andrea Ecker et al. "Camelpox virus encodes a schlafen-like protein that affects orthopoxvirus virulence" *Journal of General Virology* 88 (6): (2007): 1667-1676.
- **92.** Sergei N. Shchelkunov, I.V. Babkin, T.S. Nepomnyashikh "Sequencing and computer-assisted analysis of variola virus DNA at the WHO Collaborating Centre for Orthopoxvirus Diagnosis and Repository for Variola Virus Strains and DNA" *Meetings of the WHO Advisory Committee on Variola Virus* (2006).
- **93.** S.N. Shchelkunov, R.F. Massung, J.J. Esposito "Comparison of the genome DNA sequences of Bangladesh-1975 and India-1967 variola viruses" *Virus Research* 36 (1): (1995): 107-118.
- **94.** I.N. Babkina, I.V. Babkin, U. Le et al. "Phylogenetic comparison of the genomes of different strains of variola virus" *Doklady Biochemistry and Biophysics* 398 (2004): 316-319.

- **95.** Lev Stepanovich Sandakhchiyev, Sergey Nikolayevich Shchelkunov "Molecular Genetic Studies of Orthopoxviruses Pathogenic for Humans" *International Conference "Development of International Collaboration in Infectious Disease Research"* (2004).
- **96.** Aleksey Vladimirovich Totmenin "Study of the Structural and Functional Organization of the Variola Virus Genome" *Federal Portal for Science and Innovation* (2004).
- **97.** S.N. Shchelkunov "Role of Basic Research on Orthopoxviruses in the Development of Their Diagnostic Techniques and Therapy for Human Immunopathologies" *Basic Science for Medicine* (2006).
- **98.** I.P. Gileva, T.S. Nepomnyashchikh, A.A. Grazhdantseva et al. "Comparative Study of the Properties of TNF-Binding Orthopoxviral Proteins and Their Chimeras With an IgG Heavy Chain Fragment" *Basic Science for Medicine* (2007): 114.
- **99.** I.E. Agranovski, A.S. Safatov, O.V. Pyankov et al. "Long-Term Sampling of Viable Airborne Viruses" *Aerosol science and technology* 39 (9): (2005): 912-918.
- **100.** T.S. Nepomnyashchih, L.R. Lebedev, G.V. Kochneva et al. "Orthopoxvirus Tumor Necrosis Factor binding proteins and Gamma-Interferon binding proteins: expression, purification and some characteristics" *The FEBS Journal* 272 (Supplement 1): (2005): 499.
- **101.** Tatyana Sergeyevna Nepomnyashchikh, Irina Pavlovna Gileva "Study of the Properties of Gamma-Interferon- and TNF-Binding Proteins of Orthopoxviruses" *Vector News. Archive. Defense of Dissertations* (2006).
- **102.** Irina P. Gileva, Tatiana S. Nepomnyashchikh, Denis V. Antonets et al. "Properties of the recombinant TNF-binding proteins from variola, monkeypox, and cowpox viruses are different" *Biochimica et Biophysica Acta (BBA) Proteins & Proteomics* 1764 (11): (2006): 1710-1718.
- **103.** Frank Fenner, G.M. Woodroofe "The reactivation of poxviruses. II. The range of reactivating viruses" *Virology* 11: (1960): 185–201.
- **104.** H. Hanafusa, T. Hanafusa, J. Kamahora "Transformation phenomena in the pox group of viruses. II. Transformation between several members of pox group" *Biken Journal* 2: (1959): 85-91.
- **105.** Irina Nikolayevna Babkina, S.N. Shchelkunov, O.V. Morozova et al. "Comparative Analysis and Conservation of Variola Virus Genomes from the Russian Collection" *Vector News. Archive. Defense of Dissertations* (2006): 1-20.
- **106.** Xiao-Dan Yao, David H. Evans "Construction of recombinant vaccinia viruses using leporipoxyvirus- catalyzed recombination and reactivation of orthopoxvirus DNA" *Methods in Molecular Biology* 269: (2004): 51-64.
- **107.** Zhang Jinbao "Greenhouse Effect and the Resuscitation of Variola Virus" *Journal of Liaocheng Teachers University (Natural Sciences)* 15 (1): (2002): 50-51.
- **108.** Marion E. Andrew, Barbara E.H. Coupar "Biological effects of recombinant vaccinia virus-expressed interleukin 4" *Cytokine* 4 (4): (1992): 281-286.
- **109.** Hansen Hilde, Okeke Malachy Ifeanyi, Nilssen Oivind et al. "Recombinant viruses obtained from co-infection in vitro with a live vaccinia-vectored influenza vaccine and a naturally occurring cowpox virus display different plaque phenotypes and loss of the transgene" *Vaccine* 23 (4): (2004): 499.
- **110.** Majid Laassri, Vladimir Chizhikov, Maxim Mikheev et al. "Detection and discrimination of orthopoxviruses using microarrays of immobilized oligonucleotides" *Journal of Virological Methods* 112 (1): (2003): 67-78.
- 111. K. Matz-Rensing, H. Ellerbrok, B. Ehlers et al. "Fatal Poxvirus Outbreak in a Colony of New World Monkeys" *Veterinary Pathology* 43: (2006): 212-218.
- **112.** Michael D. Hamilton, Anthony A. Nuara, Don B. Gammon et al. "Duplex strand joining reactions catalyzed by vaccinia virus DNA polymerase" *Nucleic Acids Research* 35 (1): (2007): 143-151.
- **113.** Martin Eichner "Analysis of historical data suggests long-lasting protective effects of smallpox vaccination" *American Journal of Epidemiology* 158 (8): (2003): 717-723.

- **114.** Liu Wenqiang, Yang Shaohua, Luan Jingjing et al. "Expression and bioactivity of IL18-binding protein gene from sheep poxvirus" *Chinese Journal of Virology* 21 (4): (2005): 305-307.
- 115. Zheng Min "Immunomodulatory strategies of poxviruses" Infectious Disease Information 20 (2): (2007): 83-86.
- **116.** Guan Huimin, Wang Jiafu, Liu Jianhua "Cloning and identification of antiapoptosis virulence factor gene of goatpox virus in Guizhou" *CNKI China Doctor/Master Dissertations Full-text Database* (2007).
- 117. Chen Juxiang, Xu Jian, Ying Kang et al. "Molecular cloning and characterization of a novel human BTB domain-containing gene, BTBD10, which is down-regulated in glioma" *Gene* 340 (1): (2004): 61-69.
- **118.** Donald A. Herson "The looming threat of bioterrorism" *The American Association for the Advancement of Science* 283 (5406): (1999): 1279-1282.
- **119.** Joel G. Breman, Isao Arita, Frank Fenner "Preventing the return of smallpox" *The New England Journal of Medicine* 348: (5): (2003): 463-466.
- **120.** Mirjam Kretzschmar, Jacco Wallinga, Peter Teunis et al. "Frequency of adverse events after vaccination with dtifferent vaccinia strains" *PLoS Medicine* 3 (8): (2006): 1341-1351.
- **121.** Benjamin Schwartz, Mark Lebwohl "Complications of the smallpox vaccine" *International Journal of Dermatology* 44 (4): (2005): 289-292.
- **122.** Gregory A. Poland "Smallpox vaccines: from first to second to third generation" *The Lancet* 365 (9457): (2005): 362-363.
- **123.** Emma Kerrod, Alasdair M. Geddes, Martyn Regan et al. "Surveillance and control measures during smallpox outbreaks" *Emerging Infectious Diseases* 11 (2): (2005): 291-297.
- **124.** D.G. Cerys Rees, Amanda J. Gates, Michael Green et al. "CpG-DNA protects against a lethal orthopoxvirus infection in a murine model" *Antiviral Research* 65 (2): (2005): 87-95.
- **125.** Koert J. Stittelaar, Johan Neyts, Lieve Naesens et al. "Antiviral treatment is more effective than smallpox vaccination upon lethal monkeypox virus infection" *Nature* 439 (7077): (2006): 745-748.
- **126.** Dimitri Topalis, Bruno Collinet, Cecile Gasse et al. "Substrate specificity of vaccinia virus thymidylate kinase" *The FEBS Journal* 272 (24): (2005): 6254-6265.
- **127.** Erik De Clercq, Johan Neyts "Therapeutic potential of nucleoside/nucleotide analogues against poxvirus infections" *Reviews in Medical Virology* 14 (5): (2004): 289-300.
- **128.** Yun H. Jin, Peng Liu, Jianing Wang et al. "Practical Synthesis of D- and L-2-Cyclopentenone and Their Utility for the Synthesis of Carbocyclic Antiviral Nucleosides against Orthopox Viruses (Smallpox, Monkeypox, and Cowpox Virus)" *Journal of Organic Chemistry* 70 (1): (2005): 394.
- **129.** G. Andrei, A. Holy, F. Dal Pozzo et al. "Activity of novel acyclic pyrimidine nucleoside phosphonates against poxviruses" *Antiviral Research* 62 (2): (2004): A70.
- **130.** Donald F. Smee, Miles K. Wandersee, Kevin W. Bailey et al. "Characterization and treatment of cidofovirresistant vaccinia (WR strain) virus infections in cell culture and in mice" *Antiviral Chemistry and Chemotherapy* 16 (3): (2005): 203-211.
- **131.** Johan Neyts, Erik De Clercq "Therapy and short-term prophylaxis of poxvirus infections: historical background and perspectives" *Antiviral Research* 57 (1): (2003): 25-33.
- **132.** Gilbert Philippe-Alexandre, Lacrimioara Comanita, John Barrett et al. "Current status for high titre poxvirus stock preparation in CEF under serum-free medium conditions: implication for vaccine development" *Cytotechnology* 48 (1-3): (2005): 79-88.
- 133. Geoffrey L. Smith "Vaccinia virus virulence" Nova Acta Leopoldina 92 (344): (2005): 155-160.

- **134.** Y. Sun, E. Iglesias, A. Samri et al. "A systematic comparison of methods to measure HIV-1 specific CD8 T cells" *Journal of Immunological Methods* 272 (1): (2003): 23-34.
- **135.** Florence Buseyne, Adeline Catteau, Daniel Scott-Algara et al. "A vaccinia-based elispot assay for detection of CD8+ T cells from HIV-1 infected children" *Journal of Immunological Methods* 298 (1-2): (2005): 105-118.
- **136.** Anne-Laure Puaux, Benoit Delache, Severine Marconi et al. "Loss of reactivity of vaccine-induced CD4 T cells in immunized monkeys after SIV/HIV challenge" *AIDS* 19 (8): (2005): 757-765.
- **137.** Matthias Hebben, Veronique Duquesne, Joelle Cronier et al. "Modified vaccinia virus Ankara as a vaccine against feline coronavirus:immunogenicity and efficacy" *Journal of Feline Medicine and Surgery* 6 (2): (2004): 111-118.
- **138.** Robert Drillien, Daniele Spehner, Daniel Hanau "Modified vaccinia virus Ankara induces moderate activation of human dendritic cells" *Journal of General Virology* 85 (8): (2005): 2167-2175.
- **139.** Zhang Jian, Zhao Yueran, You Li et al. "Construction of Mutant N-ras/61 Gene Recombinant Vaccinia Virus rv-N-ras/61" *Chinese Journal of Cancer Biotherapy* 7 (4): (2000): 282-284.
- **140.** Wang Xijun, Wang Qinghua, Ge Jinying et al. "Generation of Recombinant Vaccinia Virus Expressing Attachment Glycoprotein of Nipah Virus" *Acta Microbiologica Sinica* 46 (4): (2006): 644-648.
- **141.** Huang Wei, Tian Houwen, Ren Jiao et al. "Construction and identification of non-replication recombinant vaccinia virus co-expressing human papillomavirus type 16 L1/L2/E6/E7 proteins" *Chinese Journal of Experimental and Clinical Virology* 19 (3): (2005): 240-243.
- **142.** Deng Jie, Sun Xiaoou, Wang Zhiliang et al. "Expression of the G glycoprotein from the respiratory syncytial virus field strain by recombinant vaccinia virus" *Chinese Journal of Microbiology and Immunology* 24 (4): (2004): 284-286.
- **143.** Li Xun, Xue Caifang, Maio Jun et al. "Combined vaccination with different type vaccines of Plasmodium falciparum AMA1" *Chinese Journal of Microbiology and Immunology* 25 (11): (2005): 949-952.
- **144.** Fan Jiangtao, Zhao Li, Chen Xinqiu et al. "The experimental study of HPV16 vaccines in prime-boost strategy" *Chinese Journal of Microbiology and Immunology* 26 (6): (2006): 510-515.
- **145.** Li Shumei, Li Xun, Xue Caifang et al. "DNA/MVA Combined Immunization: Antibody Response to Plasmodium falciparum Merozoite Surface Protein I in Mice" *Chinese Journal of Parasitology and Parasitic Diseases* 23 (2): (2005): 93-96.
- **146.** Wang Qinghua, Hu Sen, Wang Xijun et al. "Recombinant vaccinia viruses expressing envelope glycoprotein of rift valley fever virus" *Chinese Journal of Preventive Veterinary Medicine* 29 (6): (2007): 409-413.
- **147.** Tang Deyuan, Guo Wanzhu "Research on Japanese Encephalitis Virus and Its Vaccines" *Chinese Journal of Veterinary Science* 25 (2): (2005): 217-221.
- **148.** Wang Wenling, Lu Roujian, Lu Zhenhua et al. "Cloning of prM, E and NS1 Genes from Japanese Encephalitis Virus GSS Strain and Their Expression in Non-Replicative Vaccinia Virus" *Chinese Journal of Virology* 19 (3): (2003): 210-216.
- **149.** Qi Jianguo, Gu Shuyan, Wang Weidong et al. "Construction of non-replicating recombinant vaccinia virus expressing different protein fragments of Epstein-Barr virus major membrane protein gp340/220" *Chinese Journal of Virology* 20 (1): (2004): 22-27.
- **150.** Zhang Xiangmin, Wang Shifeng, Xin Wei et al. "Effect of codon bias on the gene expression efficiency in vaccinia virus vector" *Chinese Journal of Virology* 22 (5): (2006): 350-357.
- **151.** Wang Wenling, Lu Zhenhua, Xin Wei et al. "Non-replicating vaccinia recombinant expressed antigens of Japanese encephalitis virus can induce protective immunity in mice" *Chinese Journal of Virology* 21 (5): (2005): 370-374
- **152.** Yan Kexia, Tan Wenjie, Zhang Xiangmin et al. "Identification of epitopes on SARS-CoV (coronavirus) nucleocapsid protein that induce the Cross-or specific-reactivity among SARS-CoV, HCoV- OC43 and HCoV-229E" *Chinese Journal of Virology* 22 (4): (2006): 248-255.

- **153.** Tian Houwen, Ren Jiao, Huang Wei et al. "Construction of non-replicating recombinant vaccinia virus expressing human papillomavirus 16E6/E7 fusion protein and study of its immunogenicity and antitumor response" *Chinese Journal of Virology* 22 (5): (2006): 358-363.
- **154.** Dai Chunming, Zhang Xiaoyan, Wang Shuhui et al. "Examination of the Combined Immune Efficacy of a DNA Vector and Recombinant Vaccinia Virus Expressing the Gag Protein of Equine Infectious Anemia Virus (EIAV)" *Chinese Science Bulletin* 49 (20): (2004): 2057-2061.
- **155.** Dai Chunming, Zhang Xiaoyan, Zhang Ranran et al. "Combination immunization with EIAV Env protein expressed by recombinant baculovirus and recombinant vaccinia virus containing env gene" *Journal of Cellular and Molecular Immunology* 20 (4): (2004): 410-414.
- **156.** Zhang Xiaoyan, Li Hongmei, Liang Hua et al. "Induction of EIAV-specific cellular immune response by attenuated EIAV vaccine" *Journal of Cellular and Molecular Immunology* 22 (1): (2006): 115-118.
- **157.** Wu Ting, Chen Min, Ou Shanhai et al. "Immune response induced by combinant immunization of different HBsAg vaccines in mice" *Journal of Cellular and Molecular Immunology* 22 (5): (2006): 568-570.
- **158.** Wang Pingzhong, Bai Xuefan, Zhang Ying et al. "Preparation and Identification of Recombinant Vaccinia Virus Harboring Hantaan Virus S Gene" *Journal of Medical Postgraduates* 17 (1): (2004): 18-20.
- **159.** Li Pingping, Chen Naimin, Ying Baoling et al. "Construction, expression, and genetic stability of glycoprotein of recombinant vaccinia rabies virus" *Journal of Microbiology* 22 (2): (2002): 18-20.
- **160.** Wang Pingzhong, Bai Xuefan, Pan Lei et al. "Expression of Recombinant Vaccinia Virus Containing Hantaan Virus S Gene in B-lymphoblastoid Cell Lines from HFRS Patients" *Virologica Sinica* 18 (5): (2003): 415-419.
- **161.** Wu Yingsong, Feng Yu, Dong Wenqi "A vaccinia replication system for producing recombinant hepatitis C virus" *World Journal of Gastroenterology* 10 (18): (2004): 2670-2674.
- **162.** Zhang Yuxia, Li Shu, Shan Ming et al. "Hepatitis B virus core antigen epitopes presented by HLA-A2 single-chain trimers induce functional epitope-specific CD8+ T-cell responses in HLA-A2.1/Kb transgenic mice" *Immunology* 121 (1): (2007): 105-112.
- **163.** Stephen Kent, Robert De Rose, Erik Rollman "Drug evaluation: DNA/MVA prime-boost HIV vaccine" *Current Opinion in Investigational Drugs* 8 (2): (2007): 159-167.
- **164.** Gavin Clydesdale, Jenny Pekin, Sandra Beaton et al. "Contraception in mice immunized with recombinant zona pellucida subunit 3 proteins correlates with Th2 responses and the levels of interleukin 4 expressed by CD4+ cells" *Reproduction* 128 (6): (2004): 737-745.
- **165.** Dodie S. Pouniotis, Owen Proudfoot, Gabriela Minigo et al. "A new boost for malaria vaccines" *Trends in Parasitology* 20 (4): (2004): 157-160.
- **166.** Gerhard H. Reubel, Sandra Beaton, Daryl Venables et al. "Experimental inoculation of European red foxes with recombinant vaccinia virus expressing zona pellucida C proteins" *Vaccine* 23 (35): (2005): 4417-4426.
- **167.** Charani Ranasinghe, Jill C. Medveczky, Donna Woltring et al. "Evaluation of fowlpox-vaccinia virus prime-boost vaccine strategies for high-level mucosal and systemic immunity against HIV-1" *Vaccine* 24 (31-32): (2006): 5881-5895.
- **168.** Barbara E.H. Coupar, Damian F.J. Purcell, Scott A. Thomson et al. "Fowlpox virus vaccines for HIV and SHIV clinical and pre-clinical trials" *Vaccine* 24 (9): (2006): 1378-1388.
- **169.** Lucy Heinemann, Stephanie Dillon, Anna Crawford et al. "Flow cytometric quantitation of the protective efficacy of dendritic cellbased vaccines in a human papillomavirus type 16 murine challenge model" *Journal of Virological Methods* 117 (1): (2004): 9-18.
- **170.** Oka Kiyomasa, Nagano-Fujii Motoko, Yoshida Isao et al. "Hepatitis C virus core protein selectively inhibits synthesis and accumulation of p21/Waf1 and certain nuclear proteins" *Microbiology and Immunology* 47 (6): (2003): 429-438.

- 171. Okamura Tomotaka, Someya Kenji, Matsuo Kazuhiro et al. "Recombinant vaccinia DIs expressing simian immunodeficiency virus gag and pol in mammalian cells induces efficient cellular immunity as a safe immunodeficiency virus vaccine candidate" *Microbiology and Immunology* 50 (12): (2006): 989-1000.
- 172. Celia Perales, Luis Carrasco, Maria Eugenia Gonzalez "Regulation of HIV-1 env mRNA translation by Rev protein" *Biochimica et Biophysica Acta (BBA) Molecular Cell Research* 1743 (1-2): (2005): 169-175.
- **173.** Maria M. Garcia-Briones, Esther Blanco, Cristina Chiva et al. "Immunogenicity and T cell recognition in swine of foot-and-mouth disease virus polymerase 3D" *Virology* 322 (2): (2004): 264-275.
- **174.** Carmen Elena Gomez, Fernando Abaitua, Dolores Rodriguez et al. "Efficient CD8+ T cell response to the HIV-env V3 loop epitope from multiple virus isolates by a DNA prime/vaccinia virus boost (rWR and rMVA strains) immunization regime and enhancement by the cytokine IFN-gamma" *Virus Research* 105 (1): (2004): 11-22.
- **175.** Fernando Abaitua, Juan Ramon Rodriguez, Ana Garzon et al. "Improving recombinant MVA immune responses: potentiation of the immune responses to HIV-1 with MVA and DNA vectors expressing Env and the cytokines IL-12 and IFN-gamma" *Virus Research* 116 (1-2): (2006): 11-20.
- **176.** Carolina Johnstone, Patricia de Leon, Francisco Medina et al. "Shifting immunodominance pattern of two cytotoxic T-lymphocyte epitopes in the F glycoprotein of the Long strain of respiratory syncytial virus" *Journal of General Virology* 85 (11): (2004): 3229.
- 177. Carmen Elena Gomez, Jose Luis Najera, Victoria Jimenez et al. "Generation and immunogenicity of novel HIV/AIDS vaccine candidates targeting HIV-1 Env/Gag-Pol-Nef antigens of clade C" *Vaccine* 25 (11): (2007): 1969-1992.
- 178. Carmen Elena Gomez, Jose Luis Najera, Eva Perez Jimenez et al. "Head-to-head comparison on the immunogenicity of two HIV/AIDS vaccine candidates based on the attenuated poxvirus strains MVA and NYVAC coexpressing in a single locus the HIV-1BX08 gp120 and HIV-1(IIIB) Gag-Pol-Nef proteins of clade B" *Vaccine* 25 (15): (2007): 2863-2885.
- **179.** Carmen E. Gomez, Andree Marie Vandermeeren, Maria Angel Garcia et al. "Involvement of PKR and RNase L in translational control and induction of apoptosis after Hepatitis C polyprotein expression from a vaccinia virus recombinant" *Virology Journal* 2 (1): (2005): 81.
- **180.** B. Makitalo, P. Lundholm, J. Hinkula et al. "Enhanced cellular immunity and systemic control of SHIV infection by combined parenteral and mucosal administration of a DNA prime MVA boost vaccine regimen" *Journal of General Virology* 85 (8): (2005): 2407-2419.
- **181.** Jennifer A. Slyker, Barbara L. Lohman, Dorothy A. Mbori-Ngacha et al. "Modified vaccinia Ankara expressing HIVA antigen stimulates HIV-1-specific CD8 T cells in ELISpot assays of HIV-1 exposed infants" *Vaccine* 23 (38): (2005): 4711.
- **182.** Dania Vazquez-Blomquist, Diogenes Quintana, Carlos A. Duarte "Modified-vaccinia-virus-ankara (MVA) priming and fowlpox-virus booster elicit a stronger CD8+ T-cell response in mice against an HIV- epitope than does a DNA/poxvirus prime-booster approach" *Biotechnology and Applied Biochemistry* 39 (3): (2004): 313-318.
- **183.** Diogenes Quintana-Vazquez, Dania M. Vazquez-Blomquist, Rodriguez Ernesto Galban et al. "A vaccination strategy consisting of Semliki-Forest-virus (SFV) DNA prime and fowlpox-virus boost significantly protects mice from a recombinant (HIV-1) vaccinia-virus infection" *Biotechnology and Applied Biochemistry* 41 (1): (2005): 59-66.
- **184.** J.L. Roque-Resendiz, R. Rosales, P. Herion "MVA ROP2 vaccinia virus recombinant as a vaccine candidate fortoxoplasmosis" *Parasitology* 128 (4): (2004): 397-405.
- **185.** Marilene F. Almeida, Luzia F.A. Martorelli, Caroline C. Aires et al. "Indirect oral immunization of captive vampires, Desmodus rotundus" *Virus Research* 111 (1): (2005): 77-82.
- **186.** Luciana B. De Arruda, Priya R. Chikhlikar, J. Thomas August et al. "DNA vaccine encoding human immunodeficiency virus-1 Gag, targeted to the major histocompatibility complex II compartment by lysosomal-associated membrane protein, elicits enhanced long-term memory response" *Immunology* 112 (1): (2004): 126-135.

- **187.** A.R. Pinto, J.C. Fitzgerald, G.P. Gao et al. "Induction of CD8+ T cells to an HIV-1 antigen upon oral immunization of mice with a simian E1-deleted adenoviral vector" *Vaccine* 22 (5-6): (2004): 698-704.
- **188.** M.F. Ferrer, G. Calamante "Design and Construction of Transfer Vectors for the Obtention of Recombinant Modified Vaccinia Virus Ankara" *XIX PanAmerican Congress of Veterinary Sciences* (2004).
- **189.** Claudia C. Motran, Fernando Lopez Diaz, Carolina L. Montes et al. "In vivo expression of recombinant pregnancy-specific glycoprotein 1ainduces alternative activation of monocytes and enhances Th2-type immune response" *European Journal of Immunology* **33** (11): (2003): 3007-3016.
- **190.** Marco Franchini, Hanspeter Hefti, Sabine Vollstedt et al. "Dendritic Cells from Mice Neonatally Vaccinated with Modified Vaccinia Virus Ankara Transfer Resistance against Herpes Simplex Virus Type I to Naive One-Week-Old Mice" *Journal of Immunology* 172 (10): (2004): 6304-6312.
- **191.** Behazine Combadiere, Alexandre Boissonnas, Guislaine Carcelain et al. "Distinct time effects of vaccination on long-term proliferative and IFN-?-producing T cell memory to smallpox in humans" *Journal of Experimental Medicine* 199 (11): (2004): 1585-1593.
- **192.** Hsieh Szumin, Pan Sungching, Chen Sheyying et al. "Age distribution for T cell reactivity to vaccinia virus in a healthy population" *Clinical Infectious Diseases* 38 (1): (2004): 86-89.
- **193.** K. Dumbell, Madalene Richardson "Virological investigations of specimens from buffaloes affected by buffalopox in Maharashtra State, India between 1985 and 1987" *Archives of Virology* 128 (3-4): (1993): 257-267.
- **194.** R.K. Singh, V. Balamurugan, M. Hosamani et al. "B5r gene based sequence analysis of Indian buffalopox virus isolates in relation to other orthopoxviruses" *Acta Virologica* 51 (1): (2007): 47-50.
- **195.** Giliane S. Trindade, Zelia I.P. Lobato, Betania P. Drumond et al. "Short report: Isolation of two Vaccinia virus strains from a single bovine vaccinia outbreak in rural area from Brazil: Implications on the emergence of zoonotic orthopoxviruses" *American Journal of Tropical Medicine and Hygiene* 75 (3): (2006): 486-490.
- **196.** G. Li, N. Chen, R.L. Roper et al. "Complete coding sequences of the rabbitpox virus genome" *Journal of General Virology* 86 (11): (2005): 2969-2977.
- **197.** Tonie E. Rocke, F. Joshua Dein, Martina Fuchsberger et al. "Limited infection upon human exposure to a recombinant raccoon pox vaccine vector" *Vaccine* 22 (21-22): (2004): 2757-2760.
- **198.** Yu Li, Susan L. Ropp, Hui Zhao et al. "Orthopoxvirus pan-genomic DNA assay" *Journal of Virological Methods* 141 (2): (2007): 154-165.
- 199. D.A. Henderson "Bioterrorism as a Public Health Threat" Emerging Infectious Diseases 4 (3): (1998): 488-492.
- **200.** Susan L. Ropp, Qi Jinn, Janice C. Knight et al. "PCR Strategy for Identification and Differentiation of Smallpox and Other Orthopoxviruses" *Journal of Clinical Microbiology* 33 (8): (1995): 2069-2076.
- **201.** M. Sofi Ibrahim, David A. Kulesh, Sharron S. Saleh et al. "Real-Time PCR Assay To Detect Smallpox Virus" *Journal of Clinical Microbiology* 41 (8): (2003): 3835-3839.
- **202.** Marcus Panning, Marcel Asper, Stefanie Kramme et al. "Rapid detection and differentiation of human pathogenic orthopox viruses by a fluorescence resonance energy transfer real-time PCR assay" *Clinical Chemistry* 50 (4): (2004): 702-708.
- **203.** K. Kolmel, C. Neumann, P. Schupp et al. "Cowpox virus in a 12-year-old boy: rapid identification by an orthopoxvirus-specific polymerase chain reaction" *British Journal of Dermatology* 145 (1): (2001): 146-150.
- **204.** Andreas Nitsche, Heinz Ellerbrok, Georg Pauli "Detection of orthopoxvirus DNA by real-time PCR and identification of variola virus DNA by melting analysis" *Journal of Clinical Microbiology* 42 (3): (2004): 1207-1213.
- **205.** Michael Schmidt, Kurt W. Roth, Hermann Meyer et al. "Nucleic acid test screening of blood donors for orthopoxviruses can potentially prevent dispersion of viral agents in case of bioterrorism" *Transfusion* 45 (3): (2005): 399-403.

- **206.** David Pulford, Hermann Meyer, Gale Brightwell et al. "Amplification refractory mutation system PCR assays for the detection of variola and Orthopoxvirus" *Journal of Virological Methods* 117 (1): (2004): 81-90.
- **207.** Duan Hongyuan, Deng Yongqiang, Zhao Hui et al. "Establishment of Detection Methods for Vaccinia Virus" *Bulletin of the Academy of Military Medical Sciences* 29 (3): (2005): 272-274.
- **208.** Feng Sulan, Chen Xizhao, Tian Kegong "Cloning and Expression of the P32 Gene of Capripoxvirus and Establishment of a PCR Diagnostic Method" *CNKI China Doctor/Master Dissertations Full-text Database* (2005).
- **209.** Kang Wenyu, Xu Zizhong, Gao Hong et al. "Capripoxvirus" *China Animal Husbandry & Veterinary Medicine* 31 (12): (2004): 33-36.
- **210.** Zhou Bijun, Yu Tiande, Yin Zhengxing et al. "Electron Microscope Observation of Virus Particles of Goat Pox" *Chinese Journal of Preventive Veterinary Medicine* 28 (3): (2006): 316-319.
- **211.** Zhou Bijun, Feng Yuanzhang, Xu Leren "Comparison study of serologic diagnosis techniques for goat pox" *Chinese Journal of Preventive Veterinary Medicine* 28 (4): (2006): 423-426.
- **212.** Yang Songtao, Xia Xianzhu, Qiao Jun et al. "Isolation and Identification of CC Virulent Strain of Mousepox Virus" *Journal of Pathogen Biology* 2 (1): (2007): 4-7.
- **213.** Wang Yan, Ma Wenli, Song Yanbin et al. "Development of a DNA Microarray for Detection of Orthopoxvirus" *Journal of Preventive Medicine of Chinese People's Liberation Army* 22 (2): (2004): 86-89.
- **214.** Wang Yan, Ma Wenli, Mao Xiangming et al. "Design and preparation of oligonucleotide microarray for vaccinia virus" *Journal of Southern Medical University* 24 (2): (2004): 180-183.
- **215.** I.E. Agranovski, A.S. Safatov, A.A. Sergeev et al. "Rapid detection of airborne viruses by personal bioaerosol sampler combined with the PCR device" *Atmospheric Environment* 40 (21): (2006): 3924-3929.
- **216.** I.E. Agranovski, A.S. Safatov, A.I. Borodulin et al. "New personal sampler for viable airborne viruses: Feasibility study" *Journal of Aerosol Science* 36 (5-6): (2005): 609-617.
- **217.** Sergei N. Shchelkunov, E.V. Gavrilova, Igor V. Babkin "Multiplex PCR detection and species differentiation of orthopoxviruses pathogenic to humans" *Molecular and Cellular Probes* 19 (1): (2005): 1-8.
- **218.** L.S. Sandakhchiev, Ye.V. Gavrilova, S.N. Shchelkunov et al. "Identification of Variola Virus (Smallpox) and other Orthopoxviruses (Monkeypox, Cowpox, Vaccinia) Pathogenic for Humans using a Multiplex PCR" *Current Technologies in the Diagnosis of Particularly Dangerous Infectious Diseases* 2003 (4): (2003): 44-46.
- **219.** S.N. Shchelkunov "Molecular-Genetic Investigations of Variola Virus and Closely Related Orthopox Viruses" *International Conference "Physico-Chemical Biology"* (2006).
- **220.** M.V. Mikheev, S.A. Lapa, S.N. Shchelkounov et al. "Species identification of Orthopoxviruses in oligonucleotide microchips" *Problems of Virology* 48 (1): (2003): 4-9.
- **221.** J.J. Esposito, F. Fenner "Poxviruses In: Fields Virology" *Lippincott Williams & Wilkins* 2 (Fourth Edition): (2001).
- **222.** S.L. Ropp, Q. Jin, J.C. Knight et al. "PCR strategy for identification and differentiation of small pox and other orthopoxviruses" *Journal of Clinical Microbiology* 33 (8): (1995): 2069-2076.
- **223.** Pavel Fedorovich Safronov "Sequencing and Analysis of the Genomic Organization of Cowpox Virus (CPV) Strain GRI-90" *Federal Portal for Science and Innovation* (2004).
- **224.** Anna M. Likos, Scott A. Sammons, Victoria A. Olson et al. "A tale of two clades: monkeypox viruses" *Journal of General Virology* 86 (10): (2005): 2661-2672.
- **225.** Sergei N. Shchelkunov, Alexei V. Totmenin, Igor V. Babkin et al. "Human monkeypox and smallpox viruses: genomic comparison" *FEBS Letters* 509 (1): (2001): 66-70.

- **226.** S.N. Shchelkunov, A.V. Totmenin, P.F. Safronov et al. "Analysis of the Monkeypox Virus Genome" *Virology* 297 (2002): 172-194.
- **227.** Koert J. Stittelaar, Johan Neyts, Lieve Naesens et al. "Comparison of the efficacy of post exposure smallpox vaccination versus antiviral treatment with acyclic nucleotides against monkeypox virus infection" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- **228.** Sue Lewis-Jones "Zoonotic poxvirus infections in humans" *Current Opinion in Infectious Diseases* 17 (2): (2004): 81-89.
- **229.** Nanhai Chen, Guiyun Li, M. Kathryn Liszewski et al. "Virulence differences between monkeypox virus isolates from West Africa and the Congo basin" *Virology* 340 (2005): 46-63.
- **230.** Mary G. Reynolds, Whitni B. Davidson, Aaron T. Curns et al. "Spectrum of Infection and Risk Factors for Human Monkeypox, United States, 2003" *Emerging Infectious Diseases* 13 (9): (2007): 1332-1339.
- **231.** Ye.A. Uvarova, A.V. Totmenin, L.S. Sandakhchiyev et al. "In genome of western African monkeypox virus strains gene of complement-binding protein, important suppressor of inflammation, is deleted" *Transactions of the Academy of Sciences* 400 (1): (2005): 126-128.
- **232.** Igor V. Babkin, M.V. Mikheev, Sergei N. Shchelkunov et al. "Analysis of Nucleotide Sequences of Individual Orthopoxvirus Genes" *Meetings of the WHO Advisory Committee on Variola Virus* 2003 (5): (2003).
- **233.** S.M. Hazel, M. Bennett, J. Chantrey et al. "A longitudinal study of an endemic disease in its wildlife reservoir: cowpox and wild rodents" *Epidemiology and Infection* 124 (3): (2000): 551-562.
- **234.** David Carslake, Malcolm Bennett, Sarah Haze et al. "Inference of cowpox virus transmission rates between wild rodent host classes using space-time interaction" *Proceedings of the Royal Society of London: Biological Sciences* 273 (1588): (2006): 775-782.
- **235.** J. Chantrey, H. Meyer, D. Baxby et al. "Cowpox: reservoir hosts and geographic range" *Epidemiology and Infection* 122 (3): (1999): 455-460.
- **236.** H. Meyer, C. Schay, H. Mahnel et al. "Characterization of orthopoxviruses isolated from man and animals in Germany" *Archives of Virology* 144 (3): (1999): 491-501.
- **237.** M.J.L. Kik, P.L. Liu, J.A.M. van Asten "Cowpoxvirus infection in the Patagonian cavy (Dolichotis patagonum) emerging disease in an educational animal park the first reported case" *Veterinary Quarterly* 28 (2): (2006): 42-44.
- **238.** Andreas Nitsche, Andreas Kurth, Georg Pauli "Viremia in human Cowpox virus infection" *Journal of Clinical Virology* 40 (2): (2007): 160-162.
- **239.** N. Rajan, A.J. Carmichael, B.M. McCarron "Human cowpox: Presentation and investigation in an era of bioterrorism" *Journal of Infection* 51 (3): (2005): 167-169.
- **240.** B. Feuerstein, M. Jugens, E. Schnetz et al. "Cowpox and catpox infection: 2 Clinical case reports" *Dermatologist: Journal for Dermatology, Allergy, Venereology and Related Areas* 51 (11): (2000): 852-856.
- **241.** C. Schulze, M. Alex, H. Schirrmeier et al. "Generalized fatal Cowpox virus infection in a cat with transmission to a human contact case" *Zoonoses and Public Health* 54 (1): (2007).
- **242.** M. Tryland, H. Myrmel, G. Haukenes et al. "Clinical Cowpox Cases in Norway" *Scandinavian Journal of Infectious Diseases* 30 (3): (1998): 301-303.
- **243.** M. Tryland, T. Sandvik, H. Hansen et al. "Characteristics of four cowpox virus isolates from Norway and Sweden" *APMIS* 106 (6): (1998): 623-635.
- **244.** U. Hinrichs, H. van de Poel, T.S. van den Ingh "Necrotizing pneumonia in a cat caused by an orthopox virus" *Journal of Comparative Pathology* 121 (2): (199): 191-196.
- **245.** Paula M. Pelkonen, Kyllikki Tarvainen, Arja Hynninen et al. "Cowpox with severe generalized eruption, Finland" *Emerging Infectious Diseases* 9 (11): (2003): 1458-1461.

- **246.** M. Pfeffer, G. Burck, H. Meyer "Cowpox viruses in Germany: an analysis of 5 cases in 1998" *Berlin-Munich Veterinary Weekly* 112 (9): (1999): 334-338.
- **247.** Katrina L. Oie, David J. Pickup "Cowpox virus and other members of the orthopoxvirus genus interfere with the regulation of NF-kappaB activation" *Virology* 288 (1): (2001): 175-187.
- **248.** A.A. Vorob'ev, V.N. PodkuA-ko, V.V. MikhaA-lov "Results of basic and applied investigations on oral immunization against smallpox" *Journal of Microbiology, Epidemiology and Immunobiology* 2002 (1): (2002): 98-104.
- **249.** E.R. Tulman, G. Delhon, C.L. Afonso et al. "Genome of Horsepox Virus" *Journal of Virology* 80 (18): (2006): 9244-9258.
- **250.** Clarissa R.A. Damaso, Sheila A. Reis, Desree M. Jesus et al. "A PCR-based assay for detection of emerging vaccinia-like viruses isolated in Brazil" *Diagnostic Microbiology and Infectious Disease* 57 (2007): 39-46.
- **251.** Juliana A. Leite, Betania P. Drumond, Giliane S. Trindade et al. "Passatempo Virus, a Vaccinia Virus Strain, Brazil" *Emerging Infectious Diseases* 11 (12): (2005): 1935-1938.
- **252.** Teresa Keico Nagasse-Sugahara, Jonas Jose Kisielius, Marli Ueda-Ito et al. "Human vaccinia-like virus outbreaks in Sao Paulo and Goias States, Brazil: virus detection, isolation and identification" *Journal of the Sao Paulo Institute for Tropical Medicine* 46 (6): (2004).
- **253.** Flavio G. da Fonseca, Giliane S. Trindade, Ricardo L.A. Silva et al. "Characterization of a vaccinia-like virus isolated in a Brazilian forest" *Journal of General Virology* 83 (2002): 223-228.
- **254.** Giliane de Souza Trindade, Flavio Guimaraes da Fonseca, Joao Trindade Marques et al. "Aracatuba Virus: A Vaccinialike Virus Associated with Infection in Humans and Cattle" *Emerging Infectious Diseases* 9 (2): (2003): 1-6.
- **255.** Clarissa R.A. Damaso, Joseph J. Esposito, Richard C. Condit et al. "An Emergent Poxvirus from Humans and Cattle in Rio de Janeiro State: Cantagalo Virus May Derive from Brazilian Smallpox Vaccine" *Virology* 277 (2000): 439-449.
- **256.** Giliane S. Trindade, Flavio G. da Fonseca, Joao T. Marques et al. "Belo Horizonte virus: a vaccinia-like virus lacking the A-type inclusion body gene isolated from infected mice" *Journal of General Virology* 85 (2004): 2015-2021.
- **257.** Ye.A. Bulatov, M. Mambetaliyev, Seydigapbar Mamadaliyevich Mamadaliyev et al. "Study on Pathogenic Characteristics of the Camelpox Virus in Different Animal and Poultry Species" *International Conference* "Development of International Collaboration in Infectious Disease Research" (2004).
- **258.** V. Bhanuprakash, B. Indrani, K. Hosamani et al. "The current status of sheep pox disease" *Comparative Immunology, Microbiology & Infectious Diseases* 29 (1): (2006): 27-60.
- **259.** Ye.S. Orlova, A.V. Scherbakov, V.I. Diyev et al. "Differentiation of vaccine and epidemic strains of sheep pox virus" *Collected Works of the Federal Center for Animal Health* 3 (2005): 102-108.
- **260.** A. Nitsche, H.R. Gelderblom, K. Eisendle et al. "Pitfalls in diagnosing human poxvirus infections" *Journal of Clinical Virology* 38 (2): (2007): 165-168.
- **261.** Marek Cyrklaff, Cristina Risco, Jose Jesus Fernandez et al. "Cryo-electron tomography of vaccinia virus" *Proceedings of the National Academy of Sciences of the United States of America* 102 (8): (2005): 2772.
- **262.** Caroline Gubser, Stephane Hue, Paul Kellam et al. "Poxvirus genomes: a phylogenetic analysis" *Journal of General Virology* 85 (1): (2004): 105-117.
- **263.** S. Welsch, L. Doglio, J.K. Locker "The Vaccinia Virus I3L Gene Product Is Localized to a Complex Endoplasmic Reticulum-Associated Structure That Contains the Viral Parental DNA" *Journal of Virology* 77 (10): (2003): 6014-6028.
- **264.** Gemma C. Carter, Mansun Law, Michael Hollinshead et al. "Entry of the vaccinia virus intracellular mature virion and its interactions with glycosaminoglycans" *Journal of General Virology* 86 (5): (2005): 1279-1290.

- **265.** I.P. Gileva, I.A. Ryazankin, Z.A. Maksyutov et al. "Comparative assessment of the properties of orthopoxviral soluble receptors for tumor necrosis factor" *Doklady Biochemistry and Biophysics* 390 (2003): (2003): 160-164.
- **266.** Jayati Mullick, John Bernet, Yogesh Panse et al. "Identification of complement regulatory domains in vaccinia virus complement control protein" *Journal of Virology* 79 (19): (2005): 12382-12393.
- **267.** John Bernet, Jayati Mullick, Yogesh Panse et al. "Kinetic analysis of the interactions between vaccinia virus complement control protein and human complement proteins C3b and C4b" *Journal of Virology* 78 (17): (2004): 209-210.
- **268.** Geraldine Maloney, Martina Schroder, Andrew G. Bowie "Vaccinia Virus Protein A52R Activates p38 Mitogenactivated Protein Kinase and Potentiates Lipopolysaccharide-induced Interleukin-10" *Journal of Biological Chemistry* 280 (35): (2005): 30838-30844.
- **269.** Julianne Stack, Ismar R. Haga, Martina Schroder et al. "Vaccinia virus protein Toll-like-interleukin-1 A46R targets multiple receptor adaptors and contributes to virulence" *Journal of Experimental Medicine* 201 (6): (05): 1007-1018.
- **270.** Gary DiPerna, Julianne Stack, Andrew G. Bowie et al. "Poxvirus Protein N1L Targets the I-?B Kinase Complex, Inhibits Signaling to NF-?B by the Tumor Necrosis Factor Superfamily of Receptors, and Inhibits NF-?B and IRF3 Signaling by Toll-like Receptors" *Journal of Biological Chemistry* 279 (35): (2004): 36570.
- **271.** Alicja Kluczyk, Marek Cebrat, Renata Zbozien-Pacamaj et al. "On the peptide-antipeptide interactions in interleukin-1 receptor system" *Acta Biochimica Polonica* 51 (1): (2004): 57.
- **272.** Colin E. Henderson, Krystyna Bromek, Nicholas P. Mullin et al. "Solution structure and dynamics of the central CCP module pair of a poxvirus complement control protein" *Journal of Molecular Biology* 307 (1): (2001): 323-339.
- **273.** Antonio Alcami, Geoffrey L. Smith "The vaccinia virus soluble interferon-gamma receptor is a homodimer" *Journal of General Virology* 83 (3): (2002): 545-549.
- **274.** Geoffrey L. Smith, Julian A. Symons, Anu Khanna et al. "Vaccinia virus immune evasion" *Immunological Reviews* 159 (1): (1997): 137-154.
- **275.** Tomas P. Carroll, , Catherine M. Greene, Clifford C. Taggart et al. "Viral Inhibition of IL-1- and Neutrophil Elastase-Induced Inflammatory Responses in Bronchial Epithelial Cells" *Journal of Immunology* 175 (11): (2005): 7594-7601.
- **276.** Lena Kash, Bruno O. Villoutreix, Marten Steen et al. "Structural stability and heat-induced conformational change of two complement inhibitors: C4b-binding protein and factor H" *Protein Science* 13 (5): (2004): 1356-1364.
- **277.** O.O. Odunuga, Y.T. Ghebremariam, G.J. Kotwal "Lister homologue of vaccinia virus complement control protein is two amino acids shorter, has putative glycosylation sites and other functional and structural differences" *The FEBS Journal* 272 (Supplement 1): (2005): 15-16.
- **278.** B.E.F. Mota, J.A. Leite, C.A. Bonjardim et al. "Production Of Extracellular Infective Forms Of Cowpox Virus Is Altered By Deletion Of The Ati Gene" *III Forum on Microbiology at the Federal University of Minas Gerais Graduate Program in Microbiology* (2007).
- **279.** Zielinski Andrzej, Stefanoff Pawel "Smallpox—historical or real threat?" *Journal of Epidemiology* 58 (Supplement 1): (2004): 22-27.
- **280.** Nishiura Hiroshi "Analysis of a previous smallpox vaccination study: estimation of the time period required to acquire vaccine-induced immunity as assessed by revaccination" *Southeast Asian Journal of Tropical Medicine & Public Health* 37 (4): (2006): 673-680.
- **281.** S. Stienlauf, M. Shoresh, A. Solomon et al. "Kinetics of formation of neutralizing antibodies against vaccinia virus following re-vaccination" *Vaccine* 17 (3): (1999): 201-204.

- **282.** Igor Vladimirovich Plyasunov, A.N. Sergeyev, S.A. Melnikov "Study of the Safety, Reactogenicity and Antigenic Activity of an Oral Recombinant Bivaccine against Smallpox and Hepatitis B" *Vector News. Archive. Defense of Dissertations* (2006).
- **283.** Kidokoro Minoru, Tashiro Masato, Shida Hisatoshi "Development of a Highly Genetically Stable and Immunogenic Attenuated Smallpox Vaccine, and Analysis of the Function of the B5R Gene" *Program and Abstracts from the Annual Meeting of the Japanese Society for Vaccinology* 9 (2005): 102.
- **284.** Shinmura Yasuhiko, Kanehara Tomomi, Sato Azusa et al. "Investigation Into the Protection Mechanisms of Attenuated Smallpox Vaccine LC16m8" *Program and Abstracts from the Annual Meeting of the Japanese Society for Virology* **54** (2006): 391.
- **285.** Mike M. Puetz, Claire M. Midgley, Mansun Law et al. "Quantification of antibody responses against multiple antigens of the two infectious forms of Vaccinia virus provides a benchmark for smallpox vaccination" *Nature Medicine* 12 (11): (2006): 1310-1315.
- **286.** V.N. Pak, V.V. Raspopin, V. Petrov et al. "Cloning of the growth factor gene from vaccinia virus LIVP strain in Escherichia coli cells" *Molecular Genetics, Microbiology and Virology* 1992 (9-10): (1992): 19-21.
- **287.** Sergei N. Shchelkunov, Sergei M. Resenchuk, Alexei V. Totmenin et al. "Comparison of the genetic maps of variola and vaccinia viruses" *FEBS Letters* 327 (3): (1993): 321-324.
- **288.** Sergei N. Shchelkunov, Vladimir M. Blinov, Lev S. Sandakhchiev "Genes of variola and vaccinia viruses necessary to overcome the host protective mechanisms" *FEBS Letters* 319 (1-2): (1993): 80-83.
- **289.** V.I. Chernos, T.S. Vovk, O.N. Ivanova et al. "Insertion mutants of the vaccinia virus: Genes E7R and D8L inactivation effect on the virus biological characteristics" *Molecular Genetics, Microbiology and Virology* 1993 (2): (1993): 30-34.
- **290.** O.I. Serpinskiy, G.V. Kochneva, I.Kh. Urmanov et al. "Construction of Recombinant Variants of Orthopoxviruses Through the Insertion of Exogenous Genes Into the Intragenic Space of the Viral Genomes" *Molecular Biology* 30 (5): (1996): 1055-1065.
- **291.** Sergei Nikolaevich Shchelkunov, Alexei Vladimirovich Totmenin "Two types of deletions in orthopoxvirus genomes" *Virus Genes* 9 (3): (1995): 231-245.
- **292.** V.M. Blinov, A.V. Totmenin, S.M. Resenchuk et al. "Study of the Structural-Functional Organization of the Variola Virus Genome. IV. Sequencing and Analysis of the Nucleotide Sequence of the Right Terminus of the India-1967 Strain Genome" *Molecular Biology* 29 (4): (1995): 7782-7789.
- **293.** S.V Seregin, I.N Babkina, A.E Nesterov et al. "Comparative studies of gamma-interferon receptor-like proteins of variola major and variola minor viruses" *FEBS Letters* 382 (1-2): (1996): 79-83.
- **294.** G. Kochneva, I. Kolosova, T. Maksyutova et al. "Effects of deletions of kelch-like genes on cowpox virus biological properties" *Archives of Virology* 150 (9): (2005): 1857-1870.
- **295.** G.V. Kochneva, O.S. Taranov, T.A. Lupan et al. "Deletion of kelch-like genes affects structural organization of a cell during cowpox virus reproduction" *Basic Science for Medicine* (2007): 61.
- **296.** I.V. Kolosova, S.V. Seregin, G.V. Kochneva et al. "Orthopoxvirus Genes for Kelch-like Proteins: II. Construction of Cowpox Virus Variants with Targeted Gene Deletions" *Molecular Biology* 37 (4): (2003): 495-502.
- **297.** I.V. Kolosova, S.V. Seregin, G.V. Kochneva et al. "The study of orthopoxvirus genes for Kelch-like proteins. II. Construction of cowpox virus variants with targeted gene deletions" *Molecular Biology* 37 (4): (2003): 585-594.
- **298.** L.A. Shundrin, V.A. Ryabinin, Ye.V. Kostina et al. "Microarray-Based Genetic Diagnosis of Human-Pathogenic Orthopoxviruses" *Basic Science for Medicine* (2005).
- **299.** Elena I. Ryabchikova, O.V. Timoshenko, I.V. Vinogradov et al. "Comparative study of extracellular enveloped virus formation by different variola virus strains" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).

- **300.** A.M. Kolchinsky, D.A. Gryadunov, Yu.P. Lysov et al. "Gel-based microchips: History and prospects" *Molecular Biology* 38 (1): (2004): 5-16.
- **301.** Ye.V. Gavrilova, I.V. Babkin, S.N. Schelkunov "Multiplex PCR analysis for species specific express-identification of orthopoxviruses" *Molecular Genetics, Microbiology and Virology* 2003 (1): (2003): 45-52.
- **302.** V.A. Ryabinin, L.A. Shundrin, E.V. Kostina et al. "An oligonucleotide microarray for detection and discrimination of orthopoxviruses based on oligonucleotide sequences of two viral genes" *Molecular Genetics, Microbiology and Virology* 2006 (4): (2006): 23-30.
- **303.** Anonymous "Biological microchip for identification of orthopoxviruses and agents causing the same clinical picture as in smallpox" *Problems of Virology* 52 (2): (2007): 41-45.
- **304.** Yelena Vasilyevna Gavrilova, S.N. Shchelkunov "Development of a Multiplex PCR-based Method for Species-Specific Diagnosis of Human-Pathogenic Orthopoxviruses" *Vector News. Archive. Defense of Dissertations* (2007).
- **305.** Sergei N. Shchelkunov, E.V. Gavrilova, I.V. Babkin et al. "Development of modern methods for species-specific differentiation of orthopoxviruses" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- **306.** E.S. Orlova, A.V. Shcherbakova, V.I. Diev et al. "Differentiation of capripoxvirus species and strains by polymerase chain reaction" *Molecular Biology* 40 (1): (2006): 158-164.
- **307.** Sergei N. Shchelkunov, A.A. Guskov, E.B. Sokunova et al. "Viability estimation of variola virus isolates from the Russian collection" *Meetings of the WHO Advisory Committee on Variola Virus* 2003 (5): (2003).
- **308.** N. Tikunova, V. Morozova, E. Bovshik et al. "Combinatorial antibodies against orthopoxviruses" *Antiviral Research* 57 (3): (2003): A73.
- **309.** V.V. Morozova, N.V. Tikunova, T.A. Batanova et al. "Human mini-antibodies to orthopoxviruses" *Bulletin of the Russian Academy of Medical Sciences* 2004 (8): (2004): 22-27.
- **310.** V.V. Dubrovskaya, A.B. Ulitin, A.G. Laman et al. "Construction of a combinatorial immune library of human single-chain antibodies to orthopoxviruses and selection of antibodies to recombinant prA30L of the variola virus" *Molecular Biology* 41 (1): (2007): 157-167.
- **311.** Anonymous "Construction of combinatorial immune library of single chain human antibodies to orthopoxviruses and selection from this library antibodies to recombinant protein prA30L of variola virus" *Molecular Biology* 41 (1): (2007): 173-185.
- **312.** T.A. Batanova, A.B. Ulitin, V.V. Morozova et al. "Construction and characterization of combinatorial naive phage library of human scFvs" *Molecular Genetics, Microbiology and Virology* 2006 (3): (2006): 35-41.
- **313.** V.V. Morozova, A.A. Il'ichev, E.B. Sokunova et al. "Human recombinant antibodies to variola virus" *Problems of Virology* 50 (6): (2005): 20-25.
- **314.** Viktoriya Vladislavovna Dubrovskaya, N.V. Tikunova, V.B. Loktev et al. "Construction of Immune Combinatorial Library of Single-Stranded Human Antibodies and Selection of Orthopoxvirus Neutralizing Antibodies" *Vector News. Archive. Defense of Dissertations* (2006).
- **315.** Vera V. Morozova, Viktoria V. Voronina, Maia V. Shveigert et al. "Group-specific and neutralizing human SCFV to orthopoxviruses from a combinatorial phage library" *Antiviral Research* 65 (3): (2005): A83.
- **316.** Nina Tikunova, Vera Morozova, Viktoria Voronina et al. "Generation of anti-orthopoxvirus fully human Mabs derived from phage display singlechain antibodies" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- **317.** N.V. Tikunova, E.I. Bovshik, T.E. Yun et al. "Human Combinatorial Antibodies against Orthopoxviruses" *Meetings of the WHO Advisory Committee on Variola Virus* 2003 (5): (2003).
- **318.** Nina Tikunova, Victoria Dubrovskaia, Vera Morozova et al. "Development of neutralizing human scFv-antibodies against pathogenic orthopoxviruses" *Meetings of the WHO Advisory Committee on Variola Virus* (2006).

- **319.** T.E. Yun, N.V. Tikunova, L.N. Shingarova et al. "The full-length recombinant human antibody to vaccinia virus" *Doklady Biochemistry and Biophysics* 407 (1): (2006): 98-101.
- **320.** I.P. Gileva, I.A. Ryazankin, T.S. Nepomnyashchikh et al. "Expression of genes for orthopoxviral TNF-binding proteins and study resulted recombinant proteins" *Molecular Biology* 39 (2): (2005): 245-254.
- **321.** T.S. Nepomnyashchikh, L.R. Lebedev, G.V. Kochneva et al. "TNF-Binding Proteins of Orthopoxvirus Species: Expression, Purification and Some Properties" *Cytokines and Inflammation* 4 (2): (2005).
- **322.** I.P. Gileva, E.M. Malkova, T.S. Nepomnyashchih et al. "Influence of Variola Virus TNF-Binding Protein on Experimental LPS-Induced Endotoxic Shock" *Cytokines and Inflammation* 5 (1): (2006): 44-49.
- **323.** T.S. Nepomnyashchikh, L.R. Lebedev, I.A. Ryazankin et al. "Comparative study of variola virus and monkeypox virus interferon-gamma-binding" *Molecular Biology* 39 (6): (2005): 1055-1062.
- **324.** A.V. Atrasheuskaya, E.K. Bukin, T.M. Fredeking et al. "Protective effect of exogenous recombinant mouse interferon-gamma and tumour necrosis factor-alpha on ectromelia virus infection in susceptible BALB/c mice" *Clinical and Experimental Immunology* 136 (2): (2004): 207-214.
- **325.** I.V. Vinogradov, G.V. Kochneva, Ye.M. Malkova et al. "Comparative Study of Pathological Changes in Internal Organs of Mice in Infection Caused by Two Orthopoxviruses" *Bulletin of Experimental Biology and Medicine* 141 (4): (2006): 433-438.
- **326.** Galina Kochneva, Ilya Vinogradov, Elena Ryabchikova et al. "Study of the Morphologic Parameters of Different Orthopoxvirus Infection in Chick Embryos" *International Chemical and Biological Medical Treatment Symposium* 2006 (6): (2006): 55.
- **327.** Yelena Ivanovna Ryabchikova, I.V. Vinogradov et al. "Studies of Cowpox and Variola Virus Infection in vitro and in ovo" *International Conference "Development of International Collaboration in Infectious Disease Research"* (2004).
- **328.** I.V. Vinogradov, G.V. Kochneva, Ye.M. Malkova et al. "Intranasal infection in mice inoculated with cowpox virus strain EP-2 isolated from the elephant" *Problems of Virology* 50 (4): (2005): 37-42.
- **329.** A.V. Totmenin, I.V. Kolosova, S.N. Shchelkunov "Orthopoxvirus genes for Kelch-like proteins. I. Analysis of species specific differences by gene structure and organization" *Molecular Biology* 36 (4): (2002): 610.
- **330.** K.I. Chuykova, L.L. Katanakhova, S.V. Vozhakov et al. "Assessment of Clinical Indicators in Smallpox Vaccination Using "OspaVir" Inactivated Vaccine" *Russian Scientific and Practical Conference "Infectious Diseases: Problems of Public Health and Military Medicine"* (2006).
- **331.** T.A. Bektimirov, V.N. Podkuiko, L.V. Grigor'eva et al. "The comparative study of the safety of a vaccinal process in both oral and epicutaneous immunization against smallpox" *Problems of Virology* 47 (4): (2002): 9-13.
- **332.** S.A. Mel'nikov, V.N. Podkuiko, R.A. Khamitov et al. "Clinical studies of vaccine TEOVac under the conditions of remote revaccination" *Journal of Microbiology, Epidemiology and Immunobiology* 2005 (6): (2005): 29-33.
- **333.** Anonymous "Accidental exposure to smallpox vaccine, Russian Federation" *Weekly Epidemiological Record* 75 (25): (2000): 202.
- **334.** A.A. Sergeyev, A.N. Sergeyev, V.A. Petrishchenko et al. "Reactogenicity, Safety and Immunogenicity of a Recombinant Bivaccine Against Smallpox and Hepatitis B in Limited Clinical Trials" *Problems of Virology* 49 (5): (2004): 22-26.
- **335.** L.S. Sandakhchiyev, I.V. Plyasunov, V.N. Podkuiko "Clinical Trials of Oral Recombinant Bivaccine Against Variola and Hepatitis B during Double Vaccination" *Problems of Virology* 51 (2): (2006): 31.
- **336.** Anonymous "4647-cell culture for preparation of recombinant bivaccine against smallpox and hepatitis B" *Problems of Virology* 52 (2): (2007): 37-40.

- **337.** I.V. Plyasunov, A.N. Sergeev, A.A. Sergeev et al. "Reactogenicity, safety and immunogenecity of the "Revax-BT" vaccine against variola and hepatitis B after a single oral vaccination" *Russian Journal of Biotechnology* 2006 (3): (2006): 83-91.
- **338.** A.N. Sergeev, I.V. Plyasunov, A.A. Sergeev et al. "The study of reactogenicity, safety, and immunogenicity of recombinant smallpox-and hepatitis B vaccine for oral administration in humans" *Meetings of the WHO Advisory Committee on Variola Virus* (2006).
- **339.** Anonymous "The Development of the Cultivated Live Recombinant Bivaccine against Variola and Hepatitis B for Enteric Administration" *ISTC Projects database* (2003).
- **340.** S.N. Shchelkunov, A.Ye. Nesterov, I.A. Ryazankin et al. "Creation of a Live Candidate Polyvalent Vaccine Against Human Immunodeficiency Virus, Hepatitis Virus B and Orthopoxviruses" *Transactions of the Academy of Sciences* 390 (6): (2003): 834-838.
- **341.** V.N. Podkuyko, A.A. Vorobyev, V.A. Maksimov "Evaluation of the vaccinal process in skin-scarification and oral immunization of rabbits with live smallpox vaccines" *Journal of Microbiology, Epidemiology and Immunobiology* 2005 (5): (2005): 40-45.
- **342.** I.D. Konstantinova, N.A. Leont'eva, G.A. Galegov et al. "Ribavirin: Biotechnological Synthesis and Effect on the Reproduction of Vaccinia Virus" *Russian Journal of Bioorganic Chemistry* 30 (6): (2004): 553-560.
- **343.** Evgeny F. Belanov, N.I. Bormotov, S.M. Balakhnin et al. "An Update on the Results of Search for Antiviral Compounds for Treatment and Prevention of Orthopoxviral Infections" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- **344.** V.L. Rusinov, I.S. Kovalev, D.N. Kozhevnikov et al. "Synthesis and antiviral activity of 2-amino-3-ethoxycarbonylpyrazine derivatives" *Pharmaceutical Chemistry Journal* 39 (12): (2005): 12.
- **345.** Anonymous "A study of neutralizing activity and cross-reactivity of monoclonal antibodies to ecrtomelia virus with orthopoxviruses pathogenic for man" *Bulletin of the Russian Academy of Medical Sciences* 2004 (8): (2004): 19.
- **346.** I.A. Razumov, I.P. Gileva, M.A. Vasilyeva et al. "Fusion proteins encoded by orf 129L of ectromelia and orf A30L of smallpox viruses cross-react with neutralizing monoclonal antibodies" *Molecular Biology* 39 (6): (2005): 1046-1054.
- **347.** Orlova Elena Sergeyevna, Aleksey Vladimirovich Shcherbakov "Development of Methods for Diagnosis of Sheeppox and Goatpox Viruses" *Vector News. Archive. Defense of Dissertations* (2007): 1-23.
- **348.** Ye.S. Orlova, A.V. Scherbakov, A.S. Yakovleva et al. "P17, P18 and P32 Recombinant Proteins of Sheeppox Virus as Potential Antigens for Enzyme-Linked Immunosorbent Assay" *Collected Works of the Federal Center for Animal Health* 4 (2006).
- **349.** S.Zh. Tsybanov "Study of Biological and Molecular-Genetic Properties of Viruses and Prions for Potential Use in Development and Assessment of the Effectiveness of the Diagnosis Methods for Dangerous and Socially Significant Infections in Humans and Animals" *Federal Portal for Science and Innovation* (2005).
- **350.** Wang Yan, Ma Wenli, Zheng Wenling "Research on Smallpox and Other Orthopoxvirus Genome Structures" *Foreign Medical Sciences (Epidemiology, Lemology)* 29 (5): (2002): 263-265.
- **351.** Zhang Yanming, Wu Yingsong, Dong Wenqi et al. "Establishment of a BHK21 cell line stably expressing vaccinia virus D13L gene" *Journal of Tropical Medicine* 4 (3): (2004): 230-233.
- **352.** Hong Kunxue, Duan Danli, Li Haishan "The construction of attenuated Tiantan recombinant vaccinia virus vector with IFN-gamma receptor gene deletion" *Chinese Journal of Experimental and Clinical Virology* 18 (1): (2004): 43-46.
- **353.** Zhao Mei, Gao Yitong "Problems of Smallpox Recurrence Following Its Global Eradication" *Progress in Microbiology and Immunology* 34 (2): (2006): 46-47.
- **354.** Dong Xiaoman, Dong Yi, Chen Xiuzhen "Detection of the Potency of Smallpox Vaccine by Microtitration" *Progress in Microbiology and Immunology* 35 (1): (2007): 35-39.

- **355.** Fang Qing, Chen Zhiwei, Zhang Linqi "Study on the Biological Characteristics of Vaccinia Virus Tian Tan Strain and Construction of Its Genetically Engineered Attenuated Vector" *Wanfang Dissertations of China (DOC) Database* (2005).
- **356.** Fang Qing, Yang Lin, Zhu Weijun et al. "Host range, growth property, and virulence of the smallpox vaccine: vaccinia virus Tian Tan strain" *Virology* 335 (2): (2005): 242-251.
- **357.** Guo Qinyuan, An Huanyu, Fan Shulan et al. "Development of a plaque reduction neutralization test (PRNT) for determination of titer of anti-smallpox immunoglobulins" *Progress in Microbiology and Immunology* 34 (4): (2006): 37-39.
- **358.** Hu Sen, Wang Qinghua, Wang Xijun et al. "Expression of Nipah Virus Fusion Protein in Recombinant Vaccinia Virus and Assays of Its Biological Function" *Chinese Journal of Virology* 22 (3): (2006): 220-224.
- **359.** Yan Dongmei, Li Chunjiang, Meng Xiangdong et al. "Detection of mouse immunity induced by HIV transmembrane modified vaccine combined with poxvirus by ELISPOT and its specific CTL function" *Heilongjiang Medicine and Pharmacy* 30 (2): (2007): 7-8.
- **360.** Zhou Weimin, Tan Wenjie, Zheng Nan et al. "Rapid Detection and Differentiation of Smallpox or Monkeypox Virus Infection by Real-Time PCR Assay" *Letters in Biotechnology* 17 (5): (2006): 703-706.
- **361.** Zheng Guangyu, Ruan Li, Zheng Nan "Smallpox Virus-specific PCR Detection Technique" *Wanfang Dissertations of China (DOC) Database* (2004).
- **362.** Wang Yan, Ma Wenli "Study on Development of DNA Microarrays for Orthopoxvirus Detection" *Wanfang Dissertations of China (DOC) Database* (2004).
- **363.** Ma Wenli, W. Yan, W. Hongmin et al. "An oligonucleotide microarray for the detection of vaccinia virus" *British Journal of Biomedical Science* 61 (3): (2004): 142-143.
- **364.** An Xuefang, Hu Xueying, Yu Siyi et al. "Pathological Studies and Molecular Detection of Mousepox Virus" *CNKI China Proceedings of Conference Full-text Database* (2006).
- **365.** Xiao Jinhua, Xin Wen, Lin Yongjie et al. "Generation of linear expression constructs by one-step PCR with vaccinia DNA topoisomerase I" *Molecular Biotechnology* 35 (1): (2007): 15-22.
- **366.** Xing Ke, Deng Riqiang, Wang Jinwen et al. "Genome-based phylogeny of poxvirus" *Intervirology* 49 (4): (2006): 207-214.
- **367.** Xiang Wenhua, Guo Wei, Xue Fei "Construction of Goatpox Virus Gene Deleted Transfer Vector and Generation of Gene Deleted Mutant" *CNKI China Doctor/Master Dissertations Full-text Database* (2005).
- **368.** Cheng Xiaowen, Bao Endong, Bu Zhigao "Construction of Recombinant Goatpox Virus Expressing Green Fluorescence Protein and Expression of the G Protein of Bovine Ephemeral Fever Virus in the Recombinant Vaccinia Virus" *CNKI China Doctor/Master Dissertations Full-text Database* (2006).
- **369.** Liu Qi, Huang Xia, Guo Jiangang et al. "Isolation, identification and biological characteristics of goat poxvirus" *Chinese Journal of Preventive Veterinary Medicine* **28** (5): (2006): 494-498.
- **370.** Wang Fang, Chen Hongyan, Liu Shengwang "Molecular characterization of ORF64-ORF67 of goatpox virus vaccine strain AV41" *Chinese Journal of Preventive Veterinary Medicine* 29 (2): (2007): 96-102,111.
- **371.** Ju Houbin, Zhang Qiang, Zheng Haixue et al. "Construction of a Goatpox Virus Transfer Vector and Its Identification" *Chinese Journal of Veterinary Drug* 41 (2): (2007): 1-4, 9.
- **372.** Cheng Xiaowen, Hu Sen, Wang Xijun "Construction and Purification of Recombinant Goatpox Virus Expressing Exotic Green Fluorescence Protein" *Journal of Agricultural Biotechnology* 14 (5): (2006): 798-802.
- **373.** Zhou Bijun, He Guangzhi, Yu Tiande "Propagation of Goat Pox Virus and Preparation of Precipitation Antigen" *Journal of Wuhan University (Natural Science Edition)* 50 (Supplement 2): (2004): 53-56.

- **374.** Xu Chunzhi, Zhou Bijun, Yue Jun et al. "Cloning and sequence analysis of inverted terminal repeat from goatpox virus" *Animal Husbandry & Veterinary Medicine* 39 (7): (2007): 11-13.
- **375.** Zhou Bijun, He Guangzhi, Yu Tiande et al. "Propagation of Goatpox Virus and Preparation of Precipitation Antigen" *Journal of Mountain Agriculture and Biology* 23 (5): (2004): 400-403.
- **376.** Bi Xiaolin, Cao Ru, He Jianxin et al. "Functional and structural study of the prokaryotic enhancer-like element VV16 from vaccinia virus genome" *Chinese Journal of Biotechnology* 16 (3): (2000): 337-340.
- 377. An Huanyu, Fan Shulan, An Liying et al. "Large-scale Production of Smallpox Vaccine" *Chinese Journal of Biologicals* 19 (5): (2006): 497-498.
- **378.** An Xuefang, Liu Fengsong, Wang Hanzhong et al. "Morbidity and Epidemic Characteristics of Mousepox from an Outbreak in KM Mice" *Laboratory Animal and Comparative Medicine* 24 (4): (2004): 228-229, 256.
- **379.** Huang Xun, Xu Xiuhua "Monkeypox Virus and Its Prevention" *Chinese Journal of Infection Control* 4 (2): (2005): 186-188.
- **380.** Zhang Zheng, Yue Zhihong "Progress in Research on Monkeypox Virus" *Chinese Journal of Laboratory Medicine* 26 (8): (2003): 511-514.
- **381.** Liu Shanggao, Zha Hongbo, Wang Chunfeng "Survey of the Dangers of Monkeypox Virus for Humans and Animals" *Compilation of New Achievements in Research on the Prevention and Treatment of Zoonoses: Papers from the Sixth Academic Exchange on Zoonoses of the Livestock Infectious Diseases Branch of the Chinese Society of Animal Husbandry and Veterinary Medicine* (2004): 80-82.
- **382.** Guan Shiyun, Yin Minghong, Wang Wenhu "Clinical Practice and Prevention and Treatment of Monkeypox" *Journal of Hubei Medical Staff College* 17 (1): (2004): 91-93.
- **383.** Geeta Chaudhri, Vijay Panchanathan, R. Mark L. Buller et al. "Polarized type 1 cytokine response and cell-mediated immunity determine genetic resistance to mousepox" *Proceedings of the National Academy of Sciences of the United States of America* 101 (24): (2004): 9057-9062.
- **384.** S. Mahalingam, P.S. Foster, M. Lobigs et al. "Interferon-inducible chemokines and immunity to poxvirus infections" *Immunological Reviews* 177 (1): (2000): 127-133.
- **385.** Vijay Panchanathan, Geeta Chaudhri, Gunasegaran Karupiah "Interferon function is not required for recovery from a secondary poxvirus infection" *Proceedings of the National Academy of Sciences of the United States of America* 102 (36): (2005): 12921-12926.
- **386.** R.G. van der Most, B.W.S. Robinson, D.J. Nelson "Gene therapy for malignant mesothelioma: beyond the infant years" *Cancer Gene Therapy* 13 (10): (2006): 897-904.
- **387.** Geeta Chaudhri, Rosalind de Chazal, Geoff Sjollema et al. "A Critical Role for Tumour Necrosis Factor in the Recovery from Poxvirus Infections" *International Congress of Immunology and Annual Conference of FOCIS* 2004 (12): (2004).
- **388.** N.H.R. Hamilton, S. Mahalingam, J.L. Banyer et al. "A recombinant vaccinia virus encoding the interferon-inducible T-cellalpha chemoattractant is attenuated in vivo" *Scandinavian Journal of Immunology* 59 (3): (2004): 246-254.
- **389.** J.J. Jaunders, W.B. Dyer, M. Munier et al. "Transient CCR5+ CD38+++ CD4+ effector TH1 cells are an early component of the primary immune response to vaccinia and immediately precede IL-2+ memory CD4+ T cells" *Cytometry A* 69A (5): (2006): 400.
- **390.** I.E. Agranovski, A.S. Safatov, O.V. Pyankov et al. "Long Term Personal Sampling of Viable Airborne Viruses" *Journal of Aerosol Science* 35 (Supplement 1): (2004): 403-404.
- **391.** Chiu Wenling, Patricia Szajner, Bernard Moss et al. "Effects of a temperature sensitivity mutation in the J1R protein component of a complex required for vaccinia virus assembly" *Journal of Virology* 79 (13): (2005): 8046.

- **392.** Chung Chesheng, Huang Chengyen, Chang Wen "Vaccinia virus penetration requires cholesterol and results in specific viral envelope proteins associated with lipid rafts" *Journal of Virology* 79 (3): (2005): 1623-1634.
- **393.** Cheng Wenfang, Hung Chienfu, Sara I. Pai et al. "Repeated DNA vaccinations elicited qualitatively different cytotoxic T lymphocytes and improved protective antitumor effects" *Journal of Biomedical Science* 9 (6): (2002): 675-687.
- **394.** Kaku Hiroki "Smallpox: Key Considerations for a Bioterrorism Response Plan" *Bulletin of the Yamagata Medical Association* (631): (2004): 62-75.
- 395. Kurata Takeshi "Infectious Disease- The latest topic- Smallpox (variola)" Clinical Pediatrics 45 (4): (2004): 445-450
- **396.** Hatayama Shuji "Immunity to Smallpox" *Journal of Clinical and Experimental Medicine* 210 (7/8): (2004): 699-700
- **397.** Hatakeyama Shuji, Moriya Kyoji, Saijo Masayuki et al. "Persisting humoral antiviral immunity within the Japanese population after the discontinuation in 1976 of routine smallpox vaccinations" *Clinical and Diagnostic Laboratory Immunology* 12 (4): (2005): 520-524.
- **398.** Hatakeyama Shuji, Koike Kazuhiko, Morikawa Shigeru et al. "Conservation of Smallpox Immunity More Than 26 Years After the Cessation of Smallpox Vaccination Programs in Japan" *Journal of the Japanese Association for Infectious Diseases* 78 (8): (2004): 752-753.
- **399.** Morikawa Shigeru, Sakiyama Tokuki, Hasegawa Hideki et al. "An attenuated LC16m8 smallpox vaccine: Analysis of full-genome sequence and induction of immune protection" *Journal of Virology* 79 (18): (2005): 11873-11891.
- **400.** Gaener Rodger, Geoffrey L. Smith "Replacing the SCR domains of vaccinia virus protein B5R with EGFP causes a reduction in plaque size and actin tail formation but enveloped virions are still transported to the cell surface" *Journal of General Virology* 83 (2): (2002): 323-332.
- 401. Yamada Akio "Monkeypox" Journal of the Japan Medical Association 131 (2): (2004): 218-219.
- **402.** Saijo Masayuki "International Infectious Diseases: Monkeypox Virus" *Clinical Microbiology* 31 (1): (2004): 21-24
- **403.** Hitomi Shigemi "Monkeypox and B virus infection in humans" *Antibiotics and Chemotherapy* 22 (7): (2006): 1097-1102.
- **404.** Saijo Masayuki, Ami Yasushi, Suzaki Yuriko et al. "Analysis of the Virulence in Primates of Monkeypox Virus Zr-599 Strain (Congo Basin Type) and Liberia Strain (West-African Type)" *Program and Abstracts from the Annual Meeting of the Japanese Society for Virology* 54 (2006): 392b.
- **405.** Oliver Piskurek, Okada Norihiro "Poxviruses as possible vectors for horizontal transfer of retroposons from reptiles to mammals" *Proceedings of the National Academy of Sciences of the United States of America* 104 (29): (2007): 12046-12051.
- **406.** Sato Azusa, Kanehara Tomomi, Nagai Chigusa et al. "Establishment of a PRNT Assay Method for Smallpox Vaccines" *Program and Abstracts from the Annual Meeting of the Japanese Society for Virology* 54: (2006): 392.
- **407.** Izumi Yasuyuki, Ami Yasushi, Matsuo Kazuhiro et al. "Intravenous inoculation of replication-deficient recombinant vaccinia virus DIs expressing simian immunodeficiency virus gag controls highly pathogenic simianhuman immunodeficiency virus in monkeys" *Journal of Virology* 79 (5): (2005): 3229.
- **408.** Han Young Woo, Park Seong Ok, Kim A. Rum et al. "In Vivo CTL Activity Induced by Prime-boost Vaccination Using Recombinant Vaccinia Virus and DNA Vaccine Expressing Epitope Specific for CD8+ T Cells" *Journal of Bacteriology and Virology* 37 (1): (2007): 1-9.
- **409.** Shinoda Kaori, Xin Ke-Qin, Kojima Yoshitsugu et al. "Robust HIV-specific immune responses were induced by DNA vaccine prime followed by attenuated recombinant vaccinia virus (LC16m8 strain) boost" *Clinical Immunology* 119 (1): (2006): 32-37.

- **410.** Komano Jun, Miyauchi Kosuke, Matsuda Zene et al. "Inhibiting the Arp2/3 complex limits infection of both intracellular mature vaccinia virus and primate lentiviruses" *Molecular Biology of the Cell* 15 (12): (2004): 5197-5207.
- **411.** Izumi Yasuyukie, Ami Yasushi, Matsuo Kazuhiro et al. "Intravenous inoculation of replication-deficient recombinant vaccinia virus DIs expressing simian immunodeficiency virus Gag controls highly pathogenic simianhuman immunodeficiency virus in monkeys" *Journal of Virology* 77 (24): (2003): 13248-13256.
- **412.** Someya Kenji, Xin Ke-Qin, Matsuo Kazuhiro et al. "A consecutive priming-boosting vaccination of mice with simian immunodeficiency virus (SIV) gag/pol DNA and recombinant vaccinia virus strain DIs elicits effective anti-SIV immunity" *Journal of Virology* 78 (18): (2004): 9842-9853.
- **413.** Takeshi Mikami, Xuenan Xuan, Hideyuki Nagasawa et al. "Expression of canine interferon-gamma by a recombinant vaccinia virus and its antiviral effect" *Virus Research* 75 (2): (2001): 113-121.
- **414.** Kitamoto Noritoshi, Kobayashi Takayuki, Kato Yoji et al. "Preparation of Monoclonal Antibodies Cross-Reactive with Orthopoxviruses and Their Application for Direct Immunofluorescence Test" *Microbiology and Immunology* 49 (3): (2005): 219-225.
- **415.** Kim S.H., Yeo S.G., Park K.H. et al. "The persistence of humoral and cellular immunities more than three decades after smallpox vaccination" *Clinical Microbiology and Infection* 13 (1): (2007): 91-93.
- **416.** Kim Sung-Han, Yeo Sang-Gu, Cho Jae-Hyun et al. "Cell-mediated immune responses to smallpox vaccination" *Clinical and Vaccine Immunology* 13 (10): (2006): 1172-1174.
- **417.** Kim Sung-Han, Yeo Sang-Gu, Jang Hee-Chang et al. "Clinical responses to smallpox vaccine in vaccinia -naive and previously vaccinated populations: Undiluted and diluted lancy-vaxina vaccine in a single-blind, randomized, prospective trial" *Journal of Infectious Diseases* 192 (6): (2005): 1066-1070.
- **418.** Kang Tae-Hong, Kim Kyong-Tai "Negative regulation of ERK activity by VRK3-mediated activation of VHR phosphatase" *Nature Cell Biology* 8 (8): (2006): 863-869.
- **419.** Yoo Myung-Sook, Choi Sang-Yun "Vaccinia virus regulates transcription rate by controlling level of transcription factors" *Conference Proceedings of Korean Society for Biochemistry and Molecular Biology Annual Meeting* 2005 (1): (2005): 158.
- **420.** Kwon Soo-Young, Choi Yoon-Joo, Kang Tae-Hong et al. "Highly efficient protein expression and purification using bacterial hemoglobin fusion vector" *Plasmid* 53 (3): (2005): 274-282.
- **421.** Kang Tae Hong, Kim Kyong Tai "Apoptosis and Cell Cycle Oncogenes and Tumor Suppressor Genes: Functional characterization of Vaccinia-related Kinase-1 as a Chromatin Condenser" *Conference Proceedings of Korean Society for Biochemistry and Molecular Biology Annual Meeting* 2004 (1): (2004): 232.
- **422.** Kim J.H., Oh J.Y., Park B.H. et al. "Systemic armed oncolytic and immunologic therapy for cancer with JX-594, a targeted poxvirus expressing GM-CSF" *Molecular Therapy (2004)* 14 (3): (2006): 361-370.
- **423.** Sung Chul Ha, Neratur K. Lokanath, Dong Van Quyen et al. "A poxvirus protein forms a complex with left-handed Z-DNA: Crystal structure of a Yatapoxvirus Zα bound to DNA" *Proceedings of the National Academy of Sciences of the United States of America* 101 (40): (2004): 14367-14372.
- **424.** Shukriti Sharma, B. Bansal, K.B. Singh et al. "Clinico-epidemiological observations on outbreaks of cowpox and buffalo pox" *Indian Journal of Veterinary Medicine* 26 (1): (2006): 47-48.
- 425. S. Ratnam, M. Pal "Pseudocowpox and human infection" Veterinary World 4 (10): (2006): 293-294.
- **426.** Kumar Sanjeev, Seth Pradeep "Immunogenicity of recombinant modified vaccinia Ankara viruses (rMVA) expressing HIV-1 indian subtype C gag-protease and env-gp120 genes in mice" *Viral Immunology* 17 (4): (2004): 574-579.
- **427.** H. Erdem, A. Ozqun, A. Pahsa "Increasing infectious worries: Biological weapons" *Syndrome* 13 (12): (2001): 106-110.

- **428.** Nadav Orr, Moshe Forman, Marcus Hadar et al. "Clinical and immune responses after revaccination of Israeli adults with the Lister strain of vaccinia virus" *Journal of Infectious Diseases* 190 (7): (2004): 1295-1302.
- **429.** E. Tzahar, J.D. Moyer, H. Waterman et al. "Pathogenic poxviruses reveal viral strategies to exploit the ErbB signaling network" *The EMBO Journal* 17 (20): (1998): 5948-5963.
- **430.** Wallin Arune, Luksiene Zivile, Zagminas Kestutis et al. "Public health and bioterrorism: renewed threat of anthrax and smallpox" *Medical Research* 43 (4): (2007): 278-284.
- **431.** Z. Humlova, M. Vokurka, M. Esteban et al. "Vaccinia virus induces apoptosis of infected macrophages" *Journal of General Virology* 83 (2002): (2002): 2821-2832.
- **432.** Caroline Gubser, Daniele Bergamaschi, Michael Hollinshead et al. "A new inhibitor of apoptosis from vaccinia virus and eukaryotes" *PLoS Pathogens* 3 (2): (2007): 246-259.
- **433.** Z. Jezek, B. Kriz, V. Rothbauer "Camelpox and its risk to the human population" *Journal of Hygiene Epidemiology Microbiology and Immunology* 27 (1): (1983): 29-42.
- **434.** Iveta Vancova, Peter Kontsek, Claude La Bonnardiere "Vaccinia virus protein B18R inhibits the activity and cellular binding of the novel type interferon-delta" *Journal of General Virology* 79 (7): (1998): 1647-1649.
- **435.** Kinga Slezak, Marta Michalik, Aleksandra Kowalczyk et al. "YY1 is recruited to the cytoplasm of vaccinia virus-infected human macrophages by the Crm1 system" *Virus Research* 102 (2): (2004): 177-184.
- **436.** A. Kowalczyk, K. Guzik, K. Slezak et al. "Heat shock protein and heat shock factor 1 expression and localization in vaccinia virus infected human monocyte derived macrophages" *Journal of Inflammation* 2 (1): (2005): 12.
- **437.** Katarzyna A. Pirog, Aleksandra K. Kowalczyk, Hanna B. Rokita et al. "Changes in Bcl-2 Expression in Vaccinia Virus-Infected Human Peripheral Blood Monocytes" *Viral Immunology* 18 (1): (2005): 224-231.
- **438.** H. Rokita, W. Branicki, D. Wronska et al. "Vaccinia virus-induced changes in cytokine-regulated acute phase plasma protein synthesis by hepatoma cells" *Biochemistry and Molecular Biology International* 44 (6): (1998): 1093-1104.
- **439.** Nada Kuljic-Kapulica "Smallpox—in the past or not?" *Serbian Archives of General Medicine* 132 (7-8): (2004): 272-276.
- **440.** Mimi Remichkova, Nikolay Petrov, Angel S. Galabov "Synergistic combination effect of cidofovir and idoxuridine on vaccinia virus replication" *Antiviral Chemistry and Chemotherapy* 17 (2): (2006): 53-58.
- **441.** M. Magdalena Gherardi, Eva Perez-Jimenez, Jose Luis Najera et al. "Induction of HIV Immunity in the Genital Tract After Intranasal Delivery of a MVA Vector: Enhanced Immunogenicity After DNA Prime-Modified Vaccinia Virus Ankara Boost Immunization Schedule" *Journal of Immunology* 172 (10): (2004): 6209-6220.
- **442.** J.C. Gallego-Gomez, C. Risco, D. Rodriguez et al. "Differences in Virus-Induced Cell Morphology and in Virus Maturation between MVA and Other Strains (WR, Ankara, and NYCBH) of Vaccinia Virusin Infected Human Cells" *Journal of Virology* 77 (19): (2003): 10606-10622.
- **443.** Susana Guerra, Luis A. Lopez-Fernandez, Raquel Conde et al. "Microarray analysis reveals characteristic changes of host cell gene expression in response to attenuated modified vaccinia virus Ankara infection of human HeLa cells" *Journal of Virology* 78 (11): (2004): 5820-5834.
- **444.** M.M. Gherardi, J.L. Najera, E. Perez-Jimenez et al. "Prime-Boost Immunization Schedules Based on Influenza Virus and Vaccinia Virus Vectors Potentiate Cellular Immune Responses against Human Immunodeficiency Virus Env Protein Systemically and in the Genitorectal Draining Lymph Nodes" *Journal of Virology* 77 (12): (2003): 7048-7057.
- **445.** Juana M. Sanchez-Puig, Rafael Blasco "Isolation of vaccinia MVA recombinants using the viral F13L gene as the selective marker" *BioTechniques* 39 (5): (2005): 665-674.
- **446.** M.M. Gherardi, M. Esteban "Mucosal and systemic immune responses induced after oral delivery of vaccinia virus recombinants" *Vaccine* 17 (9-10): (1999): 1074-1083.

- **447.** Carmen Elena Gomez, Jose Luis Najera, Elena Domingo-Gil et al. "Virus distribution of the attenuated MVA and NYVAC poxvirus strains in mice" *Journal of General Virology* 88 (6): (2007): 1.
- **448.** M.P. Sanchez-Seco, L. Hernandez, J.M. Eiros et al. "Detection and identification of orthopoxviruses using a generic nested PCR followed by sequencing" *British Journal of Biomedical Science* 63 (2): (2006): 79-85.
- **449.** Maria M. Lorenzo, Rafael Immaculada Galindo "Construction and isolation of recombinant vaccinia virus using genetic markers" *Methods in Molecular Biology* 269: (2004): 15-30.
- **450.** S. Guerra, L.A. Lopez-Fernandez, A. Pascual et al. "Cellular Gene Expression Survey of Vaccinia Virus Infection of Human HeLa Cells" *Journal of Virology* 77 (11): (2003): 6493-6506.
- **451.** Claudio R. Santos, Francisco M. Vega, Sandra Blanco et al. "The vaccinia virus B1R kinase induces p53 down regulation by an Mdm2-dependent mechanism" *Virology* 328 (2): (2004): 254-265.
- **452.** Susana Guerra, Miguel Aracil, Raquel Conde et al. "Wiskott-Aldrich syndrome protein is needed for vaccinia virus pathogenesis" *Journal of Virology* 79 (4): (2005): 2133-2140.
- **453.** Rafael Blasco Lozano, Beatriz Perdiguero de la Torre "Analysis of Interactions Between Envelope Proteins of Vaccinia Virus" *TESEO Database* (2003).
- **454.** J.M. Sanchez-Puig, L. Sanchez, G. Roy et al. "Susceptibility of different leukocyte cell types to Vaccinia virus infection" *Virology Journal* 1 (1): (2004): 10.
- **455.** M. Di Nicola, C. Carlo-Stella, A. Anichini et al. "Immunization of Patients with Malignant Melanoma with Autologous CD34<sup>+</sup> Cell-Derived Dendritic Cells Transduced Ex Vivo with aRecombinant Replication-Deficient Vaccinia Vector Encoding the Human Tyrosinase Gene: A Phase I Trial" *Human Gene Therapy* 14 (14): (2003): 1347-1360.
- **456.** Massimo Di Nicola, Carmelo Carlo-Stella, Roberta Baldassari et al. "Boosting T Cell-Mediated Immunity to Tyrosinase by Vaccinia Virus-Transduced, CD34+-Derived Dendritic Cell Vaccination: A Phase I Trial in Metastatic Melanoma" *Clinical Cancer Research* (2004) 10 (16): (2004): 5381-5390.
- **457.** Sara Muraro, Attilio Bondanza, Matteo Bellone et al. "Molecular modification of idiotypes from B-cell lymphomas for expression in mature dendritic cells as a strategy to induce tumor-reactive CD4+ and CD8+ T-cell responses" *Blood* 105 (9): (2005): 3596-3604.
- **458.** Fabrizio Poccia, Cristiana Gioia, Carla Montesano et al. "Flow cytometry and T-cell response monitoring after smallpox vaccination" *Emerging Infectious Diseases* 9 (11): (2003): 1468-1470.
- **459.** T. Storni, C. Ruedl, K. Schwendener et al. "Nonmethylated CG Motifs Packaged into Virus-Like Particles InduceProtective Cytotoxic T Cell Responses in the Absence of Systemic Side Effects" *Journal of Immunology* 172 (3): (2004): 1777-1785.
- **460.** P. Zajac, D. Oertli, W. Marti et al. "Phase I/II clinical trial of a nonreplicative vaccinia virus expressing multiple HLA-A0201-restricted tumor-associated epitopes and costimulatory molecules in metastatic melanoma patients" *Human Gene Therapy* 14 (16): (2003): 1497-1510.
- **461.** Chantal Feder-Mengus, Elke Schultz-Thater, Daniel Oertli et al. "Nonreplicating recombinant vaccinia virus expressing CD40 ligand enhances APC capacity to stimulate specific CD4+ and CD8+ T cell responses" *Human Gene Therapy* 16 (3): (2005): 348-360.
- **462.** Isabelle Leparc-Goffart, Bertrand Poirier, Daniel Garin et al. "Standardization of a neutralizing anti-vaccinia antibodies titration method: an essential step for titration of vaccinia immunoglobulins and smallpox vaccines evaluation" *Journal of Clinical Virology* 32 (1): (2005): 47-52.
- **463.** Bertrand Poirier, Isabelle Leparc-Goffart, Jean-Marc Crance et al. "Establishment of a national network of validated and qualified laboratories for neutralizing anti-vaccinia antibodies titration" *Biologicals* 33 (4): (2005): 277-282.
- **464.** A. Ferrier, D. Garin, J.M. Crance "Rapid inactivation of vaccinia virus in suspension and dried on surfaces" *Journal of Hospital Infection* 57 (1): (2004): 73-79.

- **465.** I. Leparc-Goffart, B. Poirier, A. el Zaouk et al. "New Generation of Cell Culture Assay for Smallpox Vaccine Potency" *Journal of Clinical Microbiology* 41 (8): (2003): 3687-3689.
- **466.** A. Ferrier Rembert, J.M. Crance, B. Virgone et al. "Evaluation of 1st, 2nd and 3rd Generation Smallpox Vaccines" *The 1st Joint Meeting of European National Societies of Immunology, 16th European Congress of Immunology* (2006).
- **467.** H. Hillaireau, T. Le Doan, M. Besnard et al. "Encapsulation of antiviral nucleotide analogues azidothymidine-triphosphate and cidofovir in poly(iso-butylcyanoacrylate) nanocapsules" *International Journal of Pharmaceutics* 324 (1): (2006): 37-42.
- 468. J. Raulin "Development in lipid drugs" Mini Reviews in Medicinal Chemistry 5 (5): (2005): 489-498.
- **469.** Sebastien Hantz, Detlef Michel, Anne-Marie Fillet et al. "Early selection of a new UL97 mutant with a severe defect of ganciclovir phosphorylation after valaciclovir prophylaxis and short-term ganciclovir therapy in a renal transplant recipient" *Antimicrobial Agents & Chemotherapy* 49 (4): (2005): 1580-1583.
- **470.** Remy N. Charrel, Bernard La Scola, Didier Raoult "Multi-pathogens sequence containing plasmids as positive controls for universal detection of potential agents of bioterrorism" *BMC Microbiology* 4 (1): (2004): 21-32.
- **471.** Natale Scaramozzino, Audrey Ferrier-Rembert, Anne-Laure Favier et al. "Real-time PCR to identify variola virus or other human pathogenic orthopox viruses" *Clinical Chemistry* 53 (4): (2007): 606-613.
- **472.** Anne-Laure Puaux, Delphine Marsac, Stephane Prost et al. "Efficient priming of simian/human immunodeficiency virus (SHIV)-specific T-cell responses with DNA encoding hybrid SHIV/hepatitis B surface antigen particles" *Vaccine* 22 (27-28): (2004): 3535-3545.
- **473.** Margaret Liu, Bruce Acres, Jean-Marc Balloul et al. "Gene-based vaccines and immunotherapeutics" *Proceedings of the National Academy of Sciences of the United States of America* 101 (2): (2004): 14567-14571.
- **474.** F. Cliquet, M. Aubert "Elimination of terrestrial rabies in Western European countries" *Developments in Biologicals* 119 (2004): (2004): 185-204.
- **475.** Jonathan D. Turner "A bioinformatic approach to the identification of bacterial proteins interacting with Toll-interleukin 1 receptor-resistance (TIR) homology domains" *FEMS Immunology and Medical Microbiology* 37 (1): (2003): 13-21.
- **476.** Natale Scaramozzino, Guenhael Sanz, Jean Marc Crance et al. "Characterization of the substrate specificity of homogeneous vaccinia virus uracil-DNA glycosylase" *Nucleic Acids Research* 31 (16): (2003): 4950-4957.
- **477.** A. Ferrier Rembert, D. Garin, B. Virgone et al. "Essential Role of CD4+ T-Cells in Natural Protection against Cowpox Virus Intranasal Infection" *The 1st Joint Meeting of European National Societies of Immunology, 16th European Congress of Immunology* (2006).
- **478.** Alexander Kofman, Marcus Graf, Ludwid Deml et al. "Codon usage-mediated inhibition of HIV-1 gag expression in mammalian cells occurs independently of translation" *Cytology* 45 (1): (2003): 94-100.
- **479.** Andreas Nitsche, Andreas Kurth, Anna Dunkhorst et al. "One-step selection of Vaccinia virus-binding DNA aptamers by MonoLEX" *BMC Biotechnology* 7 (1): (2007): 48.
- **480.** K. Matz-Rensing, H. Ellerbrok, B. Ehlers et al. "Fatal poxvirus outbreak in a colony of New World monkeys" *Veterinary Pathology (2004)* 43 (2): (2006): 212-218.
- **481.** H. Meyer, A. Nitsche "Evaluation of published real-time PCR assays, designed to differentiate and identify variola virus, using a panel of non-variola orthopoxviruses" *Meetings of the WHO Advisory Committee on Variola Virus* (2005).
- **482.** M. Stemmler, H. Neubauer, H. Meyer "Comparison of closely related orthopoxvirus isolates by random amplified polymorphic DNA and restriction fragment length polymorphism analysis" *Journal of Veterinary Medicine Series B* 48 (9): (2001): 647-654.

- **483.** Silvia Palacios, Laurent H. Perez, Sonja Welsch et al. "Quantitative SUMO-1 modification of a vaccinia virus protein is required for its specific localization and prevents its self-association" *Molecular Biology of the Cell* 16 (6): (2005): 2822-2835.
- **484.** Caroline Staib, Marianne Lowel, Volker Erfle et al. "Improved host range selection for recombinant modified vaccinia virus Ankara" *BioTechniques* 34 (4): (2003): 694-696, 698, 700.
- **485.** Antonio Cosma, Silja Buehler, Rashmi Nagaraj et al. "Neutralization assay using a modified vaccinia virus Ankara vector expressing the green fluorescent protein is a high-throughput method tomonitor the humoral immune response against vaccinia virus" *Clinical and Diagnostic Laboratory Immunology* 11 (2): (2004): 406-410.
- **486.** Andreas Wollenberg, Stefanie Wetzel, Walter H.C. Burgdorf et al. "Viral infections in atopic dermatitis: Pathogenic aspects and clinical management" *Journal of Allergy and Clinical Immunology* 112 (4): (2003): 667-674.
- **487.** Nicole Stolte-Leeb, Ulrike Sauermann, Stephen Norley et al. "Sustained conservation of CD4(+) T cells in multiprotein triple modality-immunized rhesus macaques after intrarectal challenge with Simian Immunodeficiency Virus" *Viral Immunology* 19 (3): (2006): 448-457.
- **488.** Martin Sapp, Selinka Hans-Christoph "Pseudovirions as specific tools for investigation of virus interactions with cells" *Methods in Molecular Biology* 292: (2005): 197-211.
- **489.** Caroline Staib, Ingo Drexler, Gerd Sutter "Construction and isolation of recombinant MVA" *Methods in Molecular Biology* 269: (2004): 77-99.
- **490.** Doerte Ortmann, Juergen Hausmann, Felix Beuschlein et al. "Steroidogenic acute regulatory (StAR)-directed immunotherapy protectsagainst tumor growth of StAR-expressing Sp2-0 cells in a rodentadrenocortical carcinoma model" *Endocrinology* 145 (4): (2004): 1760-1766.
- **491.** S. Hornemann, O. Harlin, C. Staib et al. "Replication of Modified Vaccinia Virus Ankara in Primary Chicken Embryo Fibroblasts Requires Expression of the Interferon Resistance Gene E3L" *Journal of Virology* 77 (15): (2003): 8394-8407.
- **492.** Bernd Krone, Klaus F. Kolmel, Beate M. Henz et al. "Protection against melanoma by vaccination with Bacille Calmette-Guerin (BCG) and/or vaccinia: an epidemiology-based hypothesis on the nature of a melanoma risk factor and its immunological control" *European Journal of Cancer* 41 (1): (2005): 104-117.
- **493.** A. Ploubidou, V. Moreau, K. Ashman et al. "Vaccinia virus infection disrupts microtubule organization and centrosome function" *The EMBO Journal* 19 (5): (2000): 3932-3944.
- **494.** Holger Ludwig, Joerg Mages, Caroline Staib et al. "Role of viral factor E3L in modified vaccinia virus Ankara infection of human HeLa cells: Regulation of the virus life cycle and identification of differentially expressed host genes" *Journal of Virology* 79 (4): (2005): 2584-2596.
- **495.** Jan D. Kahmann, Diana A. Wecking, Vera Putter et al. "The solution structure of the N-terminal domain of E3L shows a tyrosine conformation that may explain its reduced affinity to Z-DNA in vitro" *Proceedings of the National Academy of Sciences of the United States of America* 101 (9): (2004): 2712-2717.
- **496.** Heike Schwarz, Olof Harlin, Annette Ohnemus et al. "Synthesis of IFN-beta by virus-infected chicken embryo cells demonstrated with specific antisera and a new bioassay" *Journal of Interferon & Cytokine Research* 24 (3): (2004): 179-184.
- **497.** Andreas Sauerbrei, Peter Wutzler "Final report about the development of selective antiviral substances and their testing regarding the efficacy against vaccinia virus and cowpox virus in vitro and in ovo" *Military Medical Research Report* 2006 (5): (2006): 1-62.
- **498.** Hiroshi Nishiura, Martin Eichner "Infectiousness of smallpox relative to disease age: estimates based on transmission network and incubation period" *Epidemiology and Infection* 135 (7): (2007): 1145-1150.
- **499.** C. Gubser, D. Bergamaschi, M. Hollinshead et al. "A camelpox virus protein that modulates apoptosis and interacts with Bcl-2" *Euroconference on Apoptosis* 11: (2003).

- **500.** A. Postigo, J.R. Cross, J. Downward et al. "Interaction of F1L with the BH3 domain of Bak is responsible for inhibiting vaccinia-induced apoptosis" *Cell Death and Differentiation* 13 (10): (2006): 1651-1662.
- **501.** Timothy P. Newsome, Ina Weisswange, Friedrich Frischknecht et al. "Abl collaborates with Src family kinases to stimulate actin-based motility of vaccinia virus" *Cellular Microbiology* 8 (2): (2006): 233-241.
- **502.** Samantha Cooray, Mohammad W. Bahar, Nicola G.A. Abrescia et al. "Functional and structural studies of the vaccinia virus virulence factor N1 reveal a Bcl-2-like anti-apoptotic protein" *Journal of General Virology* 88 (6): (2007): 1656-1666.
- **503.** Nathan W. Bartlett, Karen Buttigieg, Sergei V. Kotenko "Murine Interferon Lambdas (Type III Interferons) Exhibit Potent Antiviral Activity In Vivo In a Poxvirus Infection Model" *Journal of General Virology* 86 (6): (2005): 1589-1596.
- **504.** Nathalie Jacobs, Ron A.J. Chen, Caroline Gubser et al. "Intradermal immune response after infection with Vaccinia virus" *Journal of General Virology* 87 (5): (2006): 1157-1161.
- **505.** Philippa M. Beard, Graham C. Froggatt, Geoffrey L. Smith et al. "Vaccinia virus kelch protein A55 is a 64 kDa intracellular factor that affects virus-induced cytopathic effect and the outcome of infection in a murine intradermal model" *Journal of General Virology* 87 (5): (2006): 1521-1529.
- **506.** A.J. Chen Ron, Nathalie Jacobs, Geoffrey L. Smith "Vaccinia virus strain Western Reserve protein B14 is an intracellular virulence factor" *Journal of General Virology* 87 (6): (2006): 1451-1458.
- **507.** Caroline Gubser, Daniele Bergamaschi, Michael Hollinshead et al. "Vaccinia virus encodes a novel apoptosis inhibitor that localises to the Golgi and contributes to virus virulence" *Euroconference on Apoptosis* 12: (2004): L-30.
- **508.** M.P.H. Stumpf, H.M. Wilkinson-Herbots "Allelic histories: Positive selection on a HIV-resistance allele" *Trends in Ecology and Evolution* 19 (4): (2004): 166-168.
- **509.** Geoffrey L. Smith, Mansun Law "The exit of vaccinia virus from infected cells" *Virus Research* 106 (2): (2004): 189
- **510.** Esteban Herrero-Martinez, Kim L. Roberts, Michael Hollinshead "Vaccinia virus intracellular enveloped virions move to the cell periphery on microtubules in the absence of the A36R protein" *Journal of General Virology* 86 (11): (2005): 2961-2968.
- **511.** Geoffrey L. Smith, Alain Vanderplasschen, Mansun Law "The formation and function of extracellular enveloped vaccinia virus" *Journal of General Virology* 83: (2002): 2915-2931.
- **512.** David C. Tscharke, Patrick C. Reading et al. "Dermal infection with vaccinia virus reveals roles for virus proteins not seen using other inoculation routes" *Journal of General Virology* 83 (2002): (2002): 1977-1986.
- **513.** Jeremy D. Gardner, David C. Tscharke, Patrick C. Reading et al. "Vaccinia virus semaphorin A39R is a 50-55 kDa secreted glycoprotein that affects the outcome of infection in a murine intradermal model" *Journal of General Virology* 82 (9): (2001): 2083-2093.
- **514.** Kamel El Omari, Nicola Solaroli, Anna Karlson et al. "Structure of vaccinia virus thymidine kinase in complex with dTTP: insights for drug design" *BMC Biochemistry* 6 (1): (2006): 22.
- **515.** Claire Simons, Qinpei Wu, Thet Thet Htar "Recent advances in antiviral nucleoside and nucleotide therapeutics" *Current Topics in Medicinal Chemistry* 5 (13): (2005): 1191-1203.
- **516.** Hannah L. Maslen, David Hughes, Mike Hursthouse et al. "6-Azapyrimidine-2'-deoxy-4'-thionucleosides: Antiviral Agents against TK+ and TK-HSV and VZV Strains" *Journal of Medicinal Chemistry* 47 (22): (2004): 5482-5491.
- **517.** Cressida Auckland, Alexandra Cowlishaw, Dilys Morgan et al. "Reactions to small pox vaccine in naive and previously-vaccinated individuals" *Vaccine* 23 (32): (2005): 4185-4187.
- **518.** A.L. Phelps, A.J. Gates, M. Hillier et al. "Comparative efficacy of modified vaccinia Ankara (MVA) as a potential replacement smallpox vaccine" *Vaccine* 25 (1): (2007): 34-42.

- **519.** A.L. Phelps, A.J. Gates, M. Hillier et al. "Comparative efficacy of replicating smallpox vaccine strains in a Murine Challenge Model" *Vaccine* 23 (27): (2005): 3500-3507.
- **520.** D.J. Pulford, A. Gates, S.H. Bridge et al. "Differential efficacy of vaccinia virus envelope proteins administered by DNA immunisation in protection of BALB/c mice from a lethal intranasal poxvirus challenge" *Vaccine* 22 (25-26): (2004): 3358-3368.
- **521.** Mansun Law, Mike M. Putz, Geoffrey L. Smith "An investigation of the therapeutic value of vaccinia-immune IgG in a mouse pneumonia model" *Journal of General Virology* 86 (4): (2005): 991-1000.
- **522.** A.C. Moore, A.V.S. Hill "Progress in DNA-based heterologous prime-boost immunization strategies for malaria" *Immunological Reviews* 199 (1): (2004): 126-143.
- **523.** N. Berry, R. Stebbings, S. Brown et al. "Immunological responses and viral modulatory effects of vaccination with recombinant modified vaccinia virus Ankara (rMVA) expressing structural and regulatory transgenes of simian immunodeficiency virus (SIVmac32H/J5M)" *Journal of Medical Primatology* 36 (2): (2007): 80-94.
- **524.** Richard Harrop, Matthew G. Ryan, Kevin A. Myers et al. "Active treatment of murine tumors with a highly attenuated vaccinia virus expressing the tumor associated antigen 5T4 (TroVax) is CD4(+) T cell dependent and antibody mediated" *Cancer Immunology and Immunotherapy* 55 (9): (2006): 1081-1090.
- **525.** Eung-Jun Im, Joseph P. Nkolola, Kati di Gleria et al. "Induction of long-lasting multi-specific CD8+ T cells by a four-component DNA-MVA/HIVA-RENTA candidate HIV-1 vaccine in rhesus macaques" *European Journal of Immunology* 36 (10): (2006): 2574-2584.
- **526.** Beatrice O. Ondondo, Hongbing Yang, Tao Dong et al. "Immunisation with recombinant modified vaccinia virus Ankara expressing HIV-1 gag in HIV-1-infected subjects stimulates broad functional CD4+ T cell responses" *European Journal of Immunology* 36 (10): (2006): 2585-2594.
- **527.** Paul Britton, Sharon Evans, Brian Dove et al. "Generation of a recombinant avian coronavirus infectious bronchitis virus using transient dominant selection" *Journal of Virological Methods* 123 (2): (2005): 203-211.
- **528.** Eung-Jun Im, Tomas Hanke "MVA as a vector for vaccines against HIV-1" *Expert Review of Vaccines* 3 (4, Supplement 1): (2004): S89-S97.
- **529.** J.P. Nkolola, E.G-T. Wee, E-J. Im et al. "Engineering RENTA, a DNA prime-MVA boost HIV vaccine tailored for Eastern and Central Africa" *Gene Therapy* 11 (13): (2004): 1068-1080.
- **530.** Matilu Mwau, Inese Cebere, Julian Sutton et al. "A human immunodeficiency virus 1 (HIV-1) clade A vaccine in clinicaltrials: stimulation of HIV-specific T-cell responses by DNA and recombinant modified vaccinia virus Ankara (MVA) vaccines in humans" *Journal of General Virology* 85 (4): (2004): 911-919.
- **531.** Tomas Hanke, Andrew J. McMichael, Lucy Dorrell "Clinical experience with plasmid DNA- and modified vaccinia virus Ankara-vectored human immunodeficiency virus type 1 clade A vaccine focusing on T-cell induction" *Journal of General Virology* 88 (1): (2007): 1-12.
- **532.** Helen McShane, Ansar A. Pathan, R. Sander Clare et al. "Boosting BCG with MVA85A: the first candidate subunit vaccine for tuberculosis in clinical trials" *Tuberculosis* 85 (1-2): (2005): 47-52.
- **533.** E.J. Davidson, R.L. Faulkner, P. Sehrv et al. "Effect of TA-CIN (HPV 16 L2E6E7) booster immunization in vulval intraepithelial neoplasia patients previously vaccinated with TA-HPV (vaccinia virus encoding HPV 16/18 E6E7)" *Vaccine* 22 (21-22): (2004): 2722-2729.
- **534.** Shahriar Behboudi, Anne Moore, Sarah C. Gilbert et al. "Dendritic cells infected by recombinant modified vaccinia virus Ankara retain immunogenicity in vivo despite in vitro dysfunction" *Vaccine* 22 (31-32): (2004): 4326-4331.
- **535.** Tomas Hanke, Andrew J. McMichael, Michael J. Dennis et al. "Biodistribution and persistence of an MVA-vectored candidate HIV vaccine in SIV-infected rhesus macaques and SCID mice" *Vaccine* 23 (12): (2005): 1507-1514.

- **536.** Wieslawa Olszewska, Yasemin Suezer, Gerd Sutter et al. "Protective and disease-enhancing immune responses induced by recombinant modified vaccinia Ankara (MVA) expressing respiratory syncytial virus proteins" *Vaccine* 23 (2): (2004): 215-221.
- **537.** R. Paillot, S.A. Ellis, J.M. Daly et al. "Characterisation of CTL and IFN-gamma synthesis in ponies following vaccination with a NYVAC-based construct coding for EHV-1 immediate early gene, followed by challenge infection" *Vaccine* 24 (10): (2006): 1490-1500.
- **538.** D.P. Webster, S. Dunachie, S. McConkey et al. "Safety of recombinant fowlpox strain FP9 and modified vaccinia virus Ankara vaccines against liver-stage P. falciparum malaria in non-immune volunteers" *Vaccine* 24 (15): (2006): 3026-3034.
- **539.** Sarah C. Gilbert, Vasee S. Moorthy, Laura Andrews et al. "Synergistic DNA-MVA prime-boost vaccination regimes for malaria and tuberculosis" *Vaccine* 24 (21): (2006): 4554-4561.
- **540.** Inese Cebere, Lucy Dorrell, Helen McShane et al. "Phase I clinical trial safety of DNA- and modified virus Ankara-vectored human immunodeficiency virus type 1 (HIV-1) vaccines administered alone and in a prime-boost regime to healthy HIV-1-uninfected volunteers" *Vaccine* 24 (4): (2006): 417-425.
- **541.** Barry S. Peters, Walter Jaoko, Eftyhia Vardas et al. "Studies of a prophylactic HIV-1 vaccine candidate based on modified vaccinia virus Ankara (MVA) with and without DNA priming: effects of dosage and route on safety and immunogenicity" *Vaccine* 25 (11): (2007): 2120-2127.
- **542.** Lucy Dorrell, Patricia Williams, Annie Suttill et al. "Safety and tolerability of recombinant modified vaccinia virus Ankara expressing an HIV-1 gag/multiepitope immunogen (MVA.HIVA) in HIV-1-infected persons receiving combination antiretroviral therapy" *Vaccine* 25 (17): (2007): 3277-3283.
- **543.** Caroline L. Smith, P. Rod Dunbar, Fareed Mirza et al. "Recombinant modified vaccinia ankara primes functionally activated CTL specific for a melanoma tumor antigen epitope in melanoma patients with a high risk of disease recurrence" *International Journal of Cancer* 113 (2): (2005): 259-266.
- **544.** K.M. Copeland, A.J. Elliot, R.S. Daniels "Functional chimeras of human immunodeficiency virus type 1 gp120 and influenza A virus (H3) hemagglutinin" *Journal of Virology* 79 (10): (2005): 6459-6471.
- **545.** Daniel P. Webster, Susanna Dunachie, Jenni M. Vuola et al. "Enhanced T cell-mediated protection against malaria in human challenges by using the recombinant poxviruses FP9 and modified vaccinia virus Ankara" *Proceedings of the National Academy of Sciences of the United States of America* 102 (13): (2005): 4836-4841.
- **546.** Jenni M. Vuola, Sheila Keating, Daniel P. Webster et al. "Differential Immunogenicity of Various Heterologous Prime-Boost Vaccine Regimens Using DNA and Viral Vectors in Healthy Volunteers" *Journal of Immunology* 174 (1): (2005): 449-455.
- **576.** E. Prieur, S.C. Gilbert, J. Schneider et al. "A Plasmodium falciparum candidate vaccine based on a six-antigen polyprotein encoded by recombinant poxviruses" *Proceedings of the National Academy of Sciences of the United States of America* 101 (1): (2004): 290-295.
- **548.** Kamel Ait-Tahar, Vincenzo Cerundolo, Alison H. Banham et al. "B and CTL responses to the ALK protein in patients with ALK-positive ALCL" *International Journal of Cancer* 118 (3): (2005): 688-695.
- **549.** Lucy J.C. Smyth, Mariette I.E. van Poelgeest, Emma J. Davidson et al. "Immunological Responses in Women with Human Papillomavirus Type 16(HPV-16)-Associated Anogenital Intraepithelial Neoplasia Induced by Heterologous Prime-Boost HPV-16 Oncogene Vaccination" *Clinical Cancer Research* (2004) 10 (9): (2004): 2954-2961.
- **550.** Richard Harrop, Noel Connolly, Irina Redchenko et al. "Vaccination of Colorectal Cancer Patients with Modified Vaccinia Ankara Delivering the Tumor Antigen 5T4 (TroVax) Induces Immune Responses which Correlate with Disease Control: A Phase I/II Trial" *Clinical Cancer Research* (2004) 12 (1): (2006): 3416-3424.
- **551.** Claire L. Hutchings, Gilbert et al. "Novel Protein and Poxvirus-Based Vaccine Combinations for Simultaneous Induction of Humoral and Cell-Mediated Immunity" *Journal of Immunology* 175 (1): (2005): 599-606.

- **552.** H. McShane, A.A. Pathan, C.R. Sander et al. "Recombinant modified vaccinia virus Ankara expressing antigen 85A boosts BCG-primed and naturally acquired antimycobacterial immunity in humans" *Nature Medicine* 10 (11): (2004): 1240-1244.
- **553.** Victoria Chung, Anthony R. Carroll, Norman M. Gray et al. "Development of cell-based assays for in vitro characterization of hepatitis C virus NS3/4A protease inhibitors" *Antimicrobial Agents & Chemotherapy* 49 (4): (2005): 1381-1390.
- **554.** Caroline L. Smith, Fareed Mirza, Valerie Paquetto et al. "Immunodominance of Poxviral-Specific CTL in a Human Trial of Recombinant-Modified Vaccinia Ankara" *Journal of Immunology* 175 (12): (2005): 8431-8437.
- **555.** Shahriar Behboudi, Anne Moore, Adrian V.S. Hill et al. "Splenic dendritic cell subsets prime and boost CD8 T cells and are involved in the generation of effector CD8 T cells" *Cellular Immunology* 228 (1): (2004): 15-19.
- **556.** Richard H. Clark, Julia C. Kenyon, Nathan W. Bartlett et al. "Deletion of gene A41L enhances vaccinia virus immunogenicity and vaccine efficacy" *Journal of General Virology* 87 (1): (2006): 29-38.
- **557.** Christina A. Bursill, Robin Choudchury, Ziad Ali et al. "Adenoviral delivery of a Cc-chemokine inhibitor, "35k," reduces macrophage recruitment and atherosclerosis in Apoe knockout mice" *Circulation* 110 (Supplement 17): (2004): 252.
- **558.** Christina A. Bursill, Robin Choudhury, Ziad Ali et al. "Broad-Spectrum CC-Chemokine Blockade by Gene Transfer Inhibits Macrophage Recruitment and Atherosclerotic Plaque Formation in Apolipoprotein E-Knockout Mice" *Circulation* 110 (16): (2004): 2460-2466.
- **559.** Christophe Fraser, Steven Riley, Roy M. Anderson et al. "Factors that make an infectious disease outbreak controllable" *Proceedings of the National Academy of Sciences of the United States of America* 101 (16): (2004): 6146-6151.
- **560.** Perla Thorbjornsdottir, Ragnhildur Kolka, Eggert Gunnarsson et al. "Vaccinia virus complement control protein diminishes formation of atherosclerotic lesions. Complement is centrally involved in atherosclerotic disease" *Annals of the New York Academy of Sciences* 1056 (2005): (2005): 1-15.
- **561.** Peter Aaby, Per Gustafson, Adam Roth et al. "Vaccinia scars associated with better survival for adults. An observational study from Guinea-Bissau" *Vaccine* 24 (29-30): (2006): 5718-5725.
- **562.** Okeke Malachy Ifeanyi, Nilssen Oivind, Traavik Terje "Modified vaccinia virus Ankara multiplies in rat IEC-6 cells and limited production of mature virions occurs in other mammalian cell lines" *Journal of General Virology* 87 (1): (2006): 21-27.
- **563.** Anne-Katrine Kvissel, S. Orstavik, S. Eikvar et al. "Co-localization and in vitro phosphorylation of splicing factors by cAMP-dependent protein kinase" *FASEB Journal* 18 (Supplement 8): (2004): C259.
- **564.** Paeivi Turunen, Hanna L. Puhakka, Tommi Heikura et al. "Extracellular Superoxide Dismutase with Vaccinia Virus Anti-inflammatory Protein 35K or Tissue Inhibitor of Metalloproteinase-1: Combination Gene Therapy in the Treatment of Vein Graft Stenosis in Rabbits" *Human Gene Therapy* 17 (4): (2006): 405-414.
- **565.** Leon De Waal, Yasemin Suzer, Linda S. Wyatt et al. "T cell responses to respiratory syncytial virus fusion and attachment proteins in human peripheral blood mononuclear cells" *Viral Immunology* 19 (4): (2006): 669-678.
- **566.** Koert J. Stittelaar, Geert van Amerongen, Ivanela Kondova et al. "Modified vaccinia virus ankara protects macaques against respiratory challenge with monkeypox virus" *Journal of Virology* 79 (12): (2005): 7845-7851.
- **567.** Fernando Moreno-Herrero, Laurent Holtzer, Daniel A. Koster et al. "Atomic force microscopy shows that vaccinia topoisomerase IB generates filaments on DNA in a cooperative fashion" *Nucleic Acids Research* 33 (18): (2005): 5945-5953.
- **568.** Thomas P. Monath, Joseph R. Caldwell, Wolfgang Mundt et al. "ACAM2000 clonal Vero cell culture vaccinia virus (New York City Board of Health strain) a second-generation smallpox vaccine for biological defense" *International Journal of Infectious Diseases* 8 (Supplement 2): (2004): 31-44.

- **569.** Georg W. Holzer, Josef Mayrhofer, Werner Gritschenberger et al. "Dominant negative selection of vaccinia virus using a thymidine kinase/thymidylate kinase fusion gene and the prodrug azidothymidine" *Virology* 337 (2): (2005): 235-241.
- **570.** A. Berting, W. Goerner, M. Spruth et al. "Effective poxvirus removal by sterile filtration during manufacture of plasma derivatives" *Journal of Medical Virology* 75 (4): (05): 603-607.
- **571.** C. Konetschny, G.W. Holzer, C. Urban et al. "Generation of Transduction-Competent Retroviral Vectors by Infection with Single Hybrid Vaccinia Virus" *Journal of Virology* 77 (12): (2003): 7017-7025.
- **572.** Johan Neyts, Pieter Leyssen, Erik Verbeken et al. "Efficacy of cidofovir in a murine model of disseminated progressive vaccinia" *Antimicrobial Agents & Chemotherapy* 48 (6): (2004): 2267-2273.
- **573.** Erik De Clercq "Cidofovir in the therapy and short-term prophylaxis of poxvirus infections" *Trends in Pharmacological Sciences* 23 (10): (2002): 456-458.
- **574.** Erik De Clercq, G. Andrei, J. Balzarini et al. "Antiviral potential of a new generation of acyclic nucleoside phosphonates, the 6-[2-(phosphonomethoxy)alkoxy]-2,4-diaminopyrimidines" *Nucleosides, Nucleotides & Nucleic Acids* 24 (5-7): (2005): 331-341.
- 575. G. Andrei, J. Van den Oord, K.Y. Hostetler et al. "Activity of alkoxyalkyl and alkyl esters of (S)-3-hydroxy-2-phosphonylmethoxypropyl derivatives of cytosine (HPMPC, cidofovir) and adenine (HPMPA) and cyclic cidofovir against orthopoxyiruses" *Antiviral Research* 65 (3): (2005): A92.
- 576. P.P. Pastoret, A. Vanderplasschen "The uses of poxviruses as vectors" Current Gene Therapy 3 (6): (2003): 583-595
- **577.** Nathan W. Bartlett, Laure Dumoutier, Jean-Christophe Renauld et al. "A new member of the interleukin 10-related cytokine family encoded by a poxvirus" *Journal of General Virology* 85 (6): (2004): 1401-1412.
- **578.** N. Antoine-Moussiaux, B. Faye, G.F. Vias et al. "Tuareg ethnodiagnostic skill of camel diseases in Agadez area (Niger)" *Journal of Camel Practice and Research* 12 (2): (2005): 85-93.
- **579.** M.H. Al-Gaabary, A.A.A. Rawash "Clinical, epidemiological and histopathological aspects of an outbreak of buffalo pox" *Assiut Veterinary Medical Journal* 50 (102): (2004): 157-171.
- **580.** Mohamed Ragaa Mohamed, Edward G. Niles "Transient and inducible expression of vaccinia/T7 recombinant viruses" *Methods in Molecular Biology* 269: (2004): 41-50.
- **581.** E.M.E. Abu Elzein, F.M.T. Housawi, A.I. Al-Afaleq et al. "The Ability of Sheeppox Vaccine to Protect Goats Against a Virulent Field Capripox virus with High Affinity to Goats" *The Journal of King Faisal University (Basic and Applied Sciences)* 2: (2004): 274.
- **582.** Jacqueline Weyer, Charles E. Rupprecht, Janet Mans et al. "Generation and evaluation of a recombinant modified vaccinia virus Ankara vaccine for rabies" *Vaccine* 25 (21): (2007): 4213-4222.
- **583.** Yohannes T. Ghebremariam, Scott A. Smith, J.B. Anderson et al. "Intervention strategies and agents mediating the prevention of xenorejection" *Annals of the New York Academy of Sciences* 1056: (2005): 123-143.
- **584.** Nirvana S. Pillay, Laurie A. Kellaway, Girish J. Kotwal "Administration of vaccinia virus complement control protein shows significant cognitive improvement in a mild injury model" *Annals of the New York Academy of Sciences* 1056: (2005): 450-461.
- **585.** Amod P. Kulkarni, Laurie A. Kellaway, K. Debomoy et al. "Neuroprotection from complement-mediated inflammatory damage" *Annals of the New York Academy of Sciences* 1035: (2004): 147-164.
- **586.** Yohannes T. Ghebremariam, Odutayo O. Odunuga, Kristen Janse et al. "Humanized recombinant vaccinia virus complement control protein (hrVCP) with three amino acid changes, H98Y, E102K, and E120K creating an additional putative heparin binding site, is 100-fold more active than rVCP in blocking both classical and alterantive complement pathways" *Annals of the New York Academy of Sciences* 1056: (2005): 113-122.

- **587.** Melissa-Rose Abrahams, Zhouning Zhang, Sufan Chien et al. "The vaccinia virus N1L ORF may encode a multifunctional protein possibly targeting different kinases, one of which influences ATP levels in vivo" *Annals of the New York Academy of Sciences* 1056: (2005): 87-99.
- **588.** Hannah B. Ibanga, Roger H. Brookes, Philip C. Hill et al. "Early clinical trials with a new tuberculosis vaccine, MVA85A, in tuberculosis-endemic countries: Issues in study design" *Lancet Infectious Diseases* 6 (8): (2006): 522-528.
- **589.** E.B. Imoukhuede, T. Berthoud, P. Milligan et al. "Safety and immunogenicity of the malaria candidate vaccines FP9 CS and MVA CS in adult Gambian men" *Vaccine* 24 (42-43): (2006): 6526-6533.
- **590.** Vasee S. Moorthy, Egeruan B. Imoukhuede, Sheila Keating et al. "Phase 1 evaluation of 3 highly immunogenic prime-boost regimens, including a 12-month reboosting vaccination, for malaria vaccination in Gambian men" *Journal of Infectious Diseases* 189 (12): (2004): 2213-2219.
- **591.** Melaku Tefera, Fesha Gebreah "A Study on the Productivity and Diseases of Camels in Eastern Ethiopia" *Tropical Animal Health and Production* 33 (4): (2001): 265-274.
- **592.** Vazquez-Blomquist Dania, Quintana Diogenes, Duarte Carlos A "Modified-vaccinia-virus-Ankara (MVA) priming and fowlpox-virus booster elicit a stronger CD8+ T-cell response in mice against an HIV-1 epitope than does a DNA/poxvirus prime-booster approach" *Biotechnology and Applied Biochemistry* 39 (3): (2004): 313-318.
- **593.** Carlos Franco-Paredes, Lorena Lammoglia, Jose Ignacio Santos-Preciado "Historical perspective of smallpox in Mexico: emergence, elimination, and risk of reemergence due to bioterrorism" *Mexican Medical Gazette* 140 (3): (2004): 321-327.
- **594.** A.A. Carvajal, A. de la Garza, B.J. Quiroz et al. "MVA E2 recombinant vaccine in the treatment of human papillomavirus infection in men presenting intraurethral flat condyloma A phase I/II study" *BioDrugs* 21 (1): (2007): 47-59.
- **595.** E. Garcia-Hernandez, J.L. Gonzalez-Sanchez, A. Andrade-Manzano et al. "Regression of papilloma high-grade lesions (CIN 2 and CIN 3) is stimulated by therapeutic vaccination with MVA E2 recombinant vaccine" *Cancer Gene Therapy* 13 (6): (2006): 592-597.
- **596.** Z.I.P. Lobato, G.S. Trindade, M.C.M. Frois et al. "Outbreak of exanthema disease caused by Vaccinia virus in human and cattle in Zona da Mata region, Minas Gerais" *Brazilian Archives of Veterinary Medicine and Animal Husbandry* 57 (4): (2005): 423-429.
- **597.** J.A. Santos, V.C. Melo, N. Moussatche et al. "Occurrence of Orthopoxvirus in Dairy Cows in Farms of the Municipalities of Campos dos Goytacazes and Sao Fidelis, Rio de Janeiro State" *Brazilian Congress of Microbiology* 22: (2003).
- **598.** Bauer Airosa Cabrera, Nissin Moussatche "Induction of Apoptosis in Cells Infected by Cantagalo Virus: Possible Involvement of the Serine Protease 2 Inhibitor Gene" *CAPES Thesis Database* (2004).
- **599.** Sheila Albert Dos Reis, Clarissa Rosa de Almeida Damaso "Cantagalo Virus and Vaccine Strain VV-IOC: Characterization of the B18R Gene and of the Antiviral Effect of Type 1 Interferon" *CAPES Thesis Database* (2004).
- **600.** Teresa Keico Nagasse-Sugahara, Jonas Jose Kisielius, Marli Ueda-Ito et al. "Human vaccinia-like virus outbreaks in Sao Paulo and Goias States, Brazil: virus detection, isolation and identification" *Journal of the Sao Paulo Institute for Tropical Medicine* 46 (6): (2004): 315-322.
- **601.** Giliane S. Trindade, Flavio G. da Fonseca, Joao T. Marques et al. "Belo Horizonte virus: a vaccinia-like virus lacking the A-type inclusion body gene isolated from infected mice" *Journal of General Virology* 85 (7): (2004): 2015-2021.
- **602.** Flavio G. da Fonseca, Giliane S. Trindade, Ricardo L.A. Silva et al. "Characterization of a vaccinia-like virus isolated in a Brazilian forest" *Journal of Virology* 83: (2002): 223-228.
- **603.** J.M.S. Ferreira, F.M. Oliveira, B.E.F. Mota et al. "Study of the Pathogenesis of Vaccinia Virus Samples in BALB/c Mice" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).

- **604.** Giliane de Souza Trindade, Erna Geessien Kroon, Flavio Guimaraes da Fonseca "Isolation, Identification, and Molecular Characterization of Poxviruses Circulating in Brazil" *CAPES Thesis Database* (2003).
- **605.** G.S. Trindade, J.M.C. Vilela, J.M.S. Ferreira et al. "Use of atomic force microscopy as a diagnostic tool to identify orthopoxvirus" *Journal of Virological Methods* 141 (2): (2007): 198-204.
- **606.** Clarissa R.A. Damaso, Sheila A. Reis, Desyree M. Jesus et al. "A PCR-based assay for detection of emerging vaccinia-like viruses isolated in Brazil" *Diagnostic Microbiology and Infectious Disease* 57 (1): (2007): 39-46.
- **607.** J.S. Abrahao, J.M.S. Ferreira, F.M. Oliveira et al. "Vaccinia Virus Detection In Clinical Specimens Utilizing Polymerase Chain Reaction" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).
- **608.** Frederico Alberto Botelho Greco, Nissin Moussatche "Analysis of the Essentiality of the F11L Gene of Vaccinia Virus" *CAPES Thesis Database* (2003).
- **609.** Anna Carolina Toledo da Cunha Pereira, Claudio Antonio Bonjardim "Analysis of the Activation of MAPKs During Infection by Vaccinia Virus: Involvement of ERK, JNK, and P38" *CAPES Thesis Database* (2003).
- **610.** Jamaria Adriana Pinheiro Soares, Claudio Antonio Bonjardim "Cellular Signalling Pathways That Contribute to the Multiplication of Cowpox Virus" *CAPES Thesis Database* (2004).
- **611.** Marcelo Henrique Aguiar de Freitas, Claudio Antonio Bonjardim "Induction of the Gene of Early Response to Growth (Egr-1) After Cowpox Virus Infection in A31 Cells" *CAPES Thesis Database* (2004).
- **612.** Anderson Assuncao Andrade, Claudio Antonio Bonjardim "The Role of Mitogen-Activated Protein Kinases (MAPK) MEK/ERK and of Their Effectors RSK2, ELK-1, and EGR-1, in Multiplication of Vaccinia Virus" *CAPES Thesis Database* (2004).
- **613.** Sayuri E.M. Kato, Frederico A.B. Greco, Clarissa R.A. Damaso et al. "An alternative genetic method to test essential vaccinia virus early genes" *Journal of Virological Methods* 115 (1): (2004): 31-40.
- **614.** A.A. Andrade, P.N.G. Silva, A.C.T.C. Pereira et al. "The vaccinia virus-stimulated mitogen-activated protein kinase (MAPK) pathway is required for virus multiplication" *Biochemical Journal* 381 (2): (2004): 437.
- **615.** N.G. Silva Patricia, A.P. Soares Jamaria, S.A.F. Brasil Bruno et al. "Differential role played by the MEK/ERK/EGR-1 pathway in orthopoxviruses vaccinia and cowpox biology" *Biochemical Journal* 398 (1): (2006): 83-95.
- **616** B.S.A.F. Brasil, L.C. Oliveira, E.G. Kroon et al. "The Cellular Gene EGR-1 Is Regulated During Vaccinia Virus Infection and Facilitates its Replication" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).
- **617.** A.P.V. Castro, T.M.U. Carvalho, N. Moussatche et al. "Redistribution of Cyclophilin A to Viral Factories during Vaccinia Virus Infection and Its Incorporation into Mature Particles" *Journal of Virology* 77 (16): (2003): 9052-9068.
- **618.** Juliana de Almeida Leite, Erna Geessien Kroon, Giliane de Souza Trindade "Isolation, Identification, and Molecular Characterization of Orthopoxviruses Isolated in an Outbreak of Bovine Variola in Passatempo, Minas Gerais" *CAPES Thesis Database* (2003).
- **619.** Juliana Almeida Leite, Giliane de Souza Trindade, Claudio Antonio Bonjardim et al. "Brazilian Vaccinia virus strains show genetic polymorphism at the ati gene" *Virus Genes* 36: (2007).
- **620.** F.G.G. Leite, A.C.T.C. Pereira, P.C.P. Ferreira et al. "Contribution Of The C-Jun Transcription Factor to Vaccinia virus (VV) Replication" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).
- **621.** J.A.P. Soares, A.C.T.C. Pereira, E.G. Kroon et al. "JNK Pathway Induction By The Cowpox Virus Is Critical For Viral Biology" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).

- **622.** L.C. Oliveira, B.S.A.F. Brasil, P.C.P. Ferreira et al. "Comparative Analysis Of The Role Of EGR-1 In The Replication Cycle Of Orthopoxvirus And Cowpox" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).
- **623.** D.B. Oliveira, G. Magno de Freitas Almeida, C.A. Bonjardim et al. "Production of Recombinant Human Interferon Lambda1 and Sensitivity of Herpes Simplex Virus 1, Apeu Virus, Encephalomicarditis Virus and Vaccinia Virus" *III Forum on Microbiology at the Federal University of Minas Gerais, Graduate Program in Microbiology* (2007).
- **624.** S.A. Reis, N. Moussatche, C.R.A. Damaso "FK506, a secondary metabolite produced by Streptomyces, presents a novel antiviral activity against Orthopoxvirus infection in cell culture" *Journal of Applied Microbiology* 100 (6): (2006): 1373-1380.
- **625.** Adrian Lewis, Sergio Miguel, Javier Defillippo et al. "Residual Humoral Immunity From the Anti-Variola Vaccination" *Argentine Journal of Microbiology* 37 (Supplement 1): (2005): 79.
- **626.** A. Lewis, K. Bok, O. Perez et al. "Characterization of a vaccinia virus strain used to produce smallpox vaccine in Argentina between 1937 and 1970" *Archives of Virology* 150 (7): (2005): 1485-1491.
- **627.** M. Fraser, S. Hatt, D. Smith "Assessment of Two Potency Tests for Smallpox Vaccine Using a Study Panel Obtained from the National Institute for Biological Standards and Control (NIBSC)" *Health Canada Science Forum* 5: (2005).
- **628.** David Joseph Brick "Molecular characterization of poxviral RING finger proteins: virosome localization and identification of DNA binding and apoptosis inhibition activity" *Theses Canada Database* (2004).
- **629.** John-Paul Bacik, Angela M. Brigley, Lisa D. Channon et al. "Purification, crystallization and preliminary diffraction studies of an ectromelia virus glutaredoxin" *Acta Crystallographica* 61 (6): (2005): 550.
- **630.** John-Paul Bacik, Bart Hazes "Crystal structures of a poxviral glutaredoxin in the oxidized and reduced states show redox-correlated structural changes" *Journal of Molecular Biology* 365 (5): (2007): 1545-1558.
- **631.** Alicja Zachertowska, Dyanne Brewer, David H. Evans "Characterization of the major capsid proteins of myxoma virus particles using MALDI-TOF mass spectrometry" *Journal of Virological Methods* 132 (1-2): (2006): 1-12.
- **632.** Hana Weingartl, Markus Czub, Stefanie Czub et al. "Immunization with modified vaccinia virus Ankara-based recombinant vaccine against severe acute respiratory syndrome is associated with enhanced hepatitis in ferrets" *Journal of Virology* 78 (22): (2004): 12672-12676.
- **633.** Timothy Z. Vitalis, Qianjin Zhang, Judies Alimonti et al. "Using the TAP component of the antigen-processing machinery as a molecular adjuvant" *PLoS Pathogens* 1 (4): (2005): 289-298.
- **634.** J. Andrea McCart, Navneet Mehta, Deborah Scollard et al. "Oncolytic Vaccinia Virus Expressing the Human Somatostatin Receptor SSTR2: Molecular Imaging after Systemic Delivery Using 111In-Pentetreotide" *Molecular Therapy (2004)* 10 (3): (2004): 553-561.
- **635.** Qianjin Zhang, Robyn Seipp, Susan S. Chen et al. "TAP expression reduces IL-10 expressing tumor infiltrating lymphocytes and restores immunosurveillance against melanoma" *International Journal of Cancer* 120 (9): (2007): 1935-1941.
- **636.** Janina Q. Jiang, Amy Patrick, Ronald B. Moss et al. "CD8+ T-cell-mediated cross-clade protection in the genital trace following intranasal immunization with inactivated human immunodeficiency virus antigen plus CpG oligodeoxynucleotides" *Journal of Virology* 79 (1): (2005): 393-400.
- **637.** Mian Gao, Nicole Brufatto, Tricia Chen et al. "Expression Profiling of Herpesvirus and Vaccinia Virus Proteins Using a High-Throughput Baculovirus Screening System" *Journal of Proteome Research* 4 (6): (2005): 2225-2235.
- **638.** Sheryl Kirwan, Deborah Merriam, Nicola Barsby et al. "Vaccinia virus modulation of natural killer cell function by direct infection" *Virology* 347 (1): (2006): 75-87.
- **639.** Shawn T. Wasilenko, Logan Banadyga, David Bond et al. "The vaccinia virus F1L protein interacts with the proapoptotic protein Bak and inhibits Bak activation" *Journal of Virology* 79 (22): (2005): 14031-14043.

- **640.** Tara L. Stewart, Shawn T. Wasilenko, Michele Barry "Vaccinia virus F1L protein is a tail-anchored protein that functions at the mitochondria to inhibit apoptosis" *Journal of Virology* 79 (2): (2005): 1084-1098.
- **641.** Melissa Da Silva, Chris Upton "Using purine skews to predict genes in AT-rich poxviruses" *BMC Genomics* 6 (1): (2005): 22.
- **642.** Melissa Da Silva, Ling Shen, Vasily U. Tcherepanov et al. "Predicted function of the vaccinia virus G5R protein" *Bioinformatics* 22 (23): (2006): 2846-2850.
- **643.** Guiyun Li, Nanhai Chen, Zehua Feng et al. "Genomic sequence and analysis of a vaccinia virus isolate from a patient with a smallpox vaccine-related complication" *Virology Journal* 3 (1): (2006): 88.
- **644.** Anne Gatignol, Sebastien Laine, Guerline Clerzius "Dual role of TRBP in HIV replication and RNA interference: viral diversion of a cellular pathway or evasion from antiviral immunity?" *Retrovirology* 2 (1): (2005): 65.
- **645.** Michael D. Hamilton, David H. Evans "Enzymatic processing of replication and recombination intermediates by the vaccinia virus DNA polymerase" *Nucleic Acids Research* 33 (7): (2005): 2259-2268.
- **646.** Julie A. Yager, Lisa Hutchison, John W. Barrett "Raccoonpox in a Canadian cat" *Veterinary Dermatology* 17 (6): (2006): 443-448.
- **647.** Wendy C. Magee, Karl Y. Hostetler, David H. Evans "Mechanism of inhibition of vaccinia virus DNA polymerase by cidofovir diphosphate" *Antimicrobial Agents & Chemotherapy* 49 (8): (2005): 3153-3162.
- **648.** Isabelle Bougie, Martin Bisaillon "The Broad Spectrum Antiviral Nucleoside Ribavirin as a Substrate for aViral RNA Capping Enzyme" *Journal of Biological Chemistry* 279 (21): (2004): 22124-22130.
- **649.** Jodie M. Harrison, Ian A. Ramshaw "Cytokines, skin, and smallpox-a new link to an antimicrobial peptide" *Immunity* 24 (3): (2006): 245-247.
- **650.** Zhaochun Chen, Patricia Earl, Jeffrey Americo et al. "Chimpanzee/human mAbs to vaccinia virus B5 protein neutralize vaccinia and smallpox viruses and protect mice against vaccinia virus" *Proceedings of the National Academy of Sciences of the United States of America* 103 (6): (2006): 1882-1887.
- **651.** Zhaochun Chen, Patricia Earl, Jeffrey Americo et al. "Characterization of Chimpanzee/Human Monoclonal Antibodies to Vaccinia Virus A33 Glycoprotein and Its Variola Virus Homolog In Vitro and in a Vaccinia Virus Mouse Protection Model" *Journal of* Virology 81 (17): (2007): 8989-8995.
- **652.** John O. Agwunobi "Should the United States and Russia destroy their stocks of smallpox virus?" *British Medical Journal* 334 (7597): (2007): 774-775.